



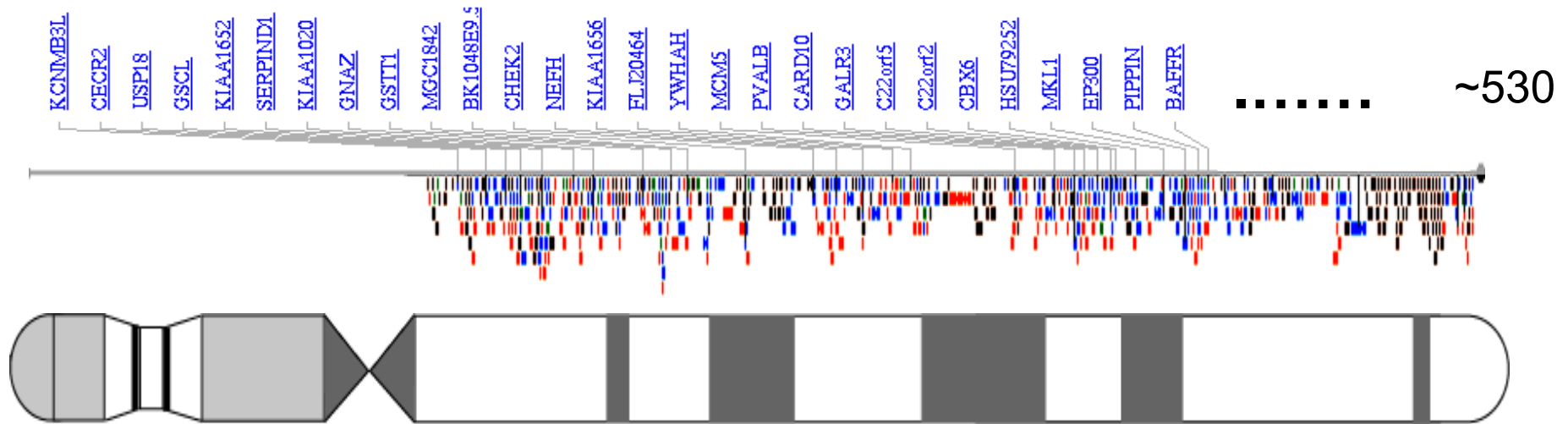
Understanding Protein Function on a Genome-scale through the Analysis of Molecular Networks

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Yale

slides at
Lectures.GersteinLab.org

(See Last Slide for References
& More Info.)

The problem: Grappling with Function on a Genome Scale?



- 250 of ~530
originally characterized on chr. 22
[Dunham et al. Nature (1999)]
- >25K Proteins in Entire Human Genome
(with alt. splicing)

Traditional single molecule way to integrate evidence & describe function

EF2_YEAST

Descriptive Name:
Elongation Factor 2

Lots of references
to papers

Summary sentence describing function:
This protein promotes the GTP-dependent translocation of the nascent protein chain from the A-site to the P-site of the ribosome.

UniProt

Basic UniProt Protein Viewer - UniProt [the Universal Protein Resource] - Microsoft Internet Explorer

Home > Database > UniProt Protein Viewer

Text Search UniProt Knowledgebase

Home About UniProt Getting Started Searches/Tools Databases Support/Documentation

General information about the UniProt/Swiss-Prot entry	
Entry name	EF2_YEAST
Primary accession number	P32324
Entered in Swiss-Prot	Release 27, 01-OCT-1993
Sequence was last modified	Release 27, 01-OCT-1993
Annotations were last modified	Release 47, 01-MAY-2005

Protein description	
Protein name	Elongation factor 2
Synonyms	EF-2

References	
[1]	NUCLEOTIDE SEQUENCE (EFT1 AND EFT2). MEDLINE=92112760; PubMed=1730643; [NCBI, ExPASy, EBI, Israel, Japan] Perentesis J.P., Phan L.D., Laporte D.C., Livingston D.M., Bodley J.W.; "Saccharomyces cerevisiae elongation factor 2. Genetic cloning, characterization of expression, and G-domain modeling."

Comments	
FUNCTION	This protein promotes the GTP-dependent translocation of the nascent protein chain from the A-site to the P-site of the ribosome.
SUBCELLULAR LOCATION	Cytoplasmic.

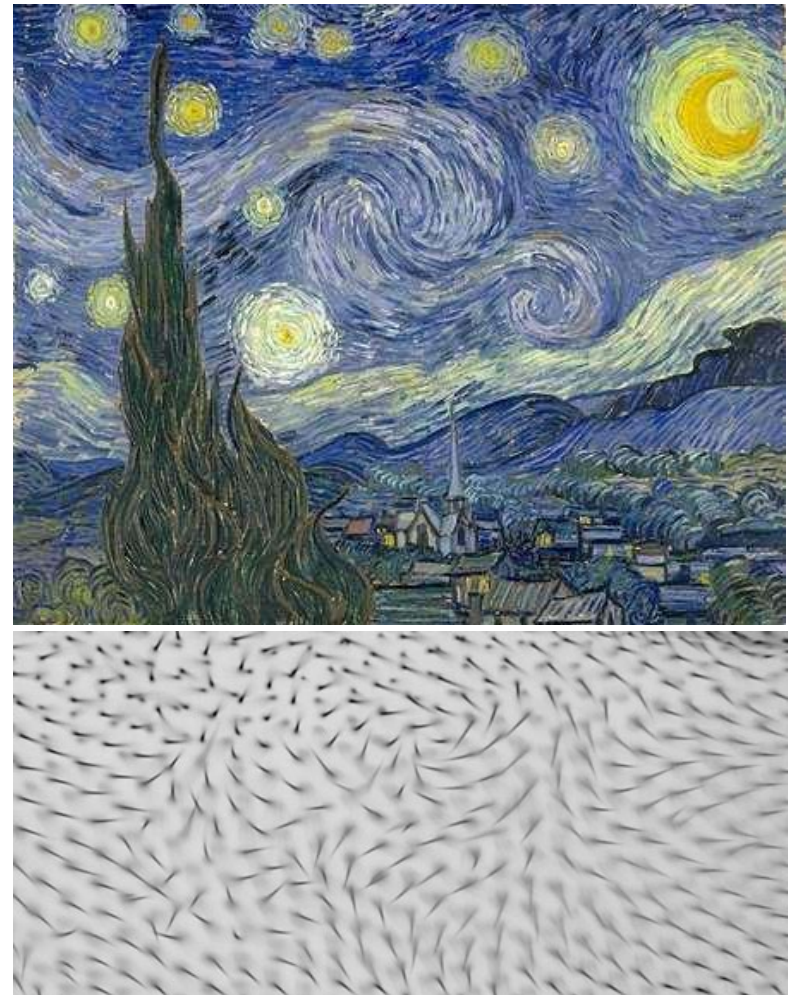
DIR Δ41778-Δ41778

Some obvious issues in scaling single molecule definition to a genomic scale

- Fundamental complexities
 - ◇ Often >2 proteins/function
 - ◇ Multi-functionality:
2 functions/protein
 - ◇ Role Conflation:
molecular, cellular, phenotypic

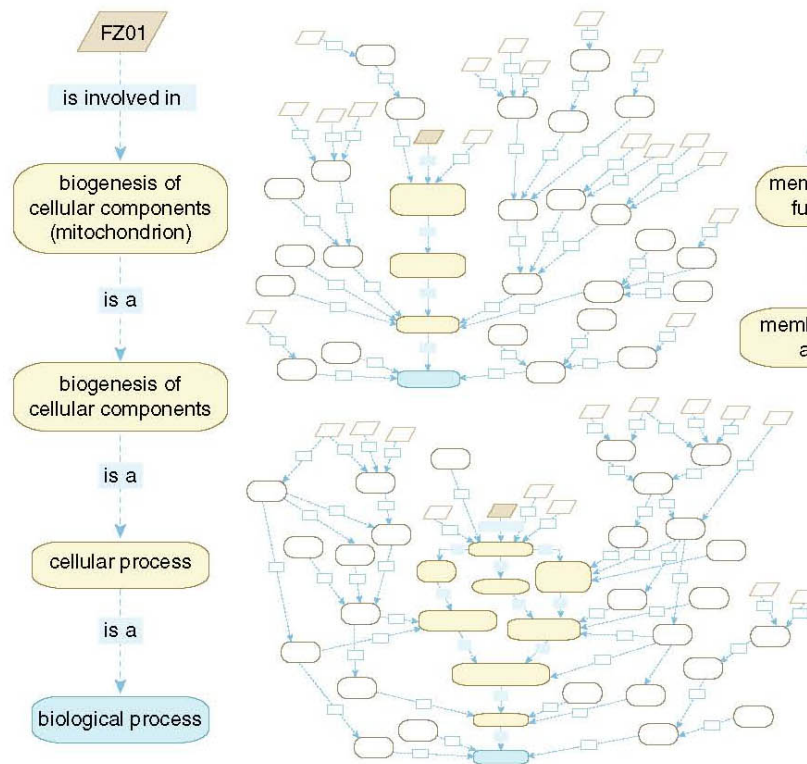
Some obvious issues in scaling single molecule definition to a genomic scale

- Fundamental complexities
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 - ◇ Multi-functionality:
2 functions/protein
 - ◇ Role Conflation:
molecular, cellular, phenotypic
- Fun terms... but do they scale?....
 - ◇ **Starry night** (P Adler, '94)

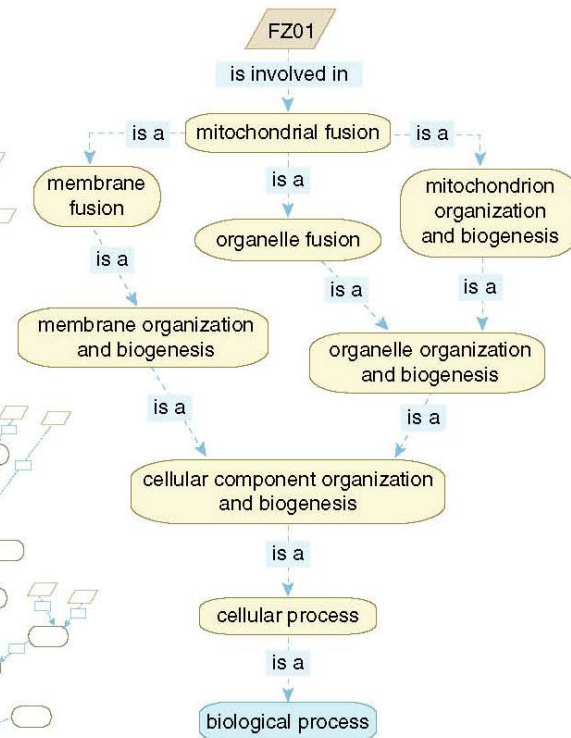


[Seringhaus et al. GenomeBiology (2008)]

Hierarchies & DAGs of controlled-vocab terms but still have issues...



MIPS (Mewes et al.)



GO (Ashburner et al.)

Towards Developing Standardized Descriptions of Function

- Subjecting each gene to standardized expt. and cataloging effect
 - ◊ KOs of each gene in a variety of std. conditions => phenotypes
 - ◊ Std. binding expts for each gene (e.g. prot. chip)
- Function as a vector

ector

nucleic acids

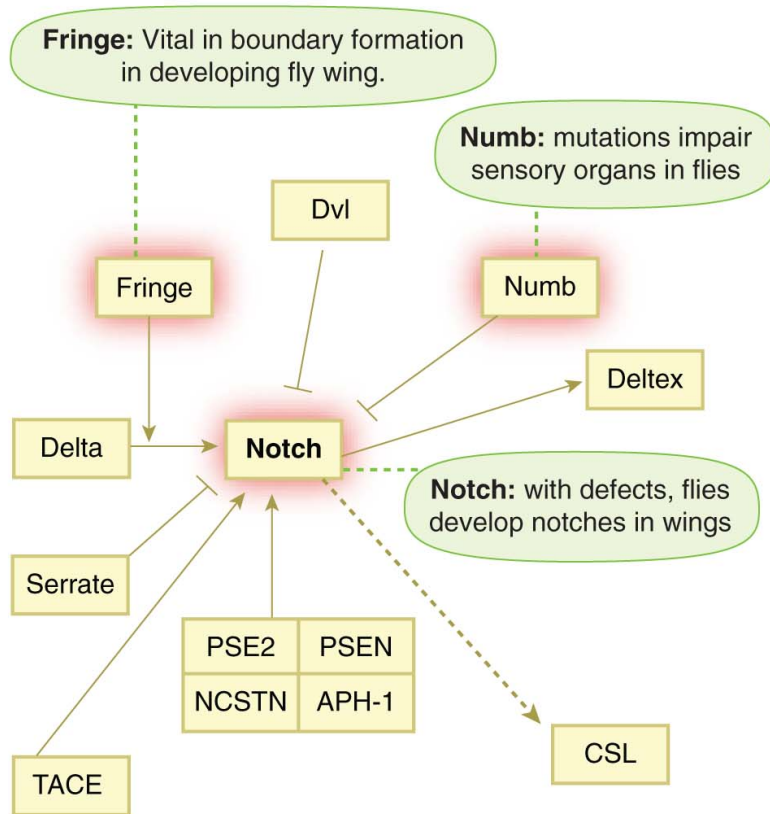
small molecules

proteins

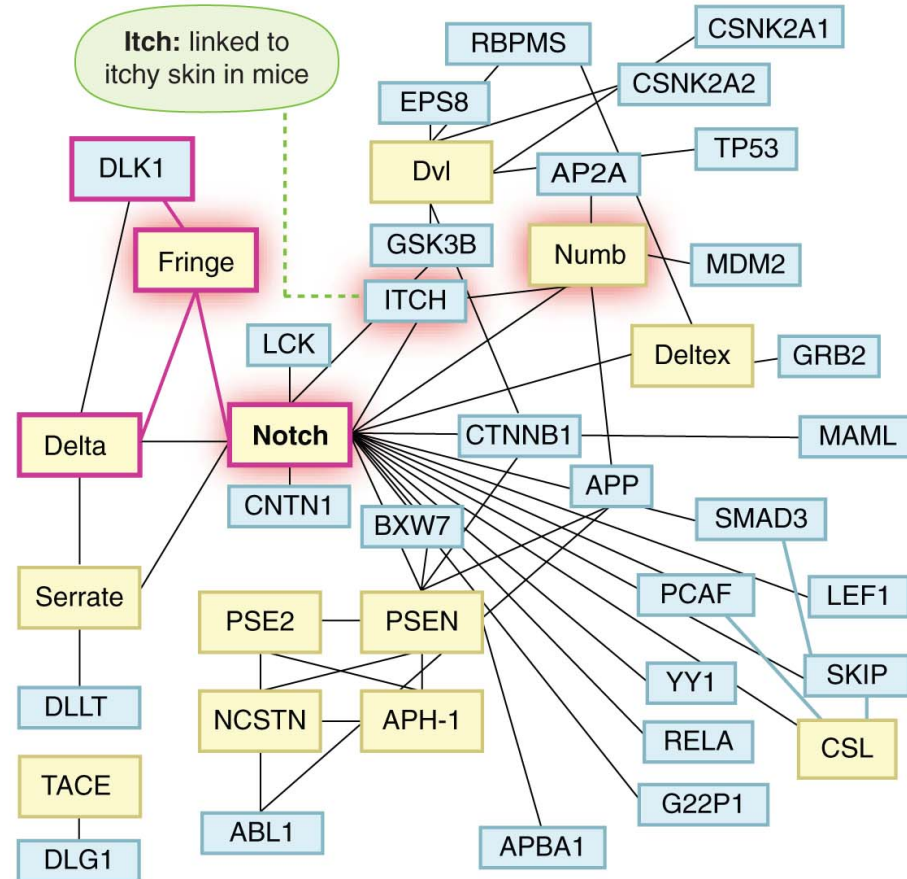
	DNA	RNA	ATP	Metal	CoA	NAD	G protein	CDC28	Calmodulin
protein 1	1.0	0	0	0	0	0	0	0	0
protein 2	0	0.9	0	0	0	0	0	0	0
protein 3	1.0	0	1.0	0	0	0	0	0	0
protein 4	0	0	0	0	0.8	0	0	0	1.0
protein 5	1.0	0	0	0	0	0	0	0.9	0
protein 6	0.9	0				
protein 7	0	0.8				
.....

Interaction Vectors [Lan et al, IEEE 90:1848]

Networks (Old & New)

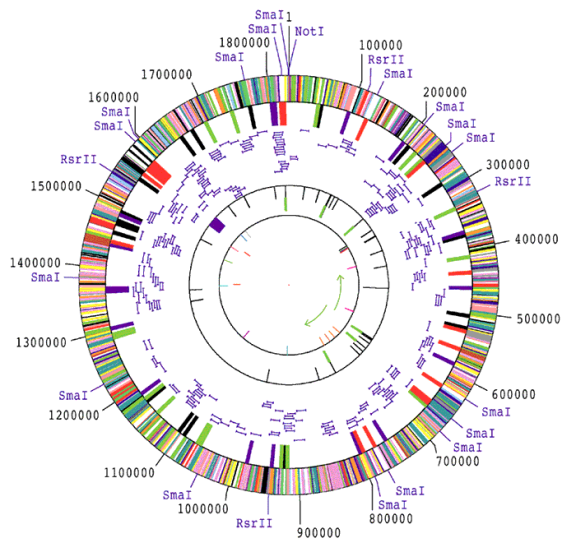


Classical KEGG pathway



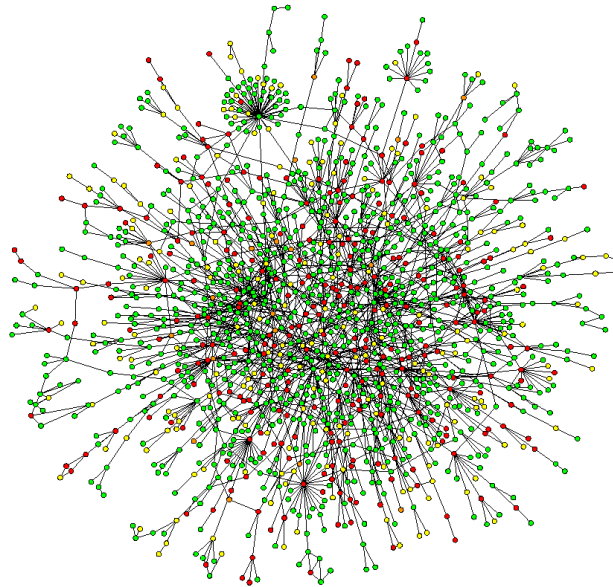
Same Genes in High-throughput Network

Networks occupy a midway point in terms of level of understanding



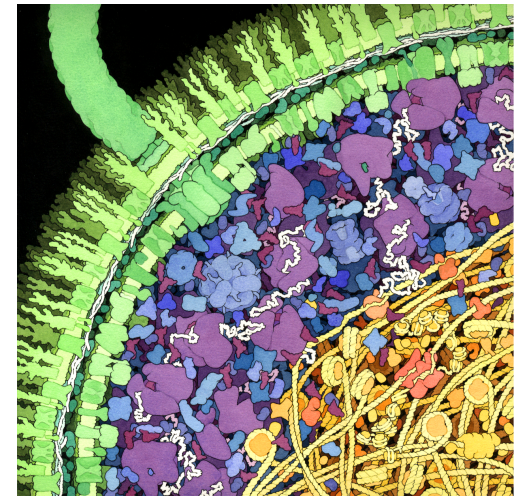
1D: Complete
Genetic Partslist

[Fleischmann et al., Science, 269 :496]



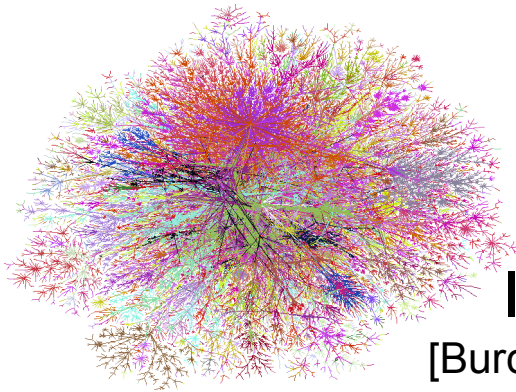
~2D: Bio-molecular
Network
Wiring Diagram

[Jeong et al. Nature, 41:411]

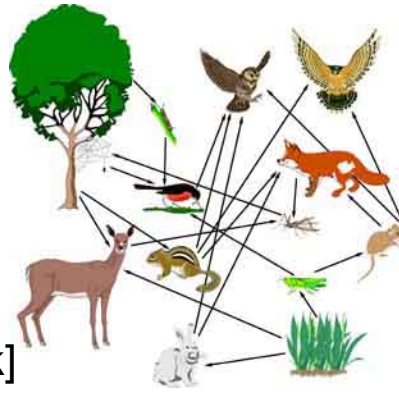


3D: Detailed
structural
understanding of
cellular machinery

Networks as a universal language



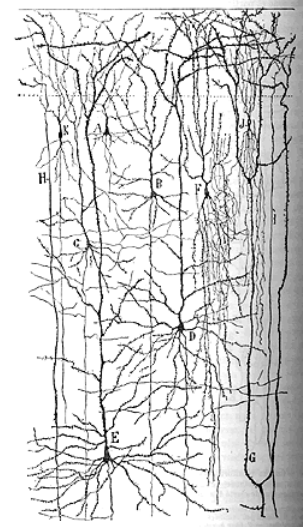
Internet
[Burch & Cheswick]



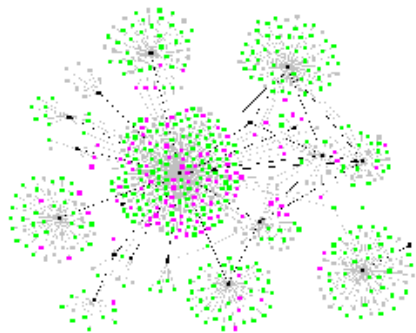
Food Web



Electronic
Circuit



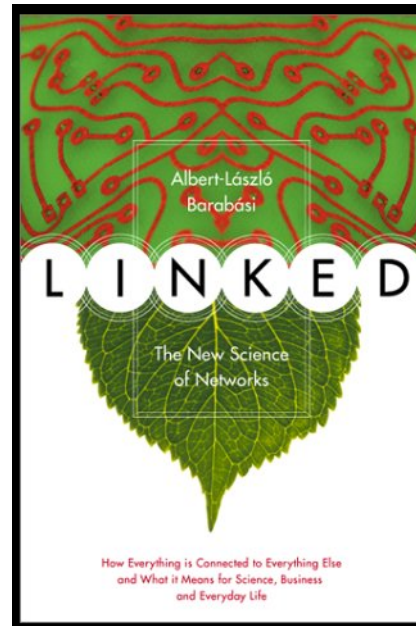
Neural Network
[Cajal]



Disease
Spread
[Krebs]



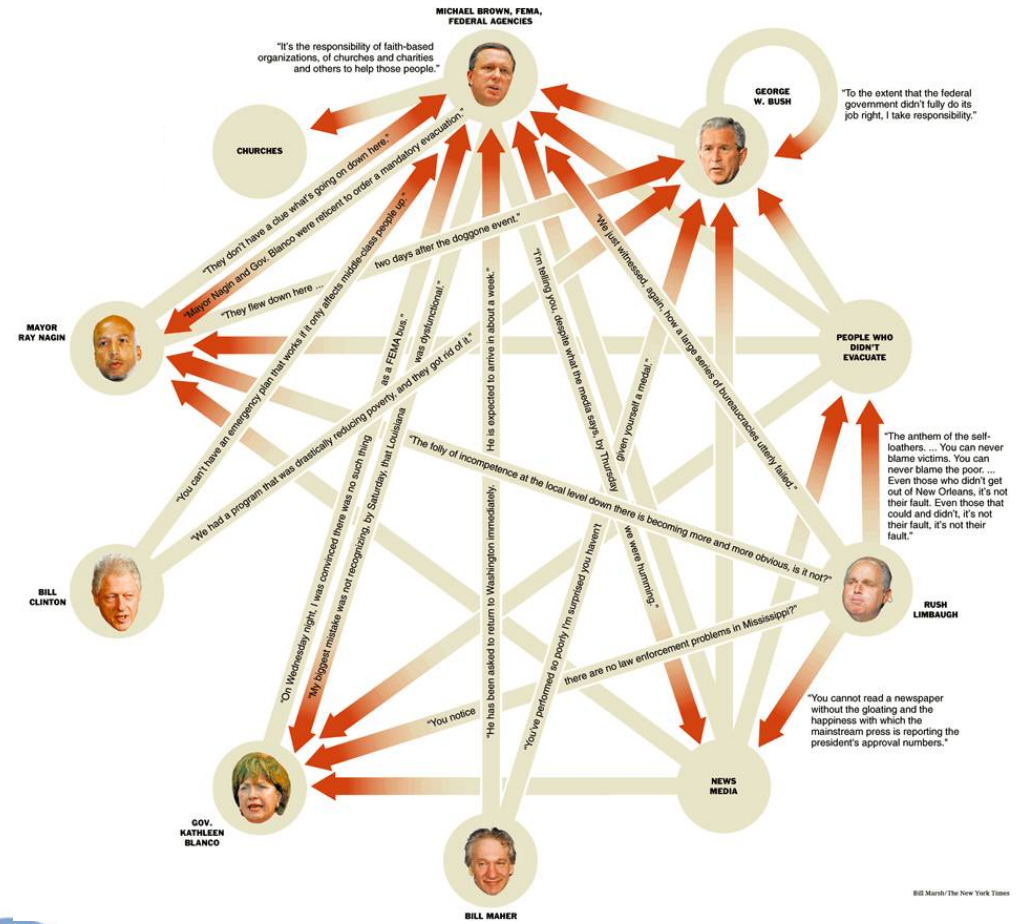
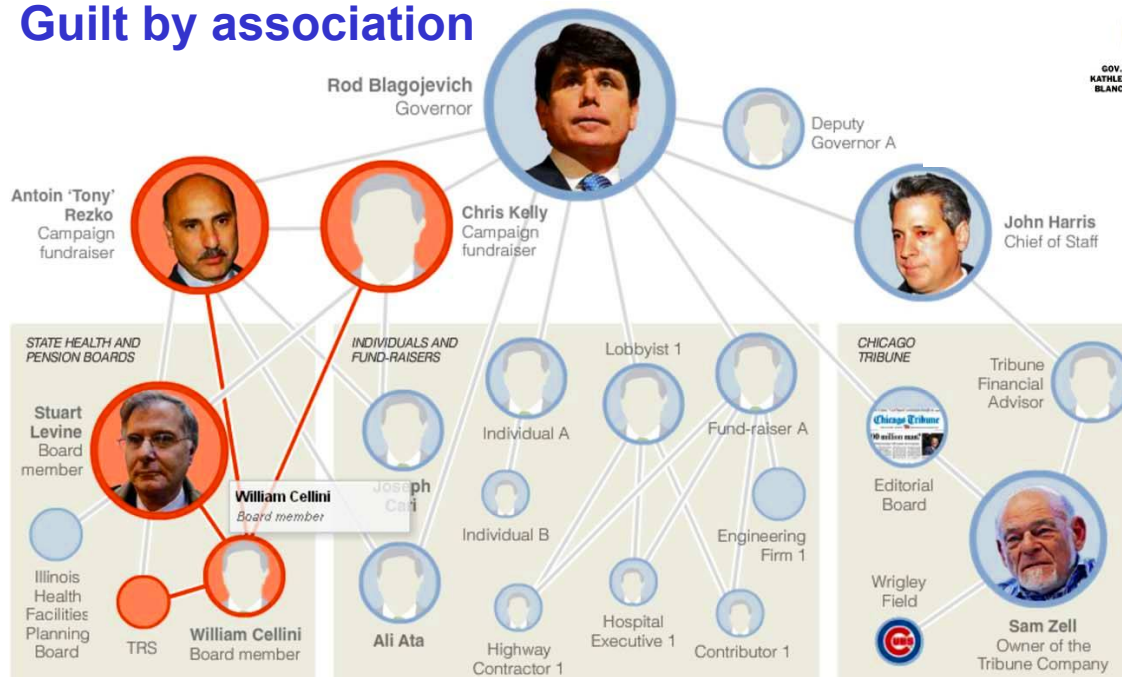
Protein
Interactions
[Barabasi]



Social Network

Using the position in networks to describe function

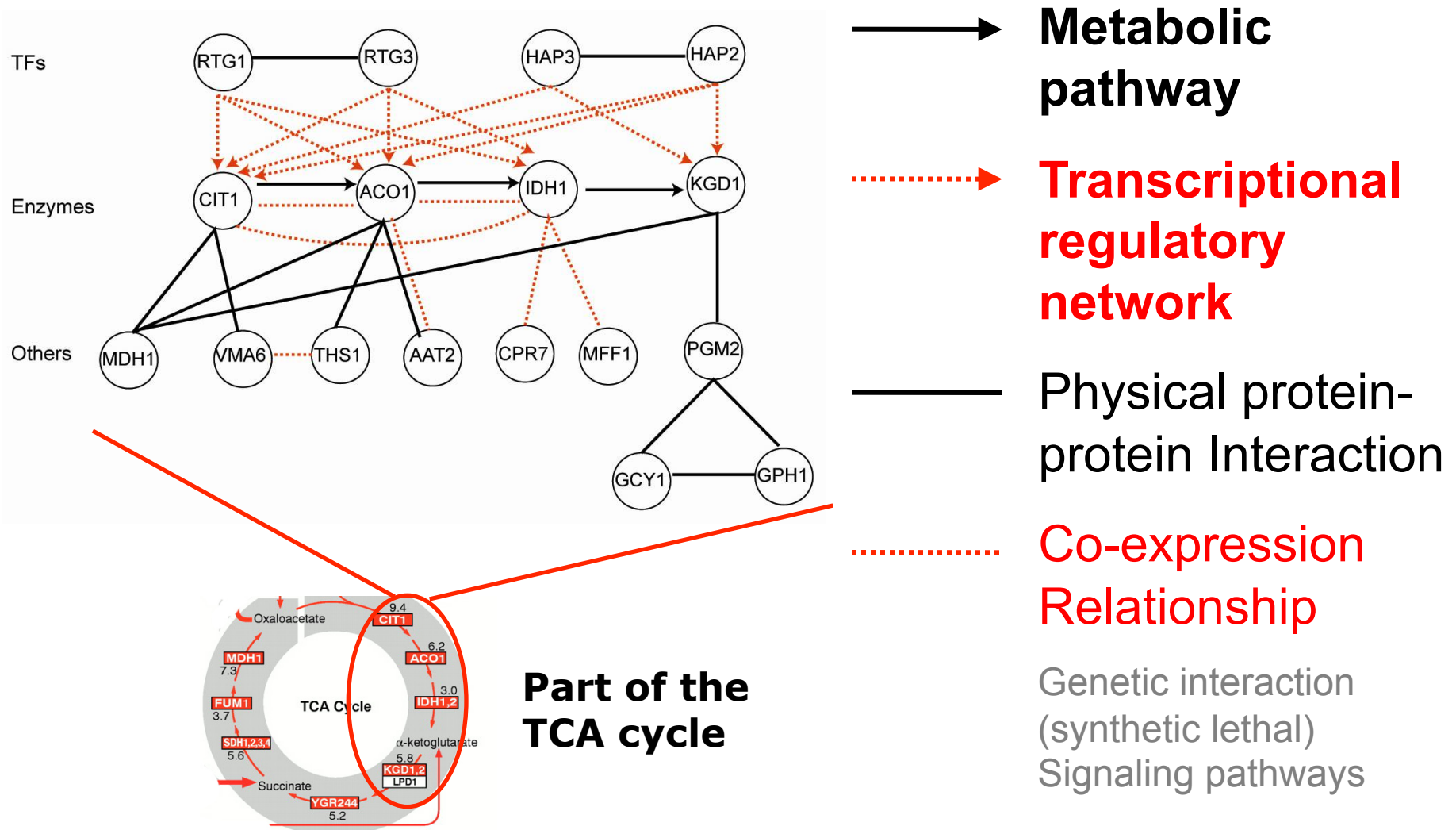
Guilt by association



Finding the causal regulator (the "Blame Game")

[NY Times, 2-Oct-05, 9-Dec-08]

Combining networks forms an ideal way of integrating diverse information



Outline: Molecular Networks

- Why Networks?
- Predicting Networks (yeast ppi)
 - ◇ Propagating known information
- Central Points in Networks
 - ◇ Hubs & Bottlenecks (yeast ppi & reg. net)
 - ◇ Tops of Hierarchies (yeast reg. net)
 - ◇ Identified by score (human miRNA-targ. net)
- Dynamics of Networks
 - ◇ Across environments (in prokaryote metab. pathways)
- Protein Networks & Variation (human ppi & miRNA-targ. net)



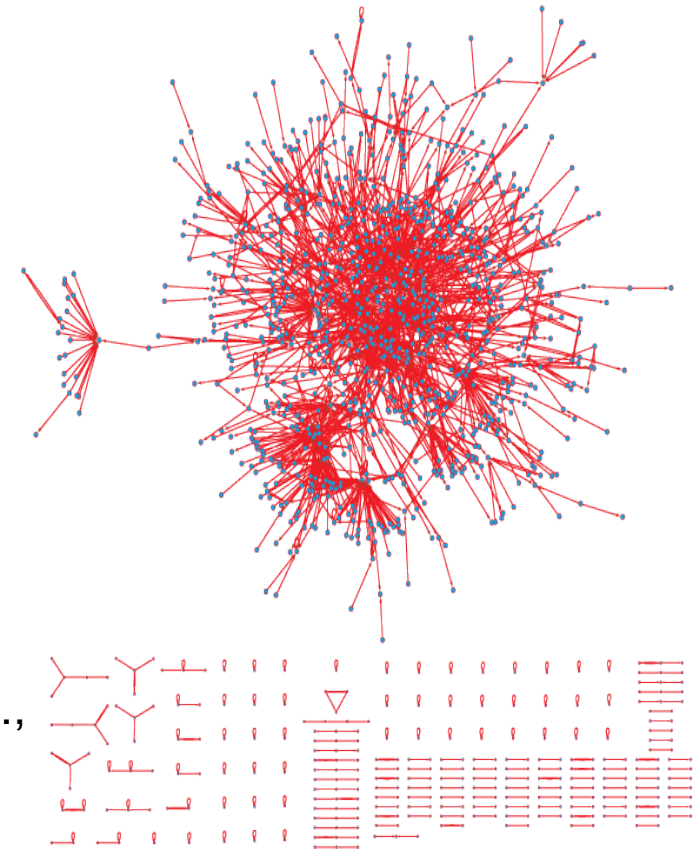
Example: yeast PPI network

Actual size:

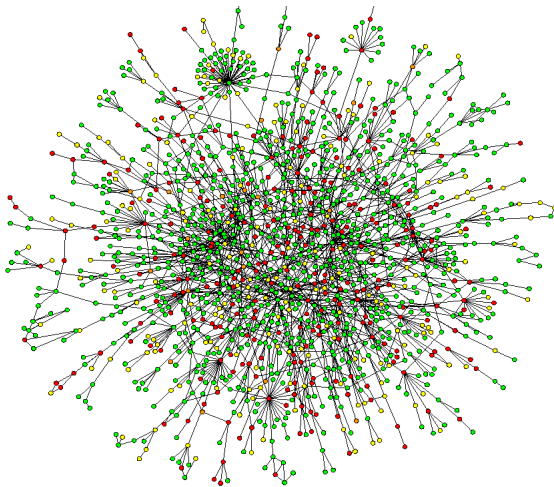
- ◇ ~6,000 nodes
→ Computational cost: ~18M pairs
- ◇ Estimated ~15,000 edges
→ Sparseness: 0.08% of all pairs (Yu et al., 2008)

Known interactions:

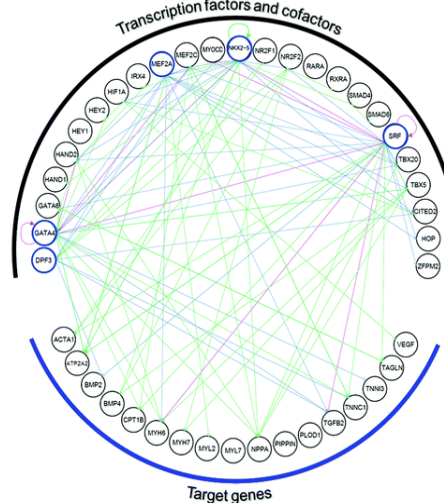
- ◇ Small-scale experiments: accurate but few
→ Overfitting: ~5,000 in BioGRID, involving ~2,300 proteins
- ◇ Large-scale experiments: abundant but noisy
→ Noise: false +ve/-ve for yeast two-hybrid data up to 45% and 90% (Huang et al., 2007)



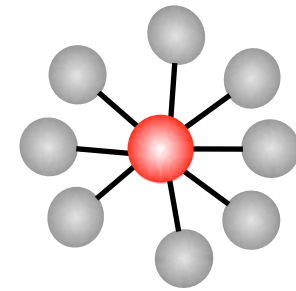
Different Types of Molecular Networks



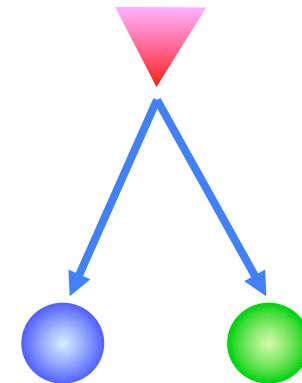
Protein-protein Interaction networks



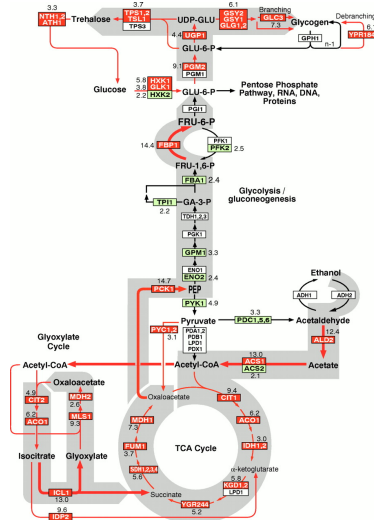
TF-target-gene Regulatory networks



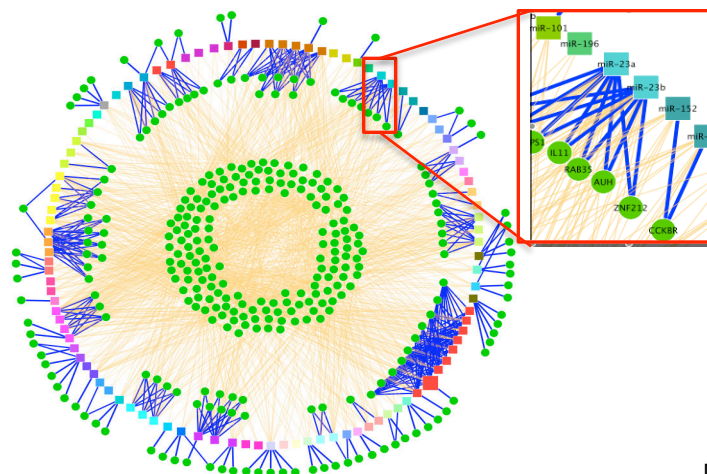
Undirected



Directed



Metabolic pathway networks



miRNA-target networks

[Toenjes, *et al*, *Mol. BioSyst.* (2008);
Jeong *et al*, *Nature* (2001); [Horak, *et al*,
Genes & Development, 16:3017-3033;
DeRisi, Iyer, and Brown, *Science*,
278:680-686]

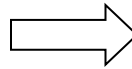
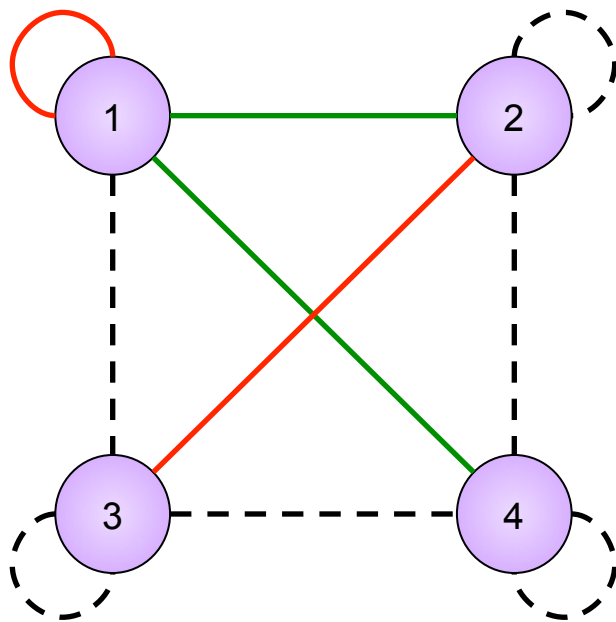
Predicting Networks

How do we construct large molecular networks?

From extrapolating correlations between functional genomics data with fairly small sets of known interactions, making best use of the known training data.



Training sets

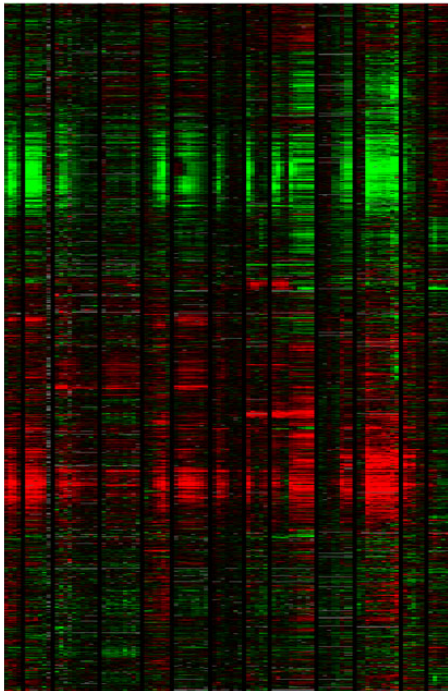


	1	2	3	4
1	0	1	?	1
2	1	?	0	?
3	?	0	?	?
4	1	?	?	?

- Known interactions
- Known non-interactions
- - - Unknown

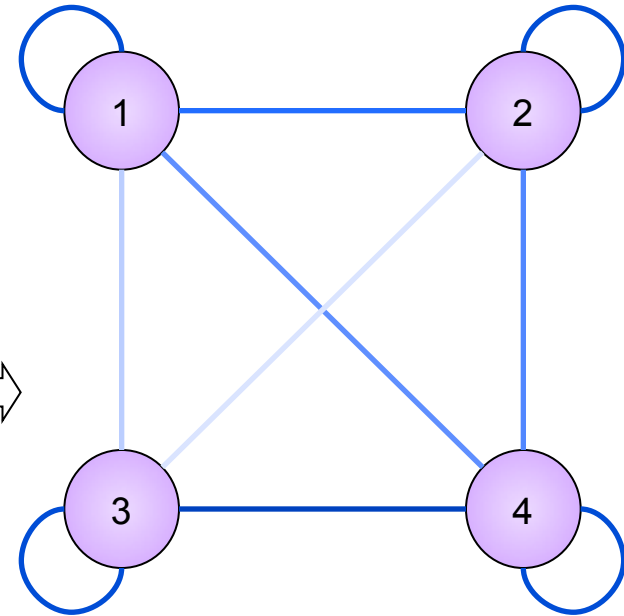
Network prediction: features

- Example 1: gene expression



Gasch et al., 2000

$x_1 = (0.2, 2.4, 1.5, \dots)$
 $x_2 = (0.8, 2.2, 1.5, \dots)$
 $\Rightarrow x_3 = (4.3, 0.1, 7.5, \dots) \Rightarrow$
 \dots
 $\text{sim}(x_1, x_2) = 0.62$
 $\text{sim}(x_1, x_3) = -0.58$
 \dots

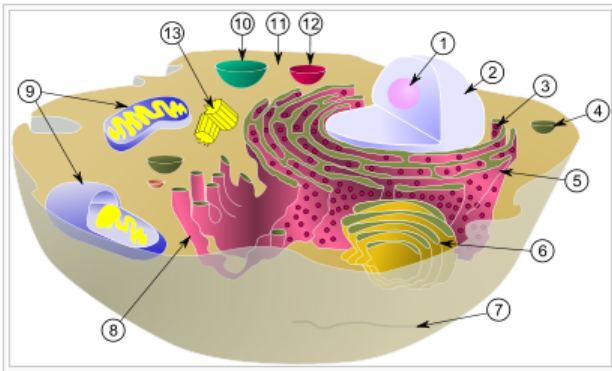


Similarity scale:



Network prediction: features

- Example 2: sub-cellular localization



<http://www.scq.ubc.ca/wp-content/yeasttwohybridtranscript.gif>

$$x_1 = (1, 1, 0, 0, \dots)$$

$$x_2 = (1, 1, 1, 0, \dots)$$

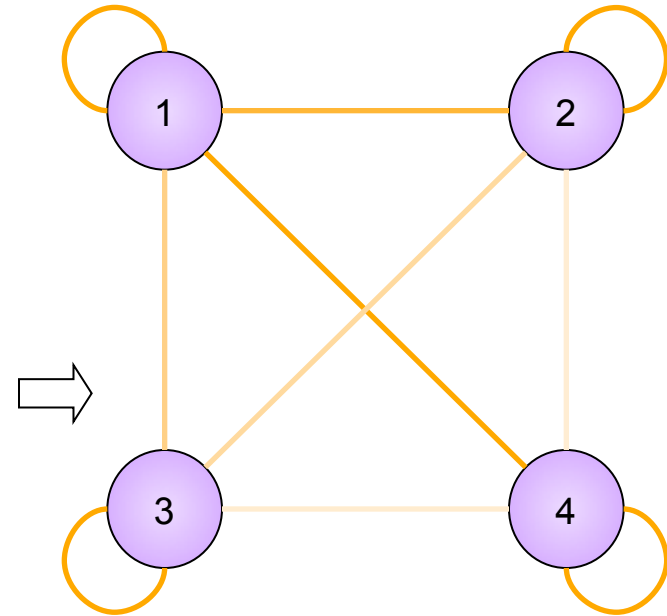
$$x_3 = (1, 0, 1, 0, \dots)$$

...

$$\text{sim}(x_1, x_2) = 0.81$$

$$\text{sim}(x_1, x_3) = 0.12$$

...



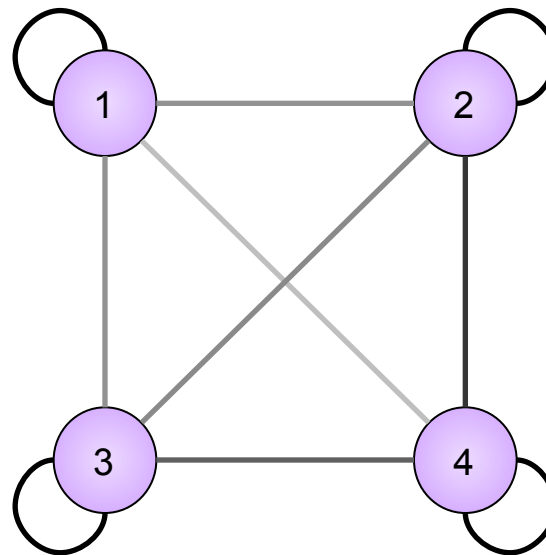
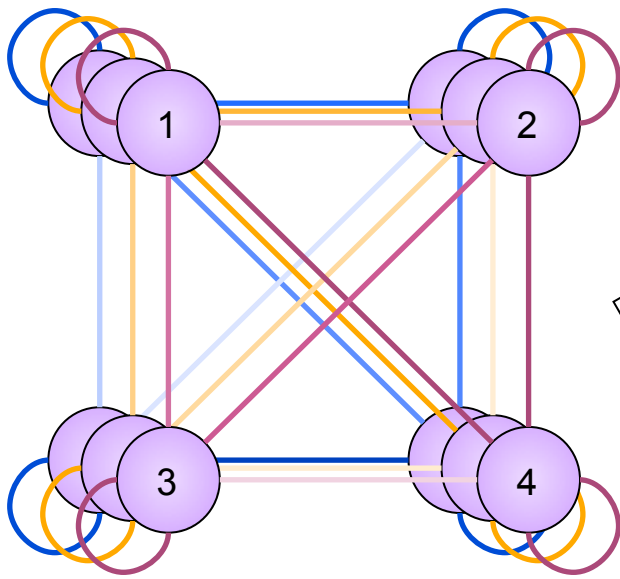
Similarity scale:

1



-1

Data integration & Similarity Matrix



	1	2	3	4
1	1.00	0.57	0.55	0.40
2	0.57	1.00	0.66	0.89
3	0.55	0.66	1.00	0.79
4	0.40	0.89	0.79	1.00

Learning methods

An endless list:

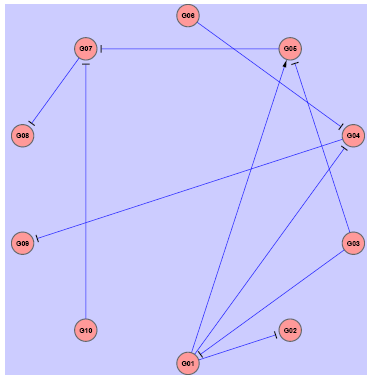
- Docking (e.g. Schoichet and Kuntz 1991)
- Evolutionary (e.g. Ramani and Marcotte, 2003)
- Topological (e.g. Yu et al., 2006)
- Bayesian (e.g. Jansen et al., 2003)
- **Kernel methods**
 - ◇ Global modeling:
 - em (Tsuda et al., 2003)
 - kCCA (Yamanishi et al., 2004)
 - kML (Vert and Yamanishi, 2005)
 - Pairwise kernel (Pkernel) (Ben-Hur and Noble, 2005)
 - ◇ Local modeling:
 - Local modeling (Bleakley et al., 2007)

Let's compare in a public challenge!

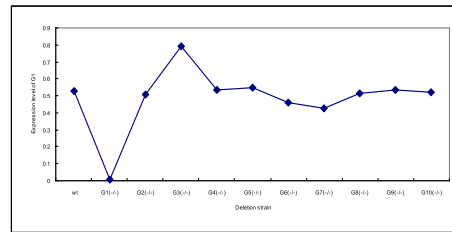
(DREAM: Dialogue for Reverse Engineering Assessment and Methods)

DREAM3: *in silico* regulatory network reconstruction

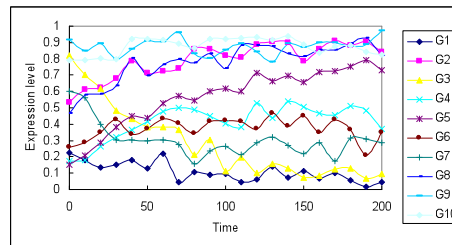
Actual network



Expression data

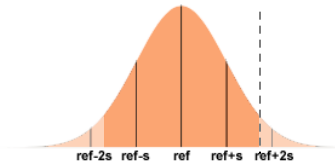


Deletion strains



**Time series after
initial perturbation**

Modeling



$$\text{Prob}(\text{signal}|\text{point}) = 2\Phi((\text{point} - \text{ref}) / s) - 1$$

Noise models

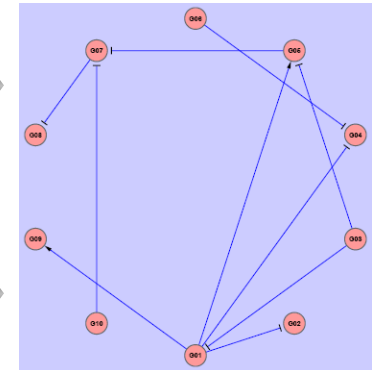
$$\frac{dy_j}{dt} = a_{j0} - a_{jj}y_j + \sum_{k \in S} a_{jk}y_k$$

$$\frac{dy_j}{dt} = \frac{b_{j1}}{1 + \exp\left(a_{j0} + \sum_{k \in S} a_{jk}y_k\right)} - b_{j2}y_j$$

$$\frac{dy_j}{dt} = a_{j0} \prod_{k_1 \in S_1} \left(\frac{b_{jk_1}}{y_{k_1}^{c_{jk_1}} + b_{jk_1}} \right) \prod_{k_2 \in S_2} \left(\frac{y_{k_2}^{c_{jk_2}}}{y_{k_2}^{c_{jk_2}} + b_{jk_2}} \right) - a_{j1}y_j$$

**Expression rate
models**

Predictions



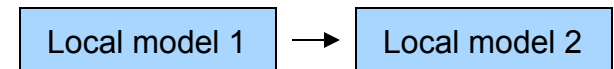
[Yip et al., DREAM3]

Accuracy (AUC)	E. Coli 1	E. Coli 2	Yeast 1	Yeast 2	Yeast 3
Size-10	0.928	0.912	0.949	0.747	0.714
Size-50	0.930	0.924	0.917	0.792	0.805
Size-100	0.948	0.960	0.915	0.856	0.783

Our work: efficiently propagating known information

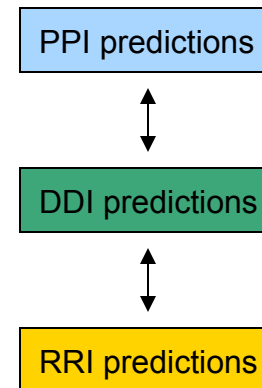
Training set expansion

- Motivation: lack of training examples
- Expand training sets horizontally



Multi-level learning

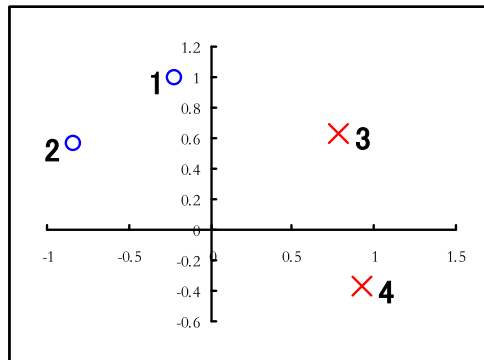
- Motivation: hierarchical nature of interaction
- Expand training sets vertically



DREAM3 *in silico* regulatory network reconstruction challenge

Kernels

Kernel: a similarity matrix that is positive semi-definite (p.s.d.)



Objects in an feature space

Compute
inner products
→
←
p.s.d. implies

	1	2	3	4
1	1.00	0.72	0.45	-0.56
2	0.72	1.00	-0.30	-0.98
3	0.45	-0.30	1.00	0.49
4	-0.56	-0.98	0.49	1.00

Similarity matrix

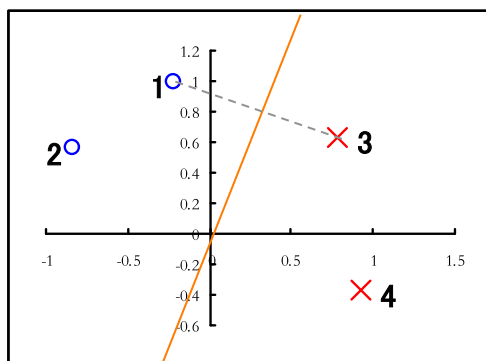
Good for integrating heterogeneous datasets (protein sequences, PSSM, gene expression, ...)

– no need to explicitly place them in a common feature space

Kernel methods

Use the kernel as proxy to work in the feature space

Example: SVM (finding the best separating hyperplane)

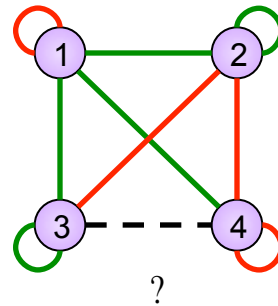


Equivalent to

$$\begin{aligned} &\text{Maximize} \quad \sum_i \lambda_i - \frac{1}{2} \sum_i \sum_j \lambda_i \lambda_j y_i y_j \langle x_i, x_j \rangle \\ &\text{Subject to} \quad \lambda \geq 0 \\ &\quad \quad \quad \sum_i \lambda_i y_i = 0 \end{aligned}$$

The only thing that we need to know about the objects: their similarity values (inner products)

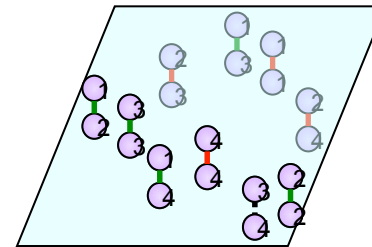
Kernel methods for predicting networks: local vs. global modeling



Global modeling: build one model for the whole network

Pairwise kernel: consider object pairs instead of individual objects

Problem: $O(n^2)$ instances, $O(n^4)$ kernel elements



Direct methods: threshold the kernel to make predictions

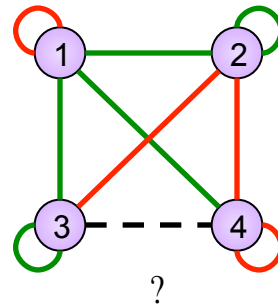
Problem: One single global model, may not be able to handle subclasses

	1	2	3	4
1	1.00	0.57	0.55	0.40
2	0.57	1.00	0.66	0.89
3	0.55	0.66	1.00	0.79
4	0.40	0.89	0.79	1.00

Threshold: 0.7

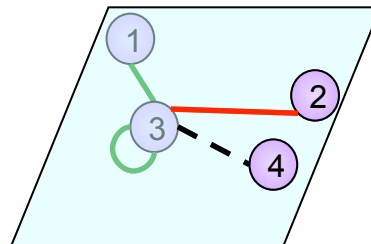
	1	2	3	4
1	1.00	0.57	0.55	0.40
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3	0.55	0.66	1.00	0.79
4	0.40	0.89	0.79	1.00

Kernel methods for predicting networks: local vs. global modeling



Local modeling: build one model for each node

Model for node 3:



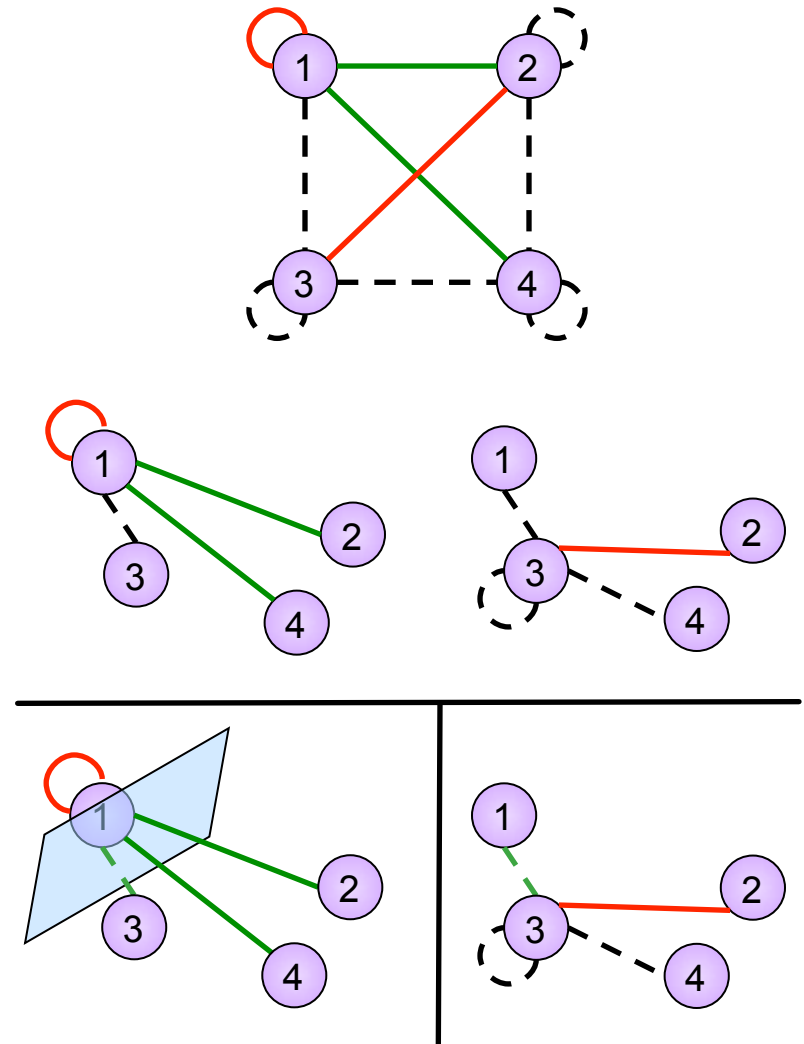
Problem: insufficient and unevenly distributed training data (what if node 3 has no known interactions at all?)

Our work: training set expansion

- Goal:
 - ◇ Utilize the flexibility of local modeling
 - ◇ Tackle the problem of insufficient training data
- Idea: generate auxiliary training data
 - ◇ Prediction propagation
 - ◇ Kernel initialization

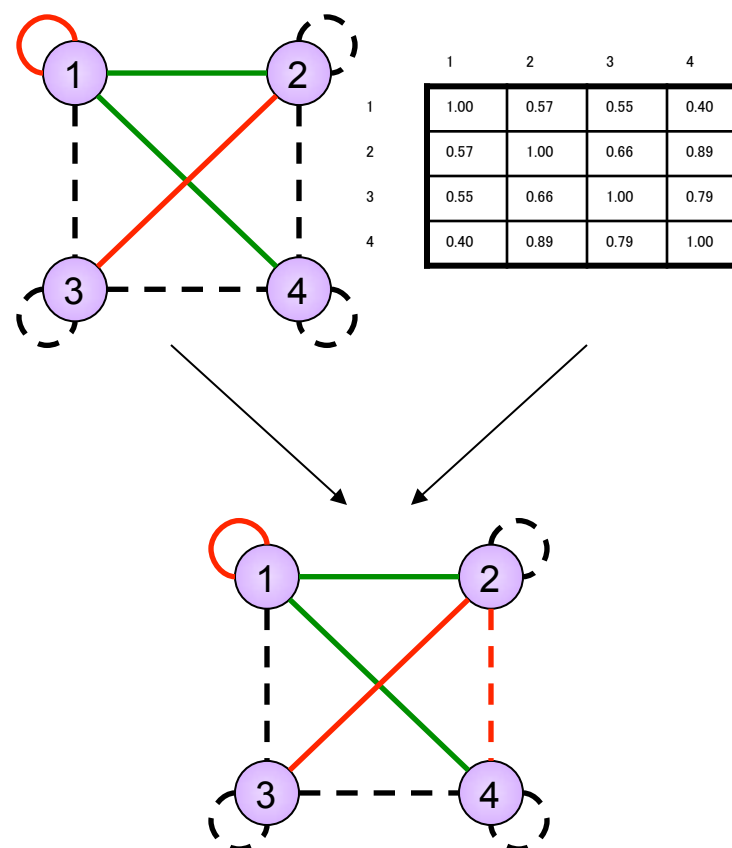
Prediction propagation

- Motivation: some objects have more examples than others
- Our approach:
 - ◇ Learn models for objects with more examples first
 - ◇ Propagate the most confident predictions as auxiliary examples of other objects



Kernel initialization

- Motivation: what if most objects have very few examples?
- Our approach (inspired by the direct method):
 - ◇ Add the most similar pairs in the kernel as positive examples
 - ◇ Add the most dissimilar pairs in the kernel as negative examples



Remarks

- Can be used in combination
- Prediction propagation theoretically related to co-training (Blum and Mitchell, 1998)
 - ◊ Semi-supervised
 - Similarity with PSI-BLAST
- Algorithm complexity $O(nf(n))$ of local modeling vs. $O(f(n^2))$ of global modeling

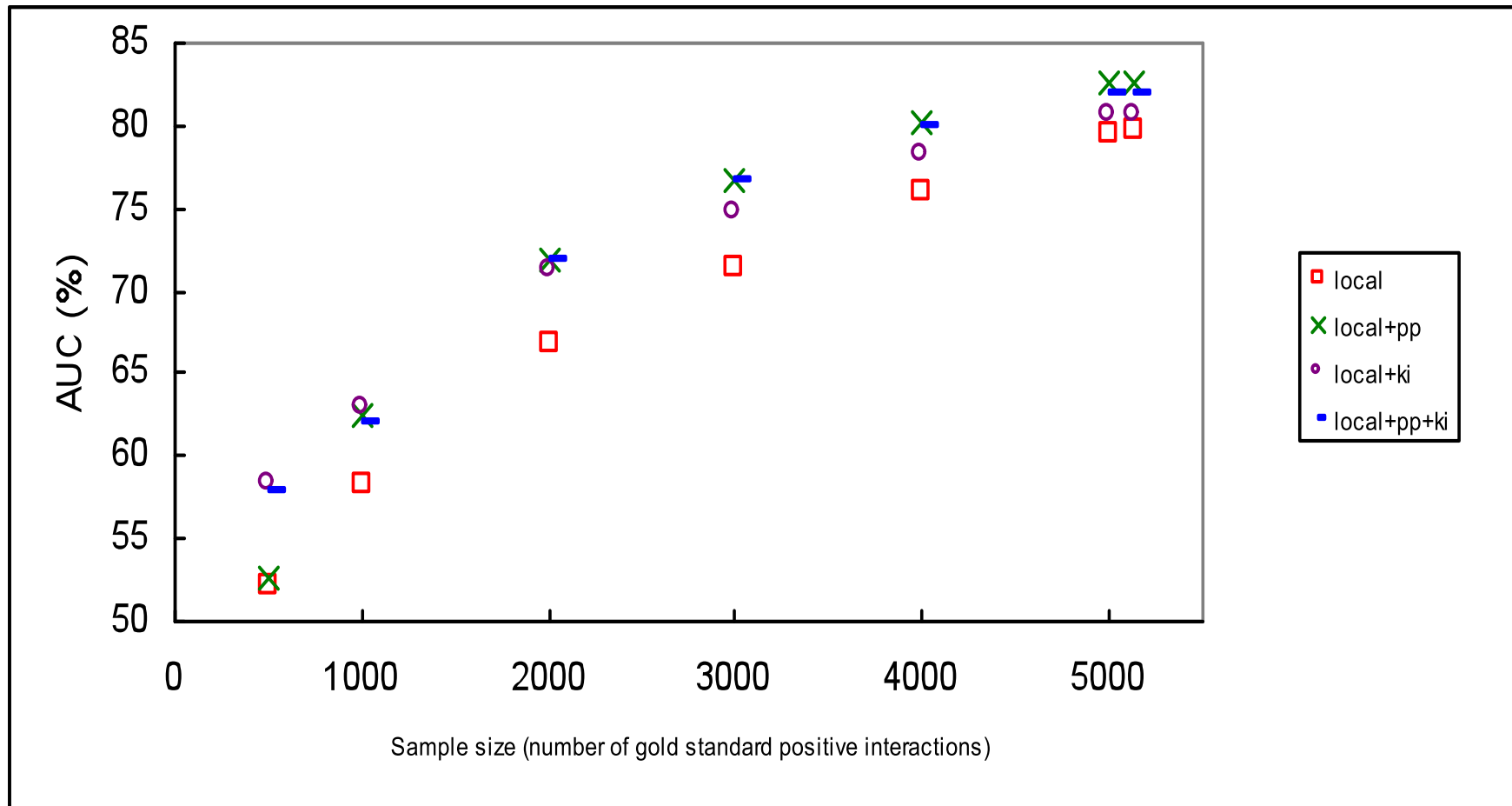
Prediction accuracy (AUC)

	phy	loc	exp-gasch	exp-spellman	y2h-ito	y2h-uetz	tap-gavin	tap-krogan	int
Mode 1									
direct	58.04	66.55	64.61	57.41	51.52	52.13	59.37	61.62	70.91
kCCA	65.80	63.86	68.98	65.10	50.89	50.48	57.56	51.85	80.98
kML	63.87	68.10	69.67	68.99	52.76	53.85	60.86	57.69	73.47
em	71.22	75.14	67.53	64.96	55.90	53.13	63.74	68.20	81.65
local	71.67	71.41	72.66	70.63	67.27	67.27	64.60	67.48	75.65
local+pp	73.89	75.25	77.43	75.35	71.60	71.51	74.62	71.39	83.63
local+ki	71.68	71.42	75.89	70.96	69.40	69.05	70.53	72.03	81.74
local+pp+ki	72.40	75.19	77.41	73.81	70.44	70.57	73.59	72.64	83.59

Observations:

- Highest accuracy by training set expansion
- Over fitting of local modeling without training set expansion
- Prediction propagation theoretically related to co-training (Blum and Mitchell, 1998)
 - ◇ Semi-supervised (Similarity with PSI-BLAST)

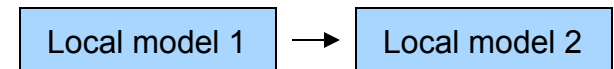
Complementarity of the two methods



From horizontal to vertical

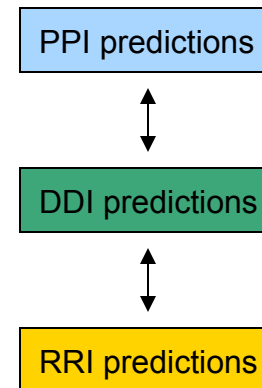
Training set expansion

- Motivation: lack of training examples
- Expand training sets horizontally

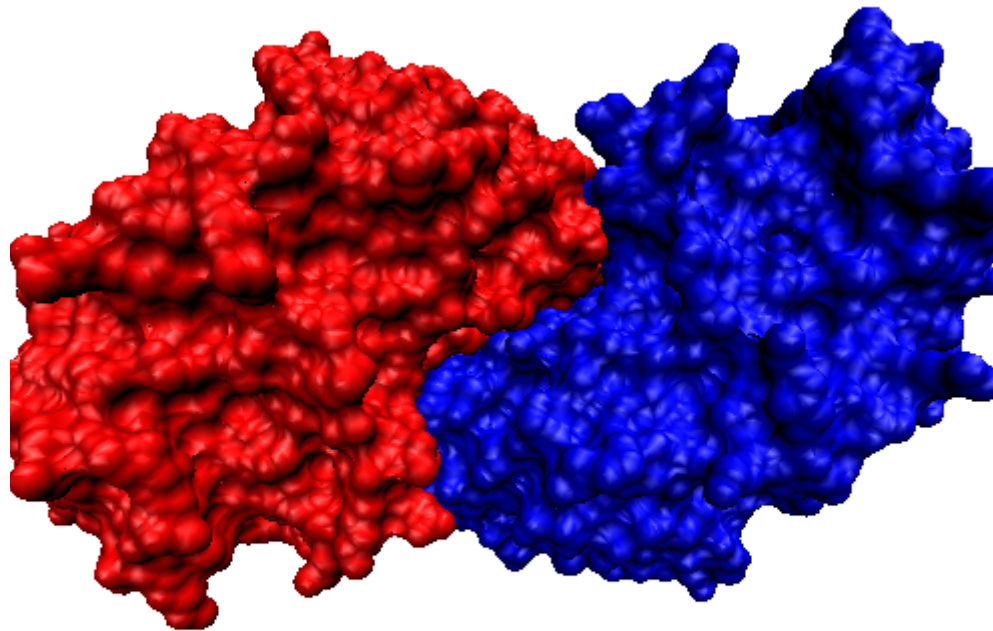


Multi-level learning

- Motivation: hierarchical nature of interaction
- Expand training sets vertically



Protein interaction

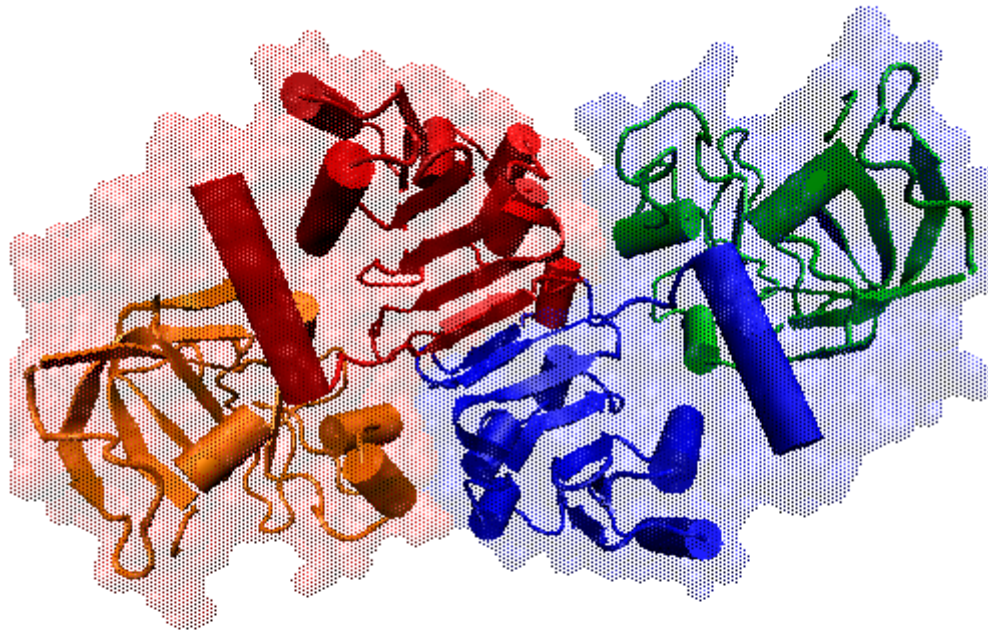


Yeast NADP-dependent alcohol dehydrogenase 6 (PDB: 1piw)

Protein-level features for interaction prediction: functional genomic information

[Yip and Gerstein, BMC Bioinfo. ('09, press)]

Domain interaction

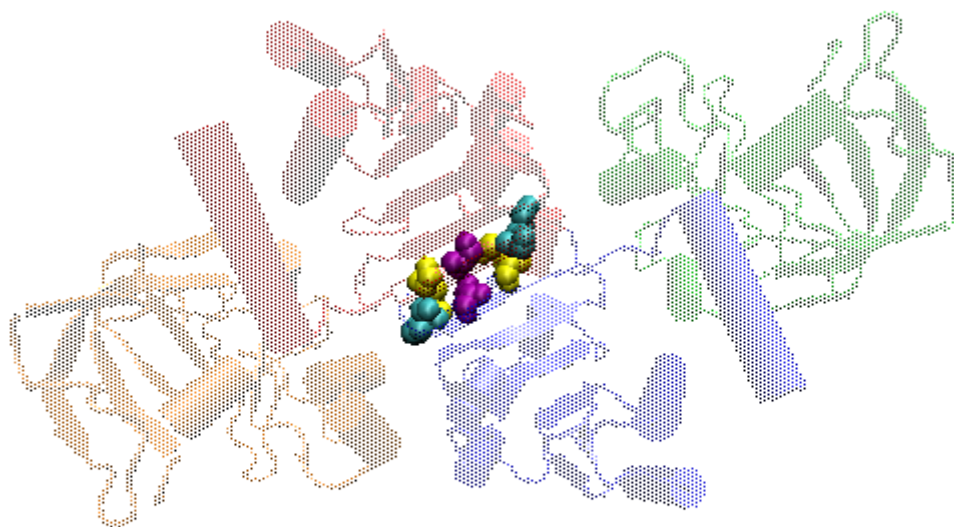


Pfam domains: PF00107 (inner) and PF08240 (outer)

Domain-level features for interaction prediction: evolutionary information

[Yip and Gerstein, BMC Bioinfo. ('09, press)]

Residue interaction

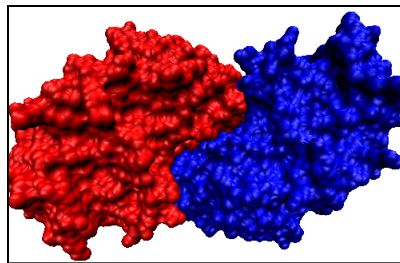


Interacting residues: 283 (yellow) with 287 (cyan), and 285 (purple) with 285

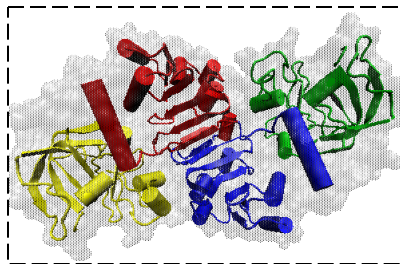
Residue-level features for interaction prediction: physical-chemical information

[Yip and Gerstein, BMC Bioinfo. ('09, press)]

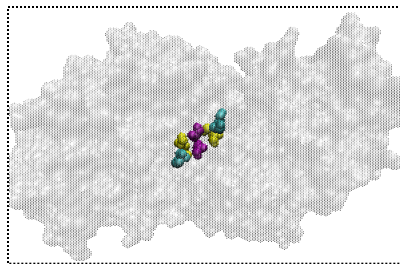
Combining the three problems



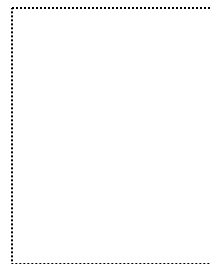
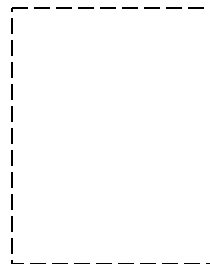
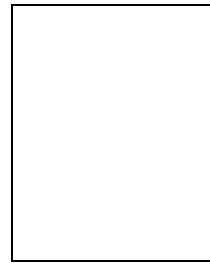
Protein interactions



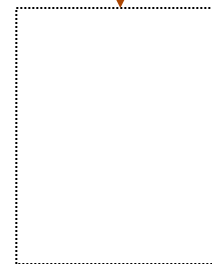
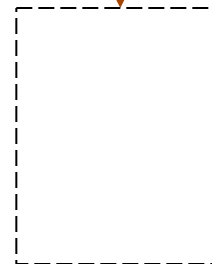
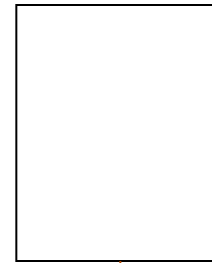
Domain interactions



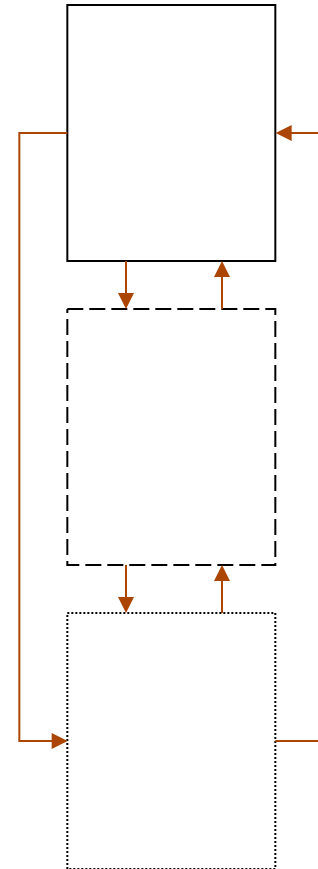
Residue interactions



i. Independent levels



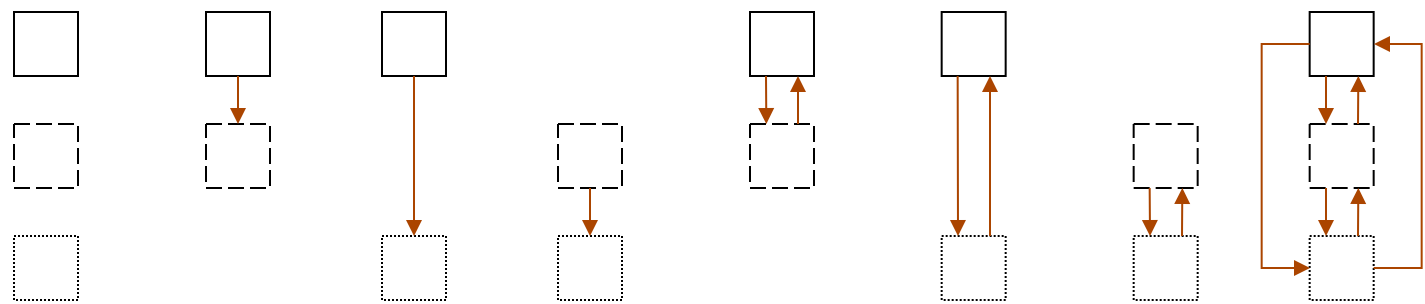
ii. Unidirectional flow



iii. Bidirectional flow

Empirical results (AUCs)

	Ind. levels	Unidirectional flow			Bidirectional flow			
Level		PD	PR	DR	PD	PR	DR	PDR
Proteins	71.68				72.23	72.50		72.82
Domains	53.18	61.51			71.71		68.94	71.20
Residues	57.36		54.89	53.81		72.26	63.16	77.86



- Highest accuracy by bidirectional flow
- Additive effect: 2 vs. 3 levels

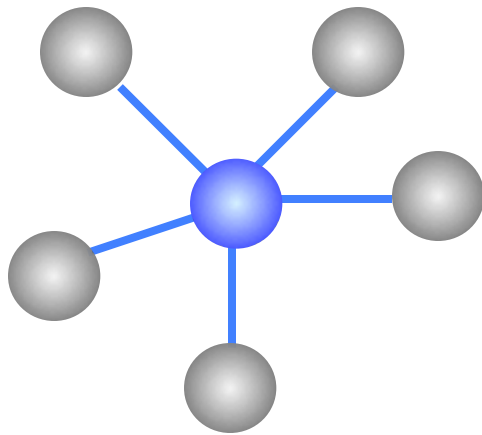
Finding Central Points in Networks: Hubs & Bottlenecks

Where are key points networks ? How do we locate them ?

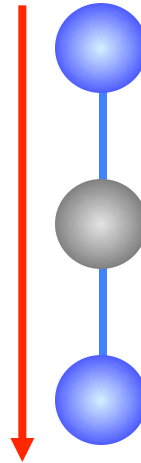


Global topological measures

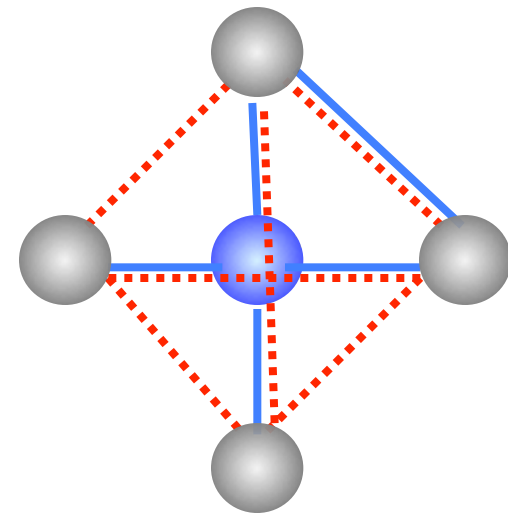
Indicate the gross topological structure of the network



Degree (K)
5



Path length (L)
2

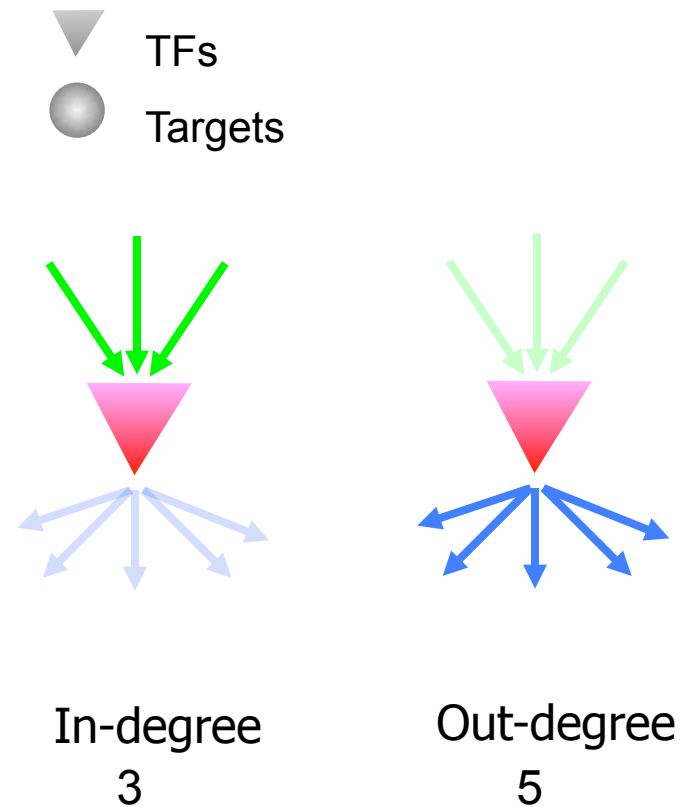


Clustering coefficient (C)
 $1/6$

Interaction and expression networks are ***undirected***

[Barabasi]

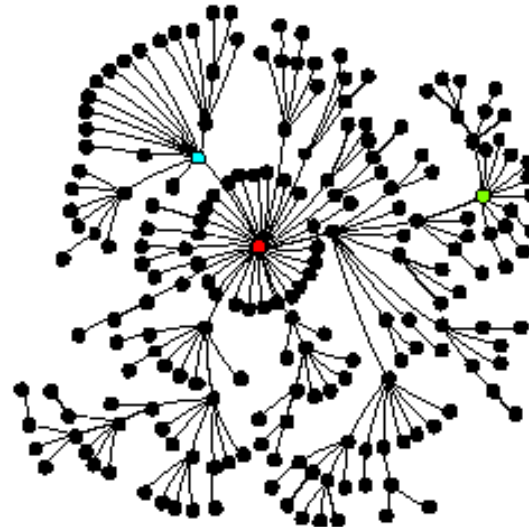
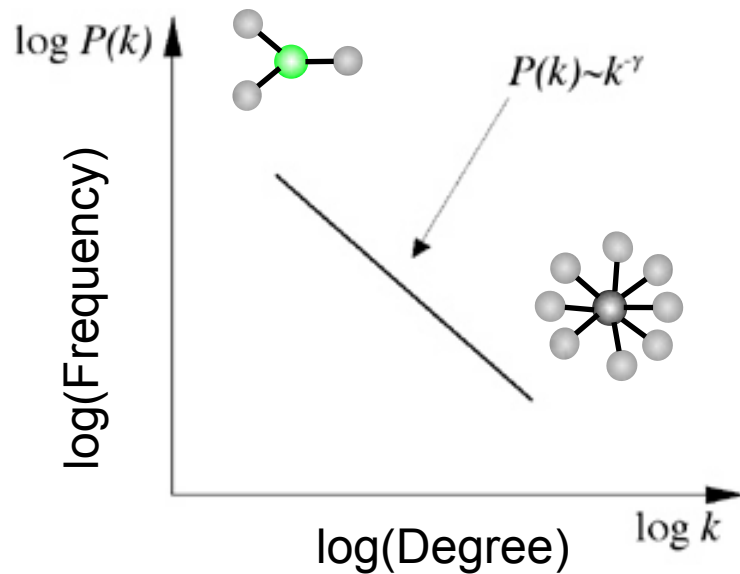
Global topological measures for directed networks



Regulatory and metabolic networks are ***directed***

Scale-free networks

Power-law distribution



Hubs dictate the structure of the network

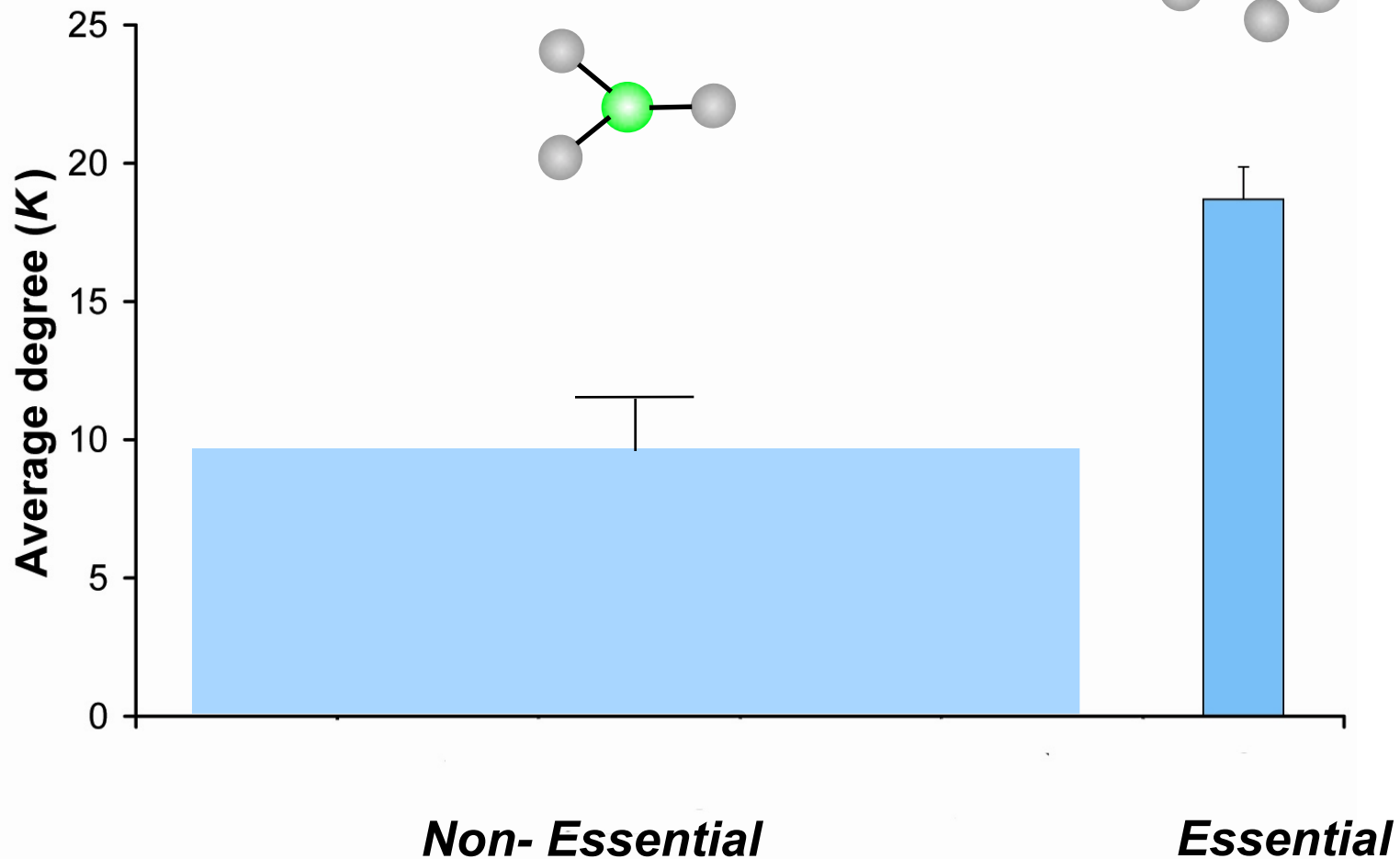
[Barabasi]

Hubs tend to be Essential

Integrate gene essentiality data with protein interaction network. Perhaps hubs represent vulnerable points?

[Lauffenburger, Barabasi]

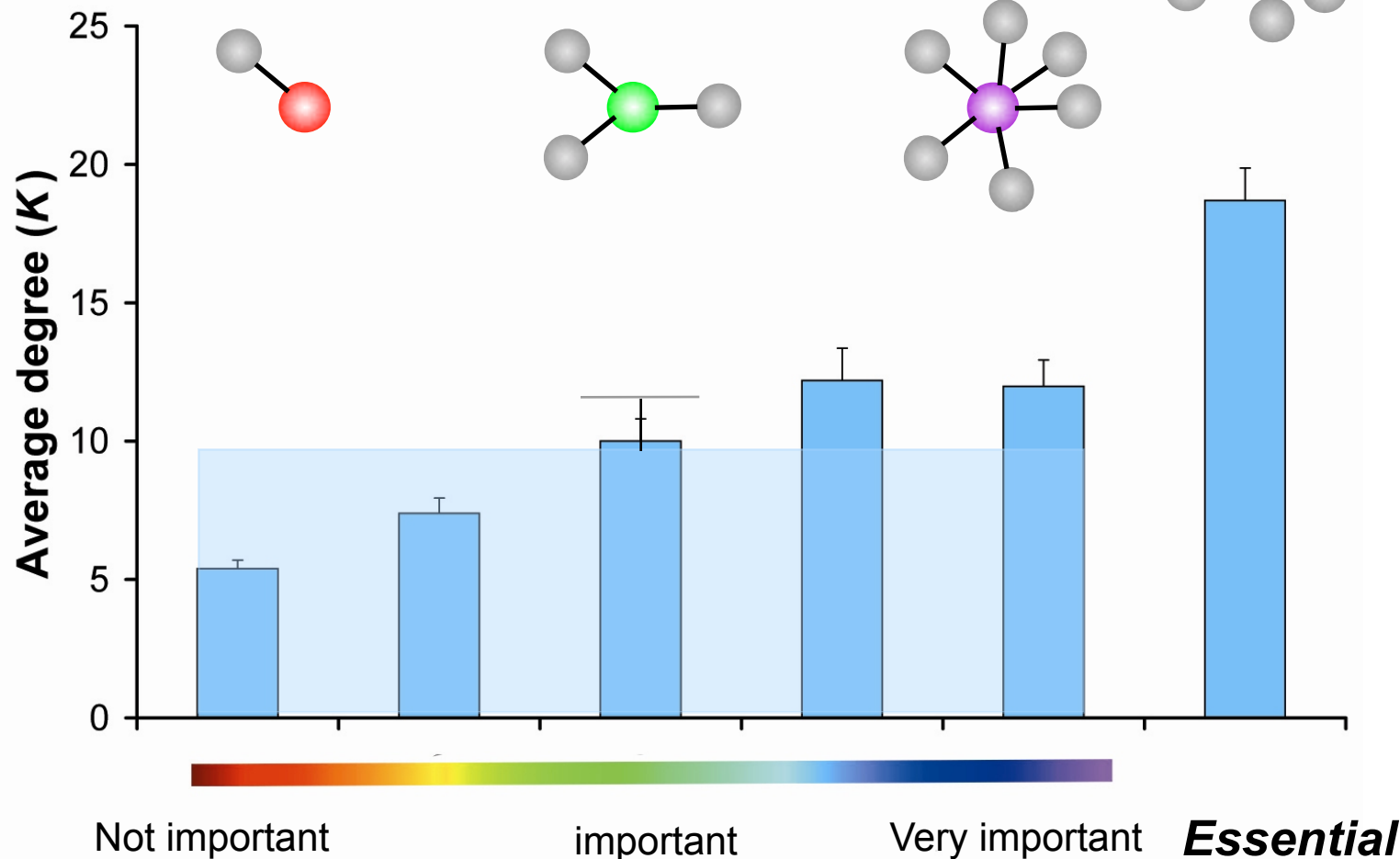
"hubbiness"



Relationships extends to "Marginal Essentiality"

Marginal essentiality measures relative importance of each gene (e.g. in growth-rate and condition-specific essentiality experiments) and scales continuously with "hubbiness"

"hubbiness"

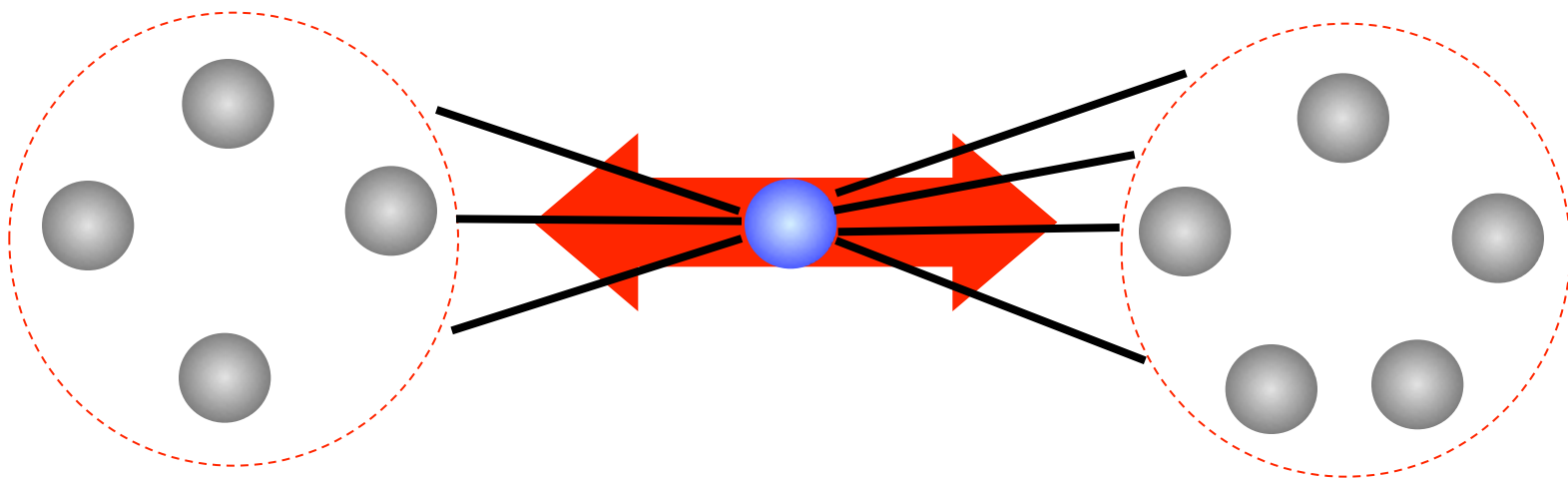


Another measure of Centrality: Betweenness centrality

Betweenness of a node is the number of shortest paths of pairs of vertices that run through it -- a measure of information flow.

Freeman LC (1977) Set of measures of centrality based on betweenness.
Sociometry 40: 35–41.

Girvan & Newman (2002) PNAS 99: 7821.



Betweenness centrality -- Bottlenecks

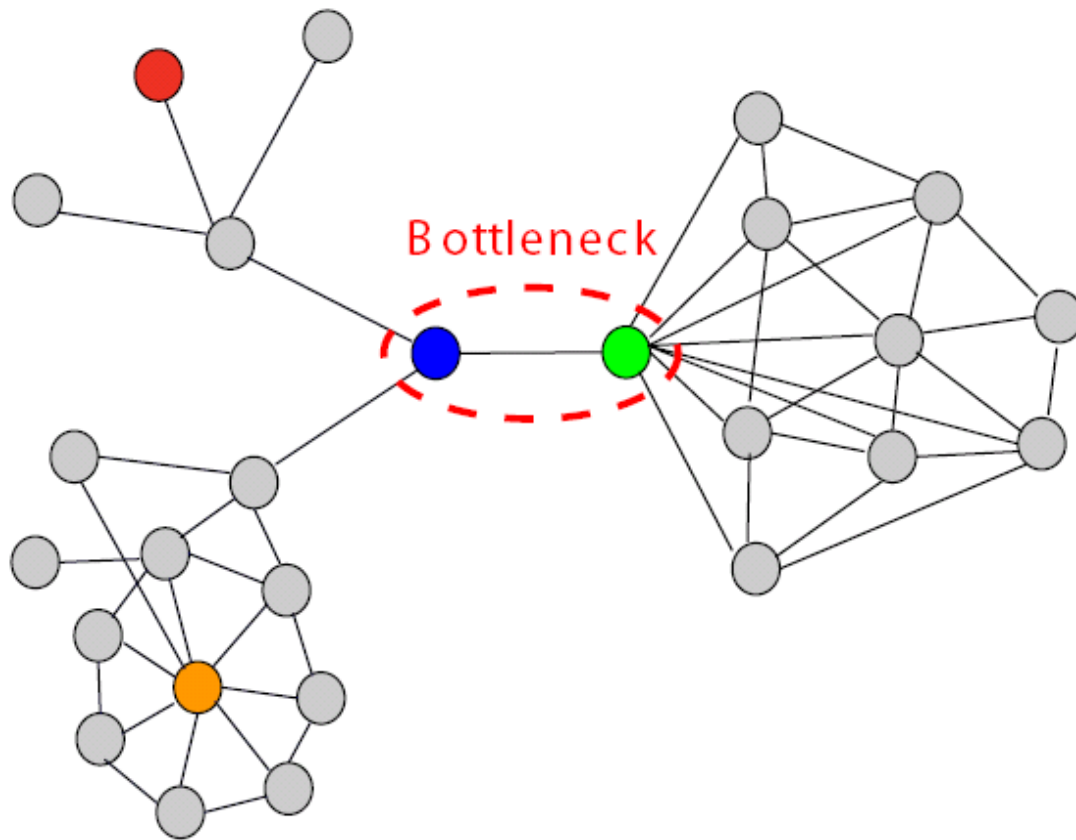
Proteins with high betweenness are defined as *Bottlenecks* (top 20%), in analogy to the traffic system







George Washington
Bridge



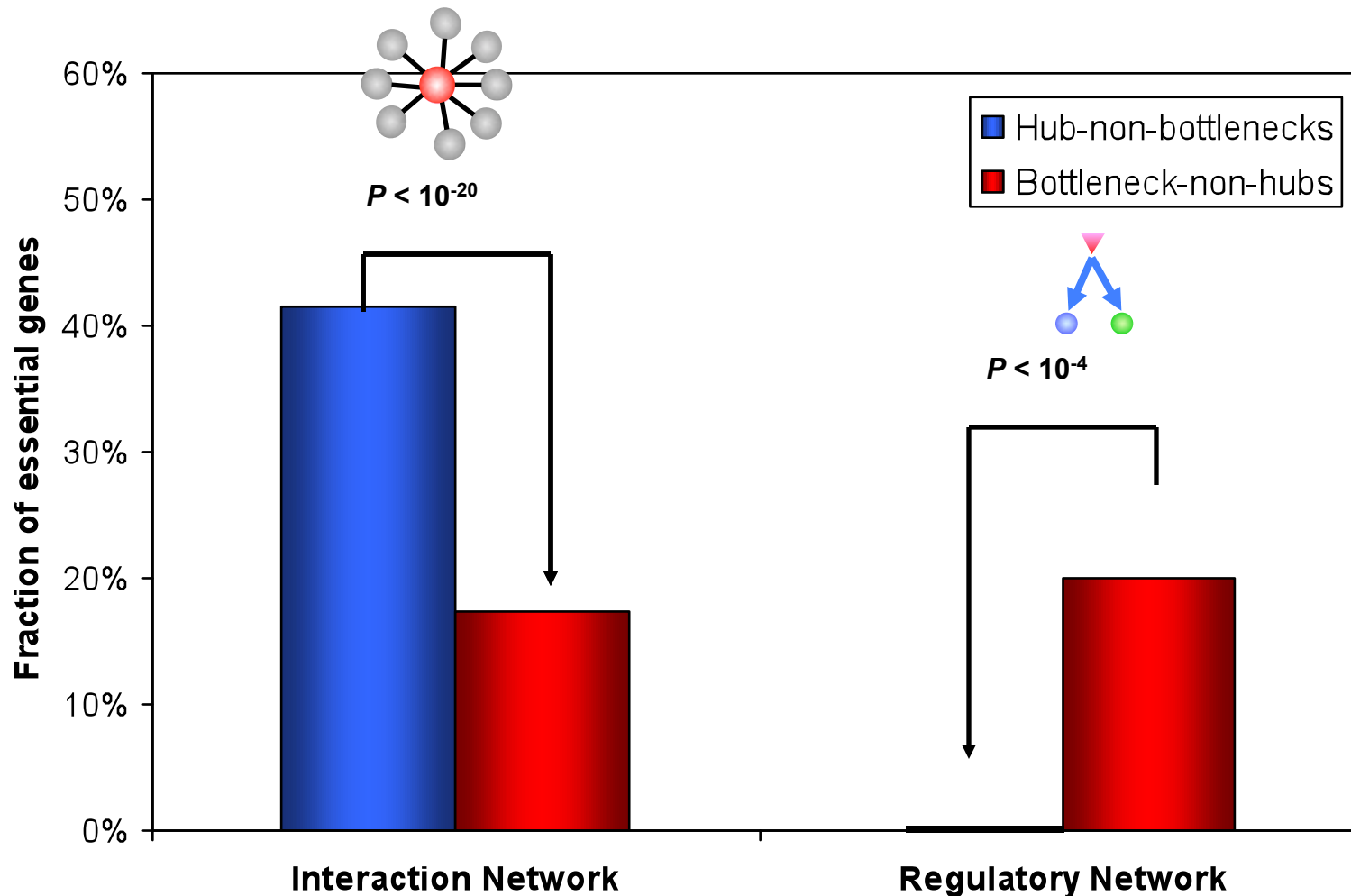
Bottlenecks & Hubs



-  Hub-bottleneck **node**
-  Non-hub-bottleneck **node**
-  Hub-non-bottleneck **node**
-  Non-hub-non-bottleneck **node**

[Yu et al., PLOS CB (2007)]

Bottlenecks are what matters in regulatory networks



[Yu et al., PLoS Comput Biol (2007)]

Finding Central Points in Networks #2: Tops of the Hierarchy

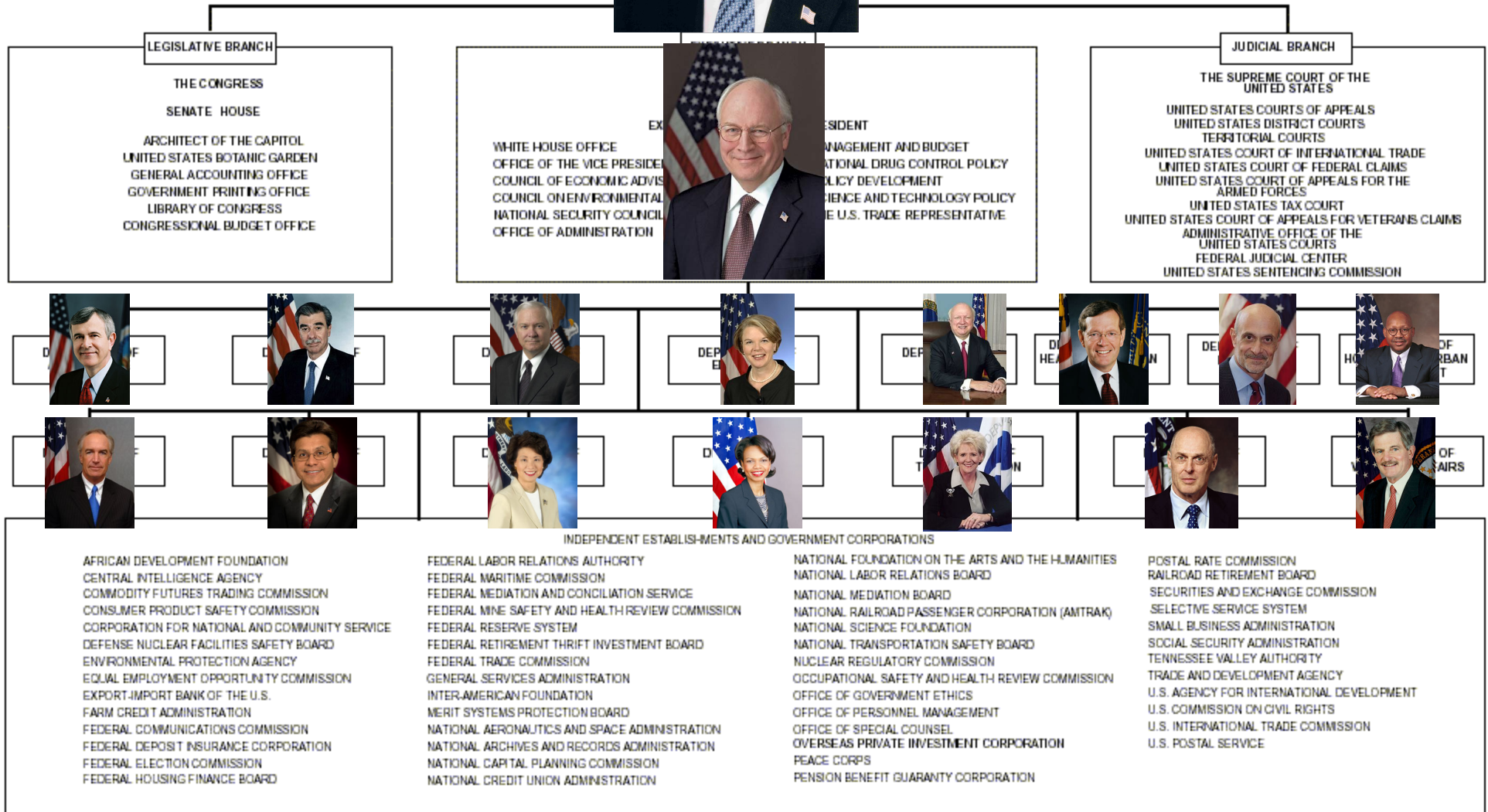
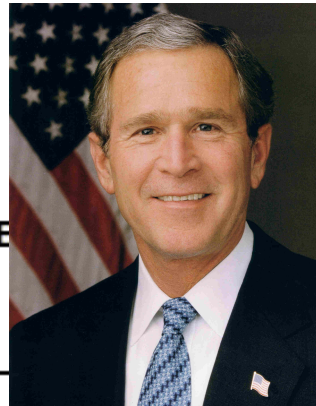
Where are key points networks ? How do we locate them ?



Social Hierarchy

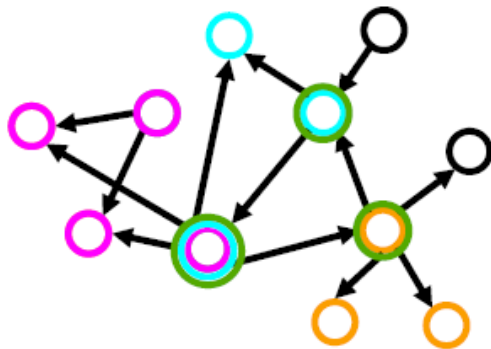
THE GOVERNMENT

UNITED STATES

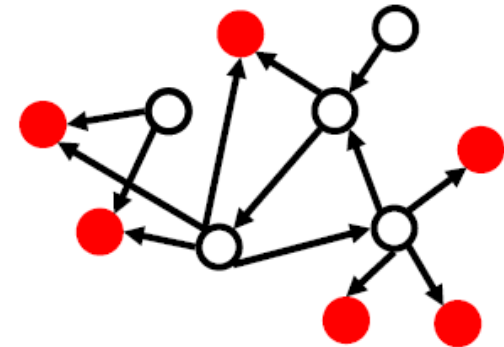


Determination of "Level" in Regulatory Network Hierarchy with Breadth-first Search

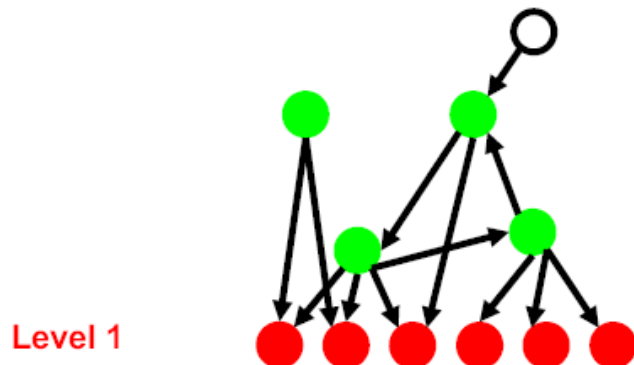
I. Example network with all 4 motifs



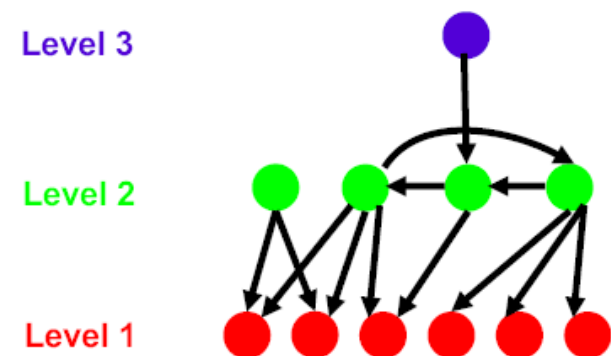
II. Finding terminal nodes (Red)



III. Finding mid-level nodes (Green)

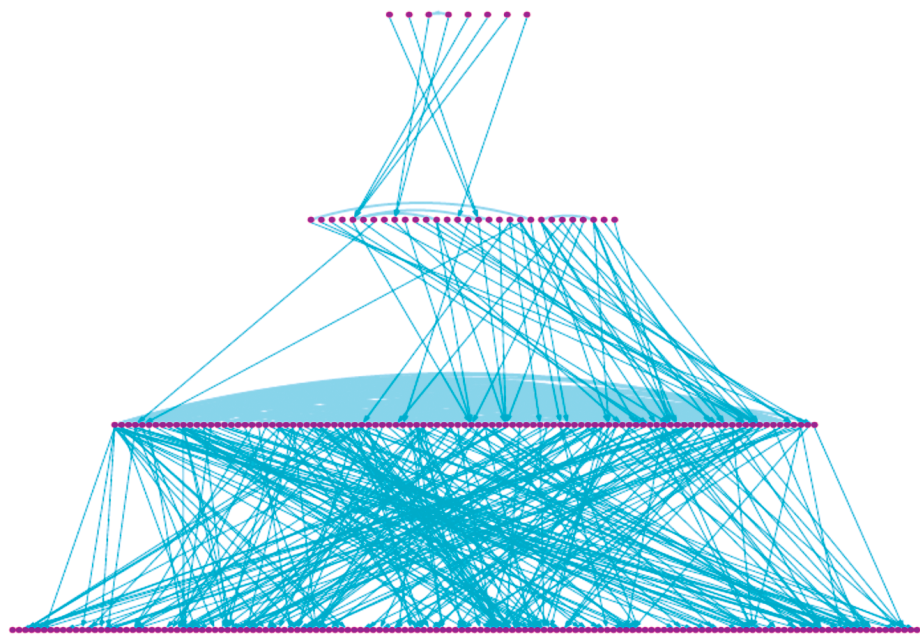


IV. Finding top-most nodes (Blue)

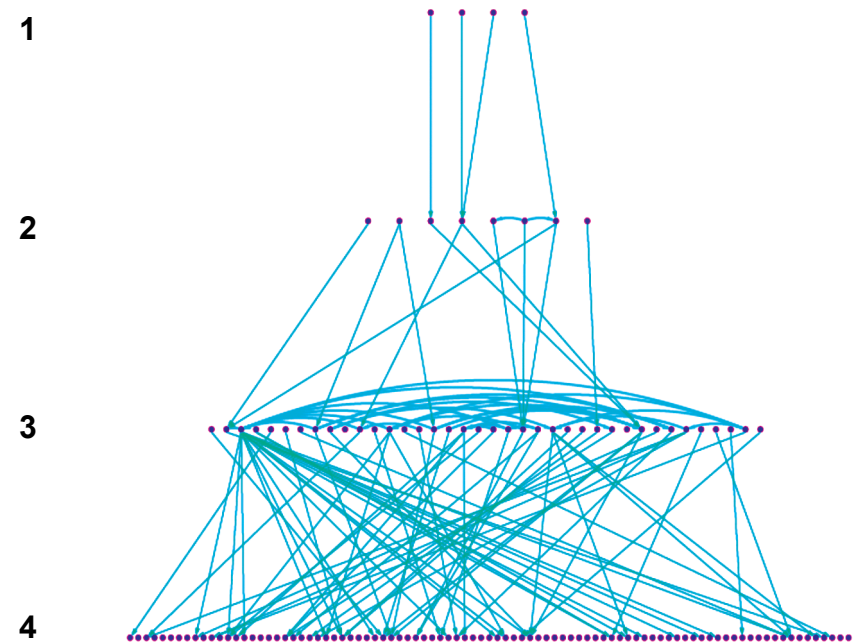


[Yu et al., PNAS (2006)]

Regulatory Networks have similar hierarchical structures



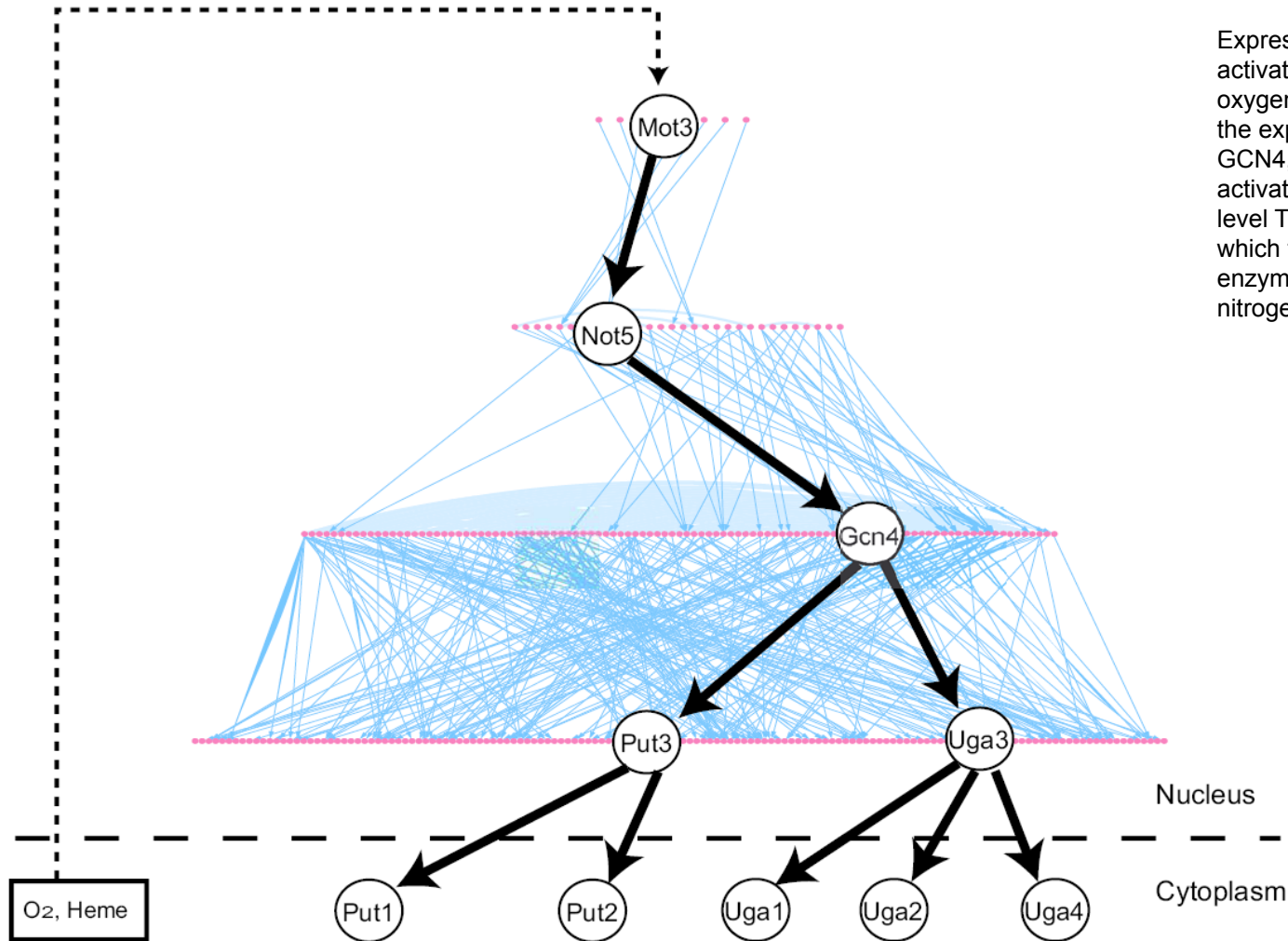
S. cerevisiae



E. coli

[Yu et al., Proc Natl Acad Sci U S A (2006)]

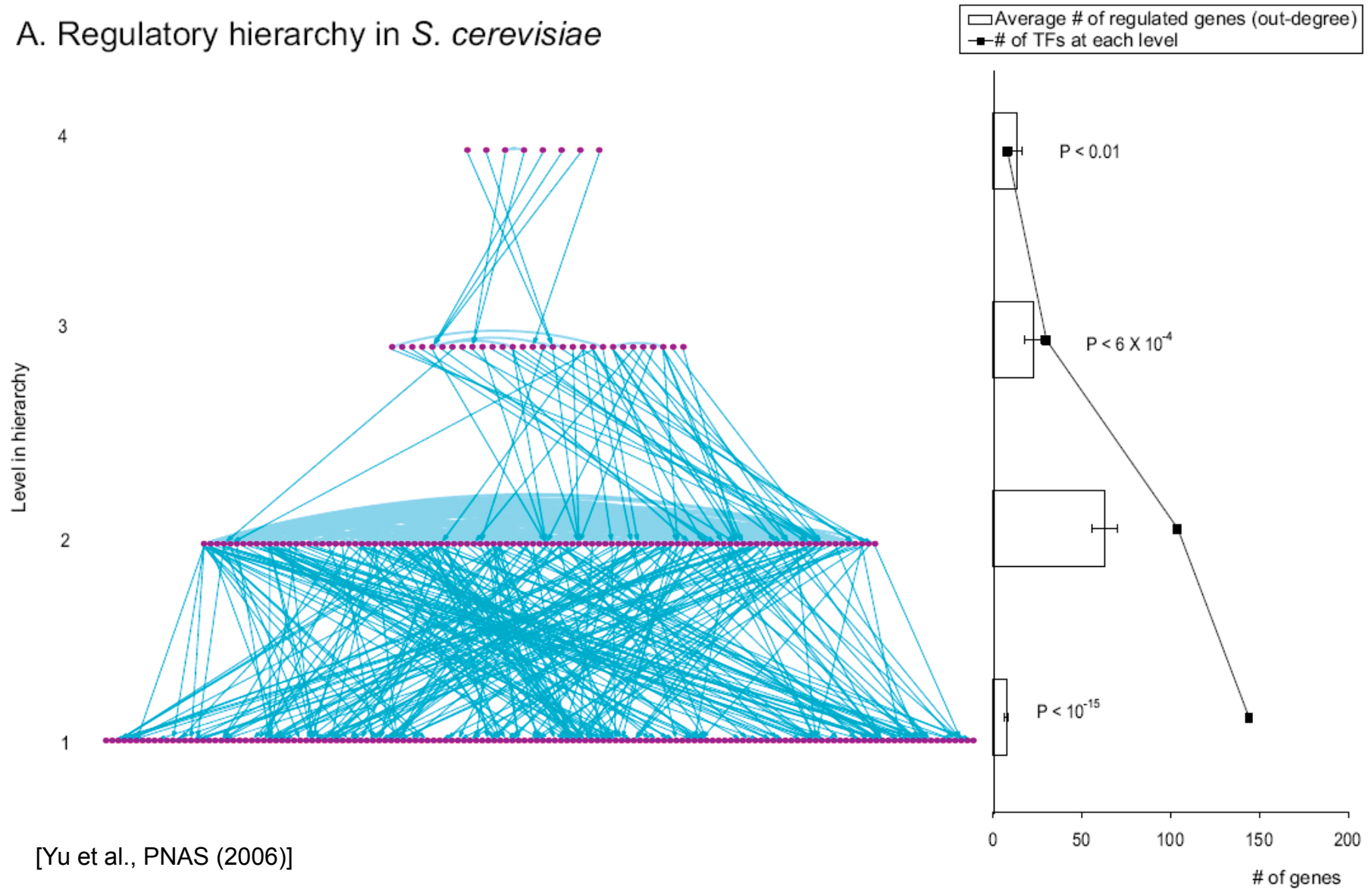
Example of Path Through Regulatory Network



[Yu et al., PNAS (2006)]

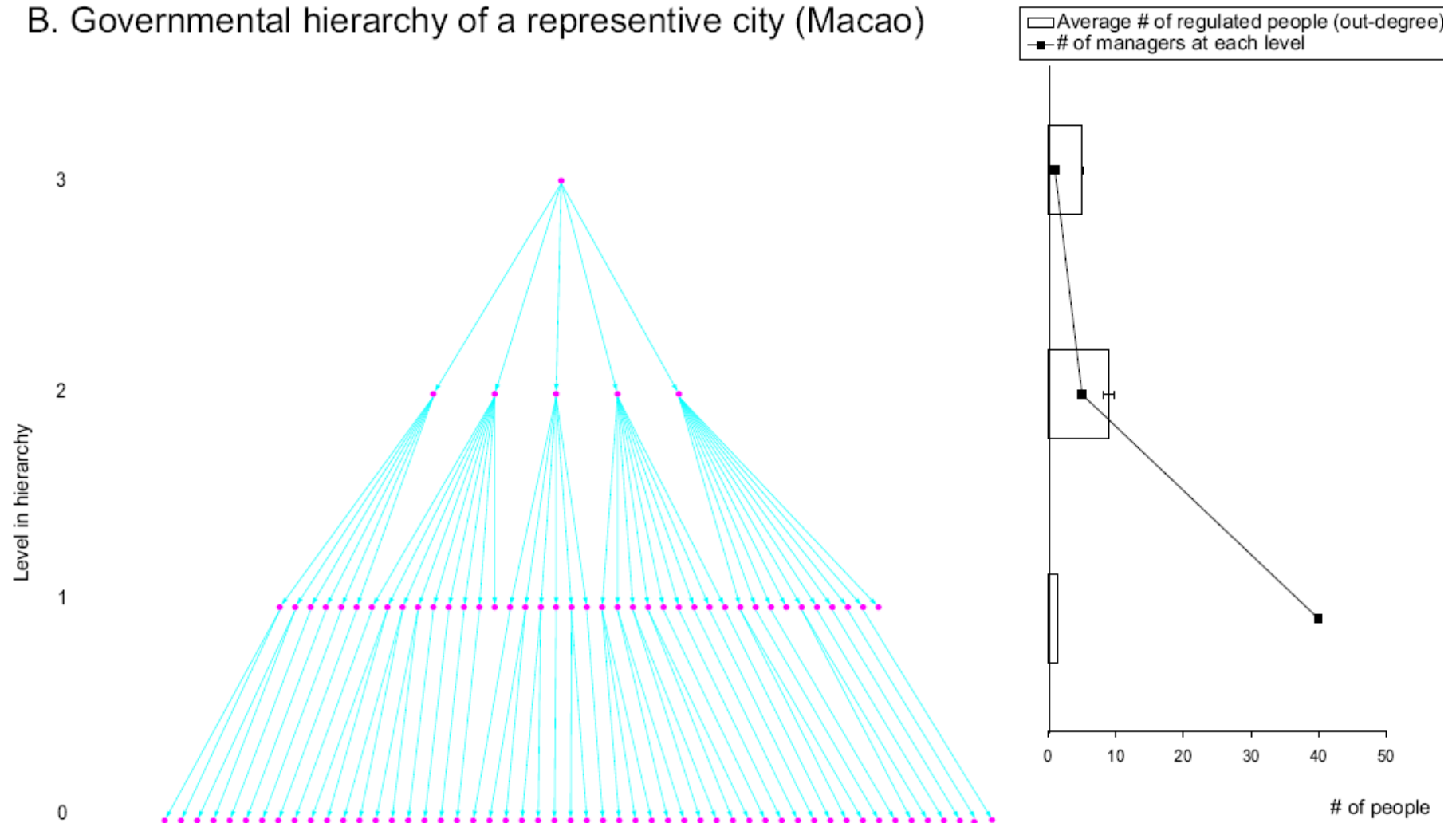
Yeast Regulatory Hierarchy: the Middle-managers Rule

A. Regulatory hierarchy in *S. cerevisiae*



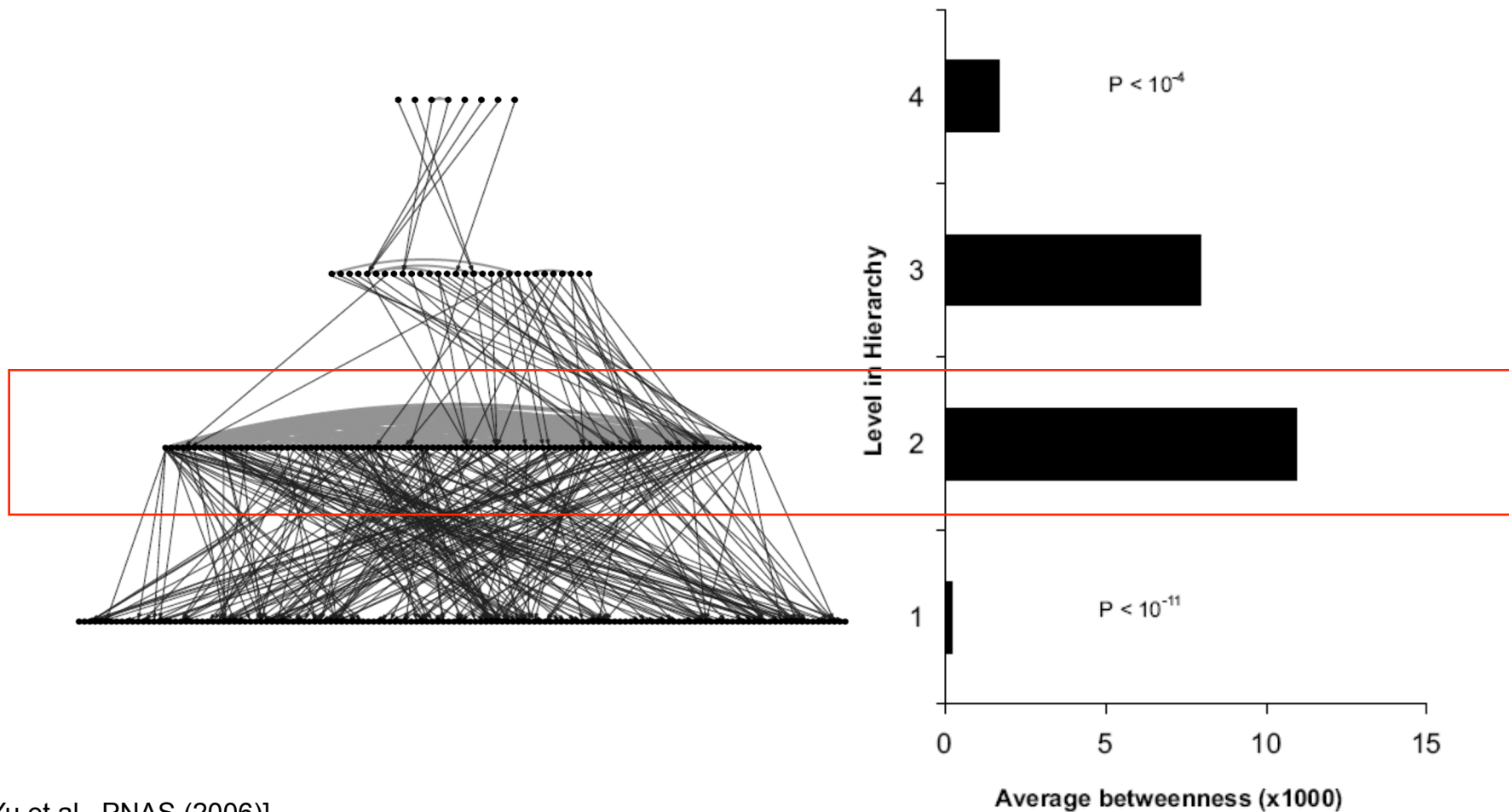
Yeast Network Similar in Structure to Government Hierarchy with Respect to Middle-managers

B. Governmental hierarchy of a representative city (Macao)



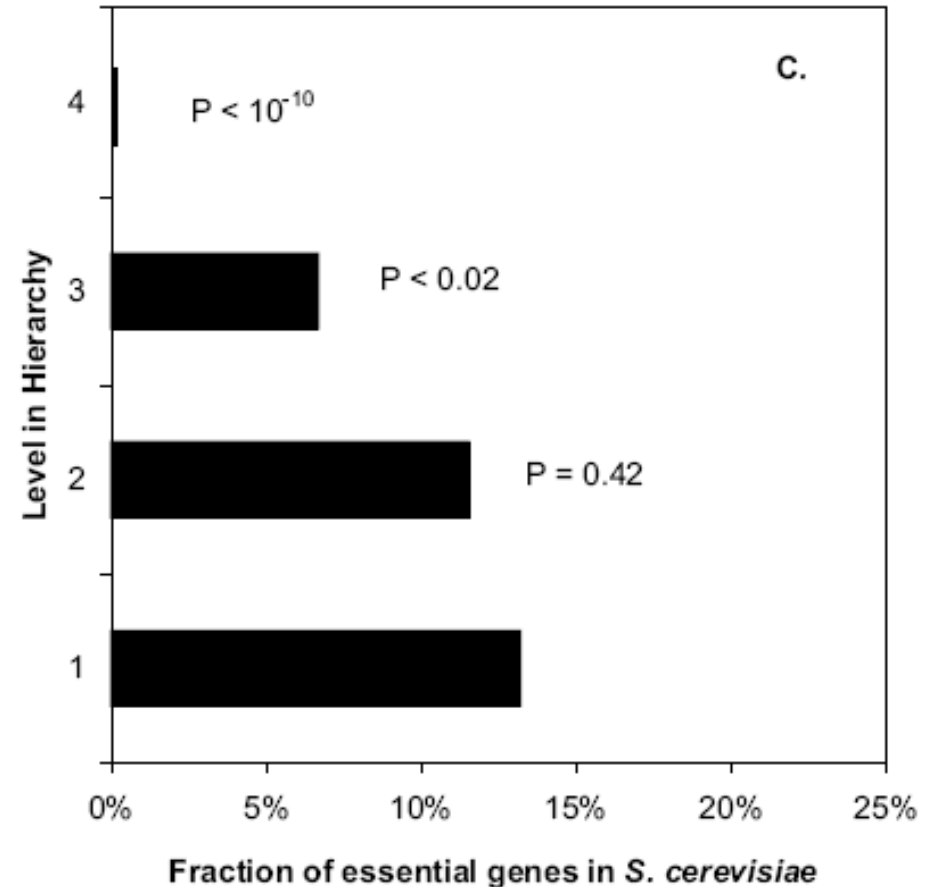
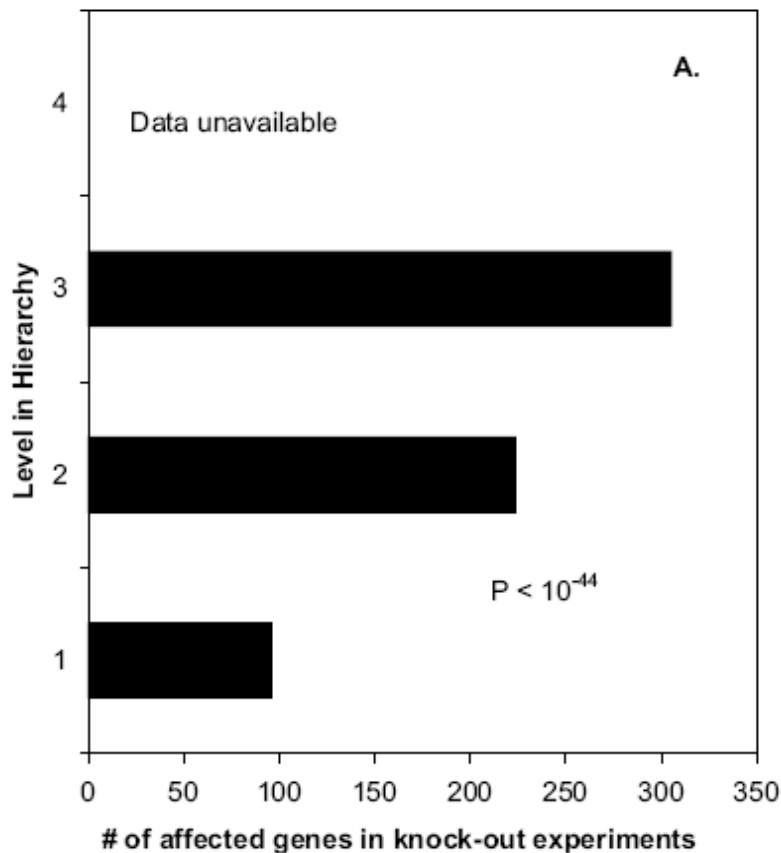
Characteristics of Regulatory Hierarchy: Middle Managers are Information Flow Bottlenecks

Average betweenness at each level



[Yu et al., PNAS (2006)]

Characteristics of Regulatory Hierarchy: The Paradox of Influence and Essentiality



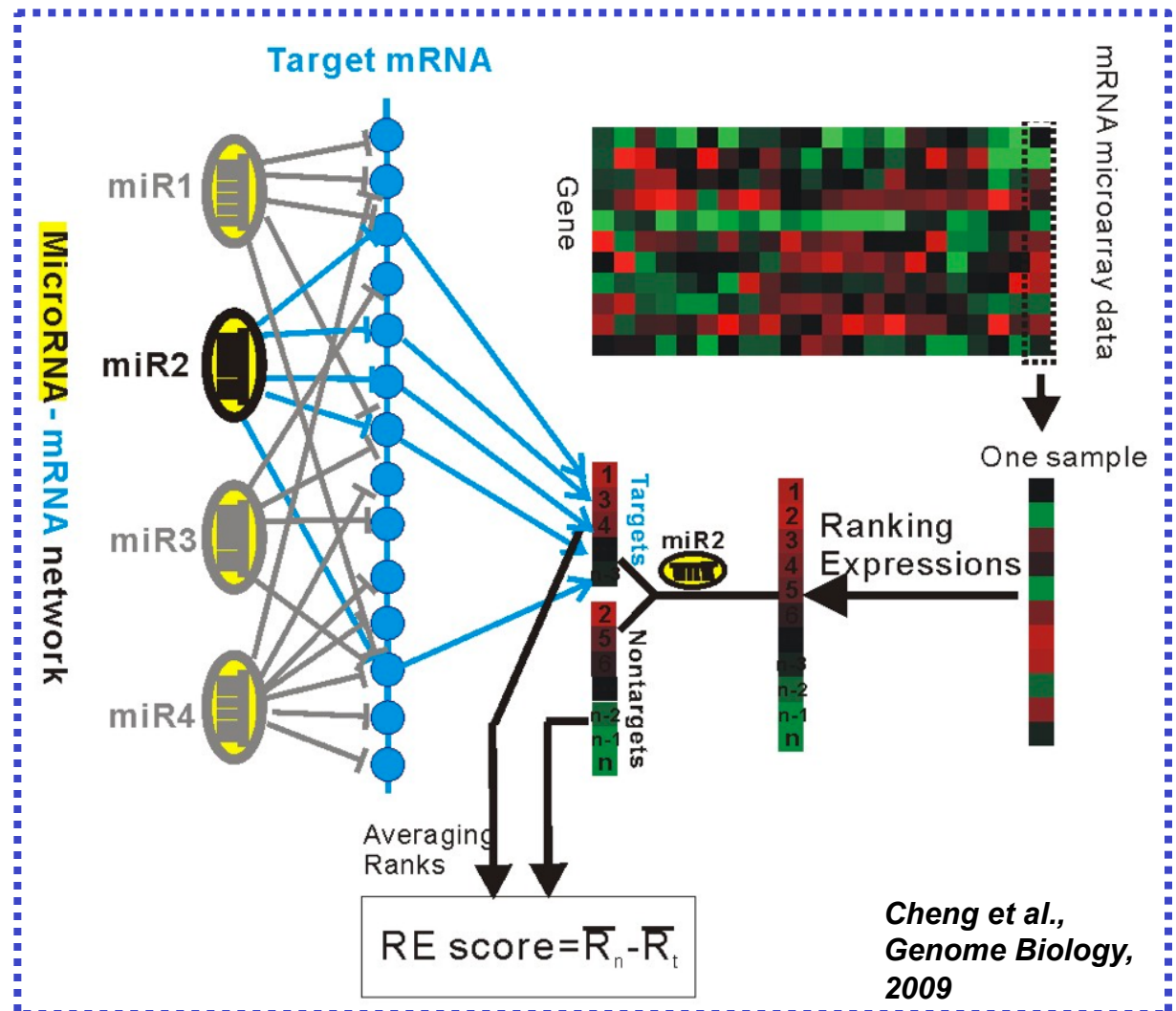
[Yu et al., PNAS (2006)]

Finding Central Points in Networks #3: Points of Maximal Regulatory Effect

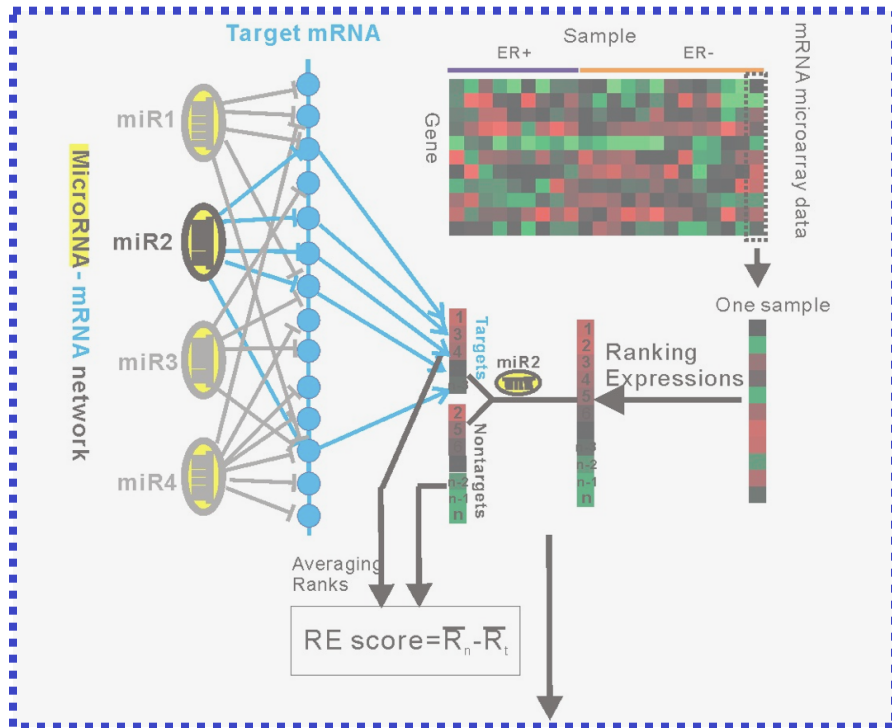


- How much does a regulator influence its targets?
- For micro-RNA-target networks easy to calculate, as all influence is down-regulation
 - ◇ target prediction methods: TargetScan, PITA, PicTar, miRanda, ...
- Look at down-reg. genes in a sample & compare with targets of a specific micro-RNA
 - ◇ more down-reg genes => stronger regulatory effect

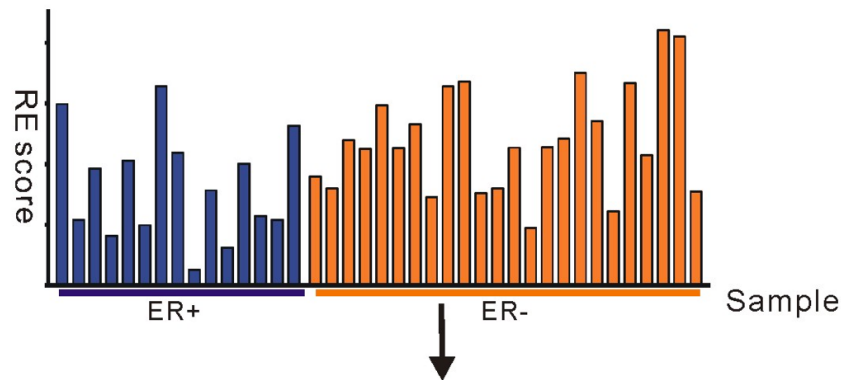
RE-score: Another way to measure "importance" in networks



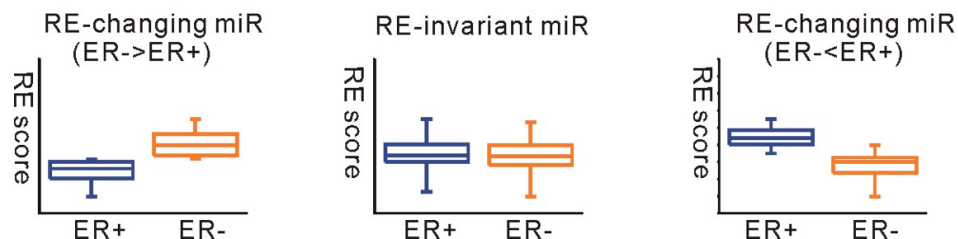
Application of RE-score to measure changing miRNA effect in different conditions (ER- and ER+ breast cancer)



Calculating RE scores of a miRNA in each sample

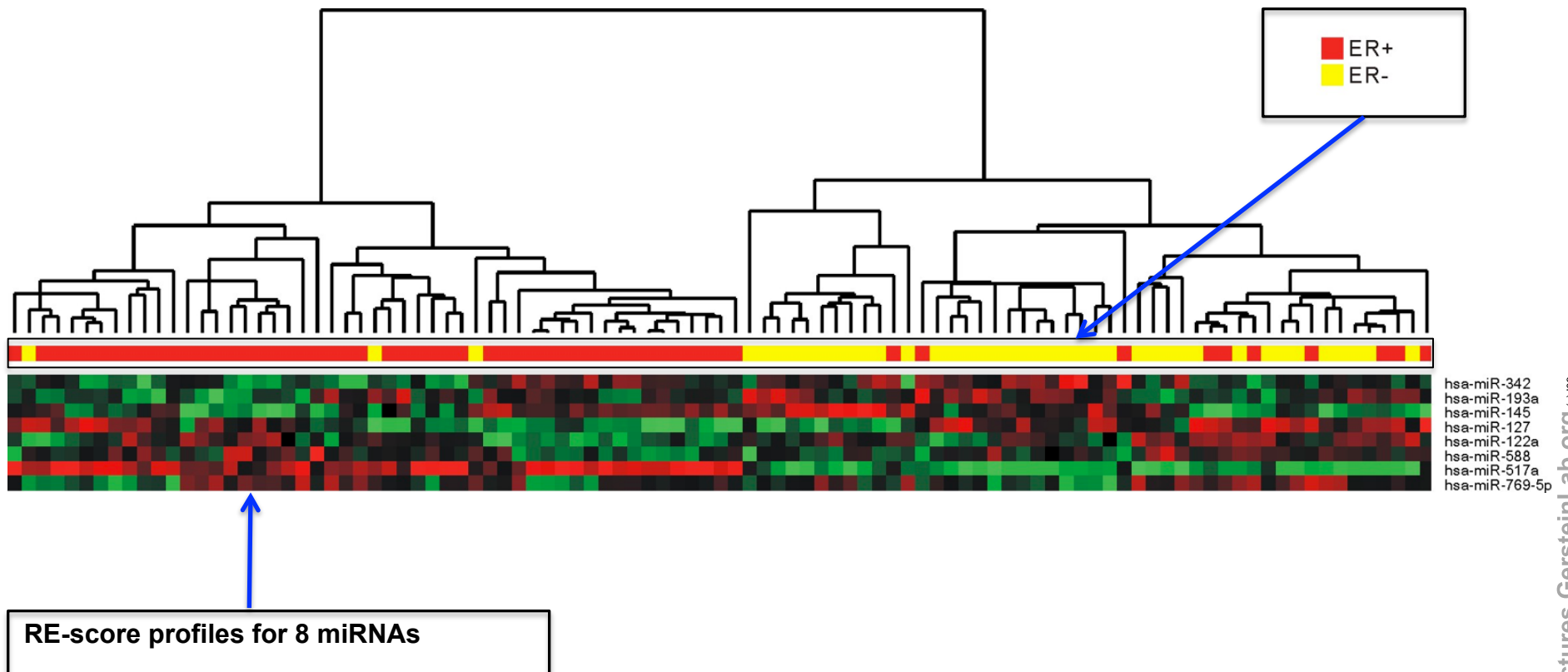


Comparing the RE scores between ER+ and ER-



Cheng et al., Genome Biology, 2009

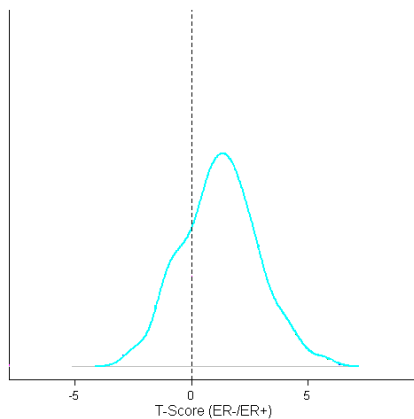
miRNA RE-scores can be used to classify cancers



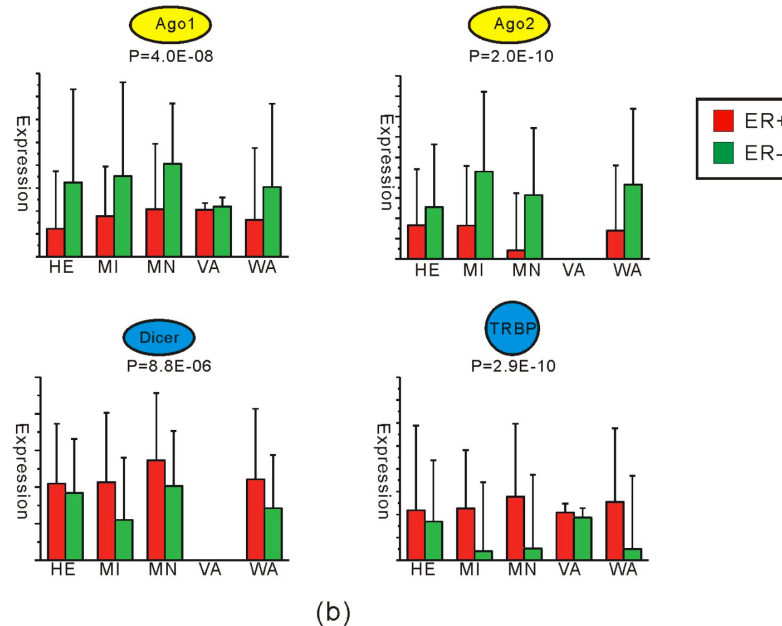
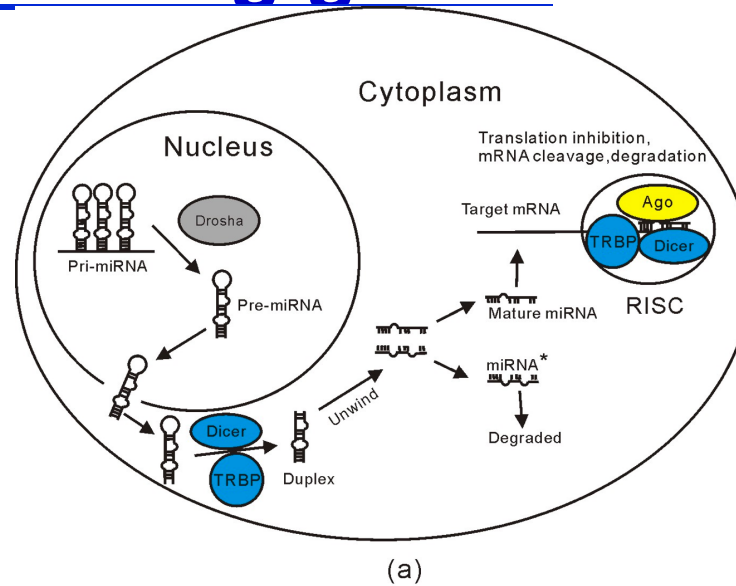
Cheng et al., *Genome Biology*,
2009

Differential expression of miRNA processing genes

Distribution of ER-/ER+ T-scores for all miRNAs



The majority of miRNAs have higher RE-score in ER- than in ER+



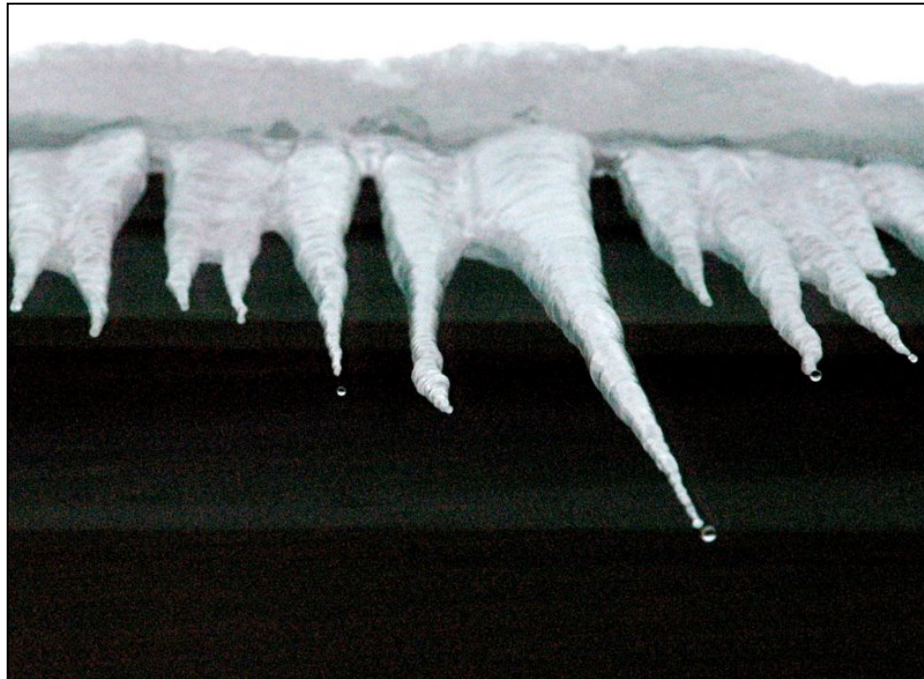
Network Dynamics #2:

Environments

How do molecular networks change across environments?

What pathways are used more ?

Used as a biosensor ?



What is metagenomics?

Genomics Approach

Culture Microbes



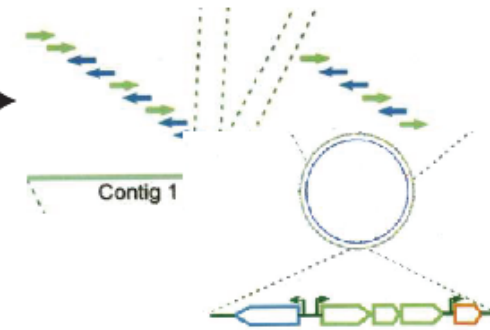
Extract DNA



Sequence

```
ATCGTATA
CGCGAAG
ACGTCTGA
AGTGCTGCT
```

Assemble and Annotate



PROBLEM: Estimated that less than 1% can be cultured in the lab

Metagenomics Approach

Collect Sample



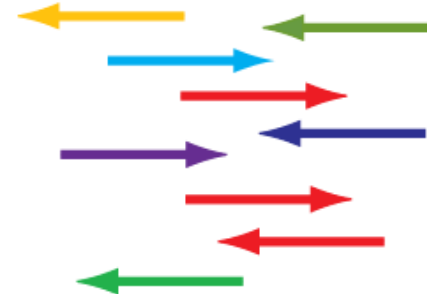
Extract DNA



Sequence

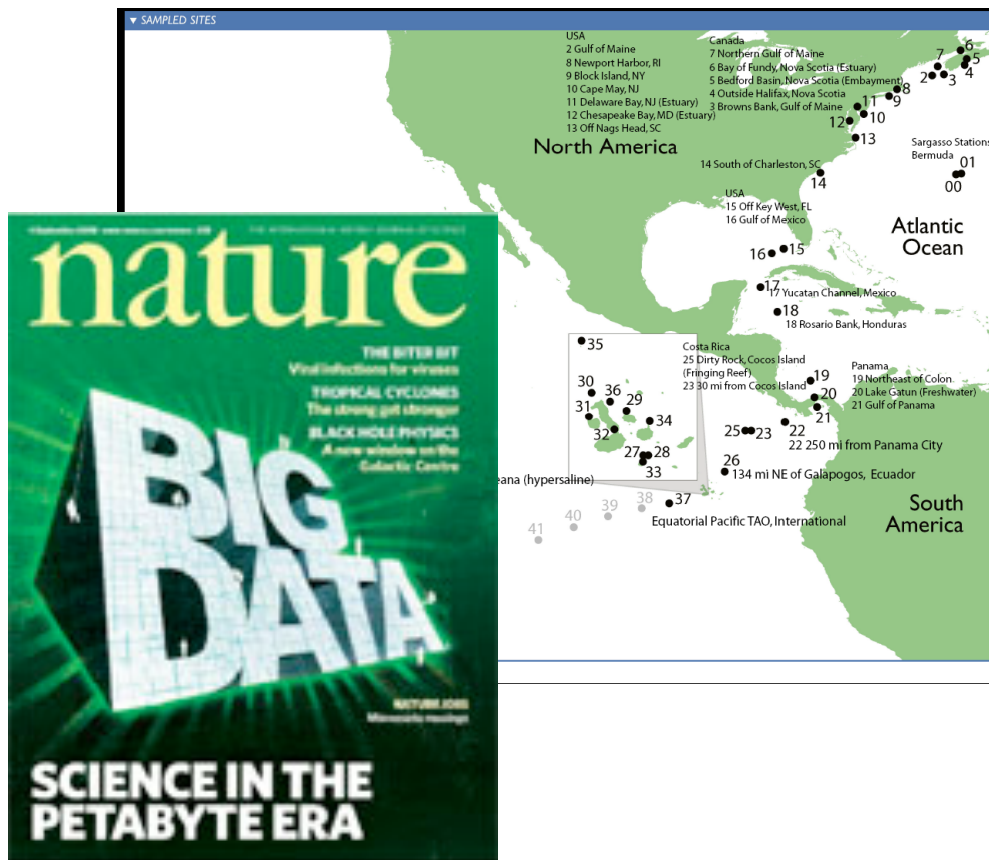
```
ATCGTGATAGATGATAGTAGA
ATGCTGCATGCATCTAGCACT
ACAGTAGCTAGCTACGTAATA
CAGCTGACTAGCTAGCTAGCT
ACGTAGCATGCTAGCTAGCAG
ACGTACGTAGCTAGCTAGTAG
ACGTACGTACGTAGCTAGCATC
AGTCGACTGAGCCAGTGATGAT
ACGATGCATGAGCAGATGCTAC
AGATCGTAGCATGCTAGCATGCT
ACGTACGTAGCTAGCTAGCTAAG
AGCTAGCATGCTAGTAGCATGAG
ACGATGCTAGCTAGCTAGCTGATA
TCGATCAGCATGCTACGATGCAAG
ACGATCGATGCTAGCTAGCAT
AGCTAGCTAGTCAGCTAGCTAGTG
```

Partially Assemble and Annotate



PROBLEM: Lose information about which gene belongs to which microbe.

Global Ocean Survey Statistics (GOS)



6.25 GB of data
7.7M Reads
1 million CPU hours
to process

Rusch, et al., PLOS Biology 2007

Pathway Sequences (Community Function)

Environmental Features

Metabolic Pathways

	P1	P2	P3		
Sites B1	3800	1400	1000		
B2	2200	100	400		
↓	---	---	---		



Environmental Metadata

	Temp	NaCl	Depth		
Sites B1	15°C	27.2	10 m		
B2	23°C	36.6	5 m		
↓	---	---	---		

READS → PROTEIN FAMILIES → PATHWAYS

CCGTGAGCACGATGCGC-----
 ATGCTCATGCT-----
 ATCGTGACGCGATGC-----
 CCGTGAGCACGATGCGC-----
 ATGCTCATGCT-----
 ATCGTGACGCGATGC-----
 ATGCTCATGCT-----
 GCGATCGATCGATCGTAGC-----
 TGCTGCTAGCATGCT-----
 GCGATCGATCGATCGTAGC-----
 TGCTGCTAGCATGCT-----
 CCGTGAGCACGATGCGC-----
 GTATCGTAGCATGCTT-----
 CCGTGAGCACGATGCGC-----
 GCGATCGATCGATCGTAGC-----



$$P_1 = f_1 + f_2 + f_3$$

$$P_2 = f_4 + f_5 + f_6$$

PATHWAYS

SITES

$$P_{1,1} = 2 + 1 + 3$$

$$P_{2,1} = 2 + 4 + 3$$

$$P_{1,2} = 5 + 2 + 6$$

$$P_{2,1} = 5 + 7 + 6$$

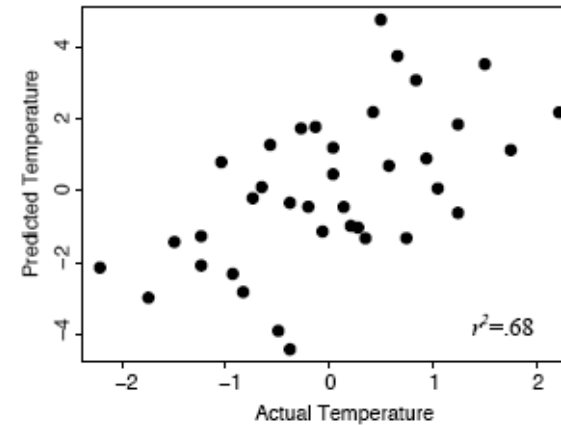
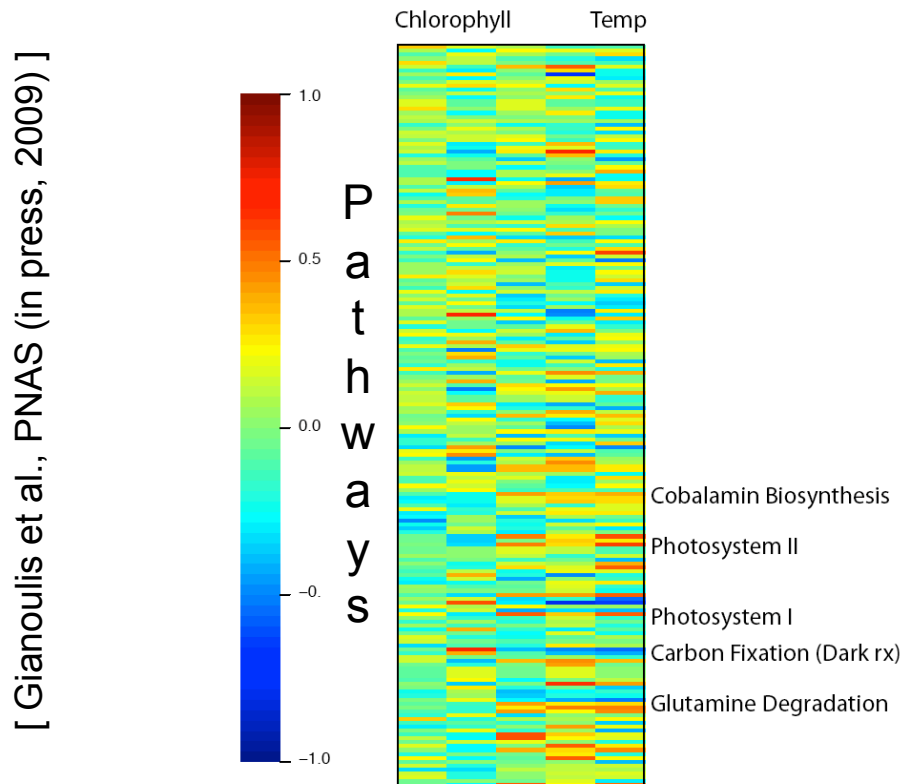
Expressing
data as
matrices
indexed by
site, env. var.,
and pathway
usage

[Rusch et. al., (2007) PLOS Biology;
 Gianoulis et al., PNAS (in press, 2009)]

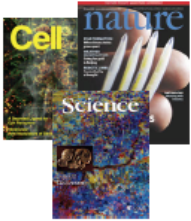
Simple Relationships: Pairwise Correlations





Environmental Features






Canonical Correlation Analysis: Simultaneous weighting

Score	# of papers published
GRE	

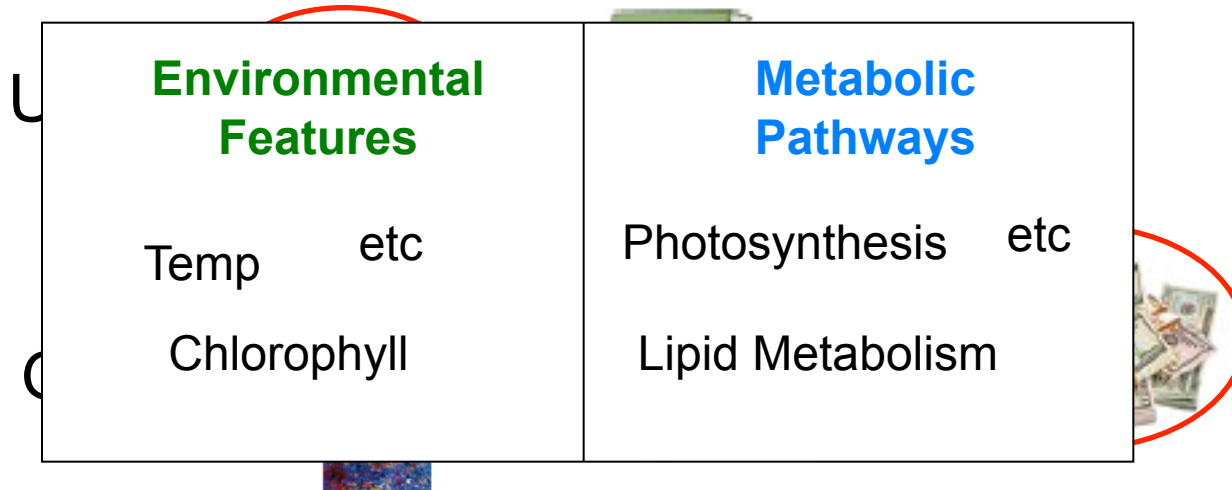
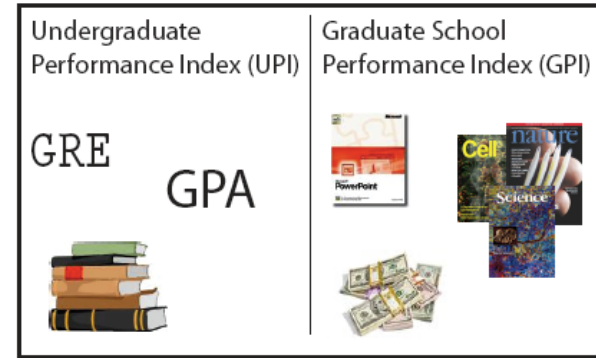
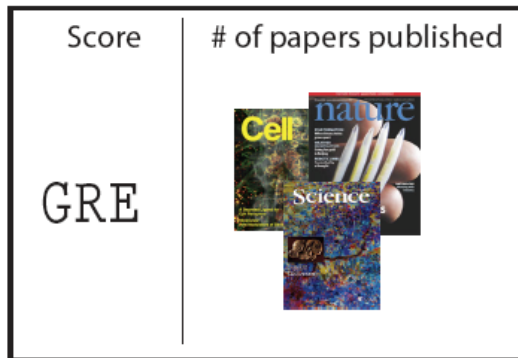
Undergraduate Performance Index (UPI)	Graduate School Performance Index (GPI)
GRE 	

$$\text{UPI} = a \text{ GRE} + b \text{ GPA}$$

$$\text{GPI} = a' \text{  + b' \text{  + c' \text{ $$

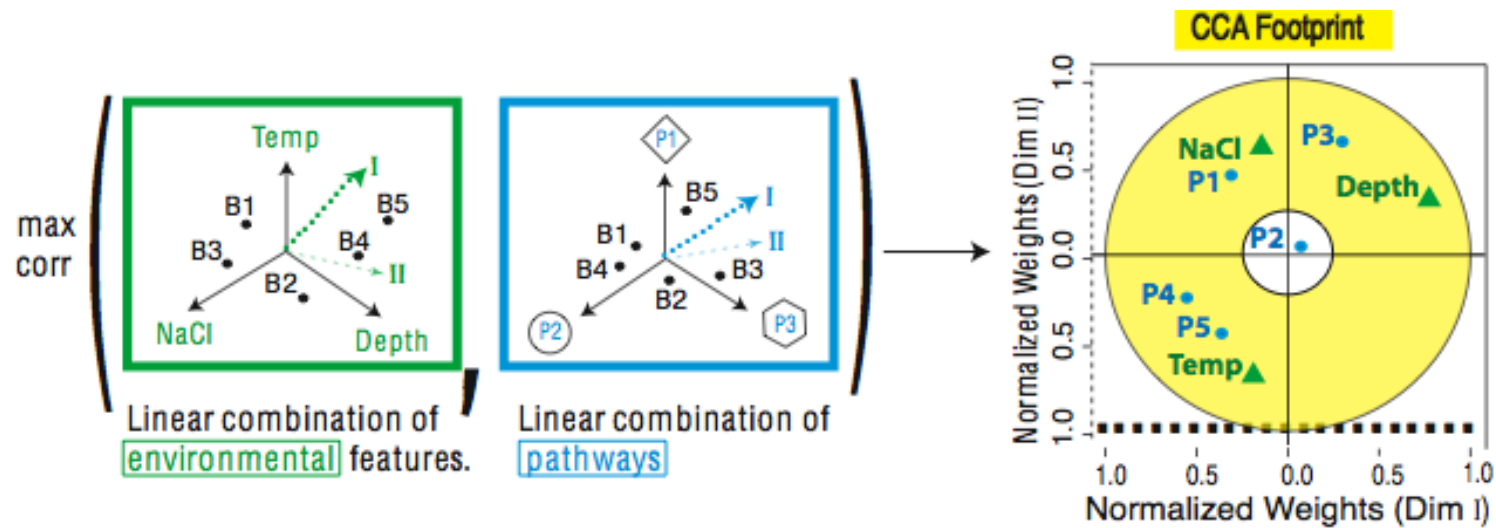
[Gianoulis et al., PNAS (in press, 2009)]

Canonical Correlation Analysis: Simultaneous weighting



[Gianoulis et al., PNAS (in press, 2009)]

Environmental-Metabolic Space

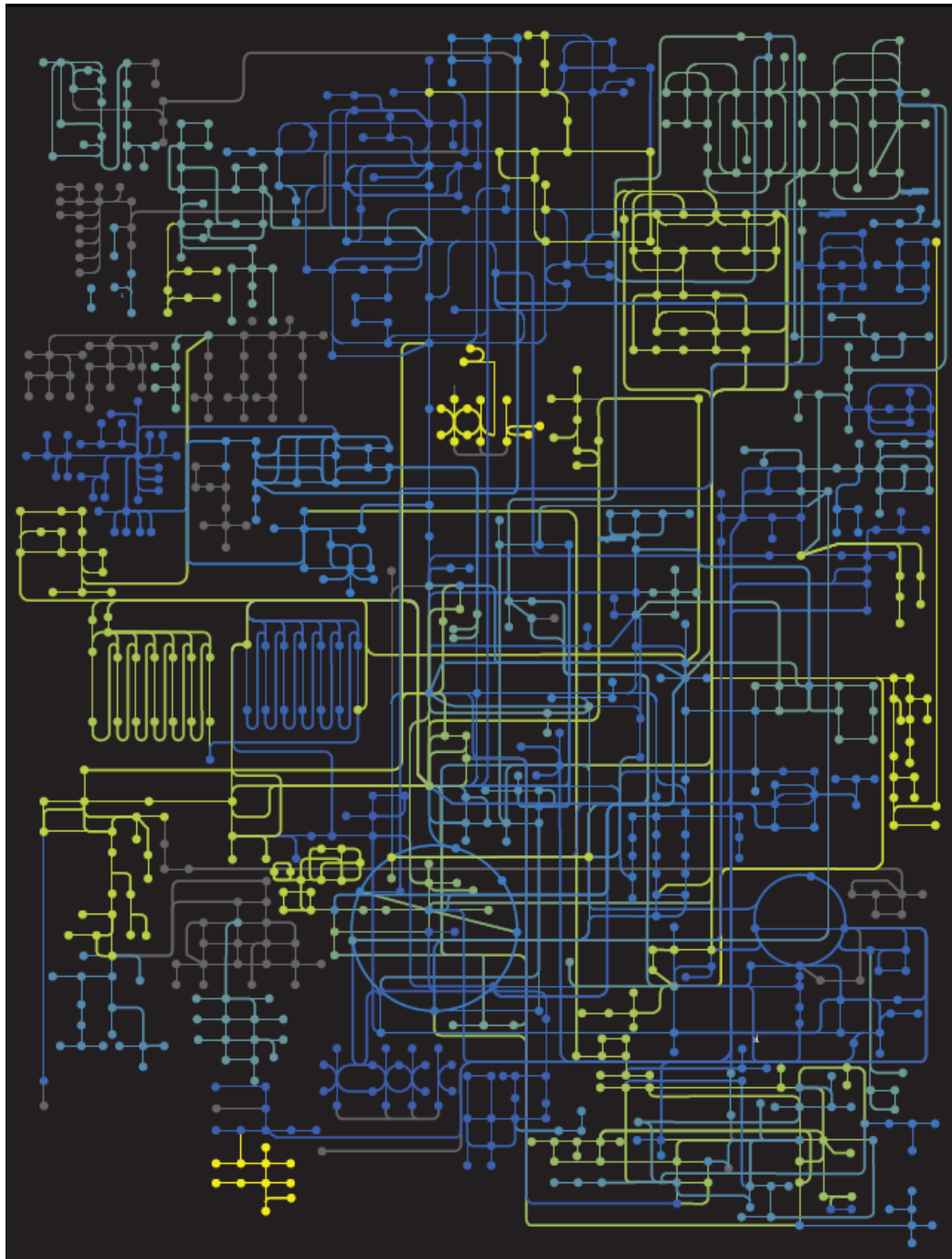


The goal of this technique is to interpret cross-variance matrices
We do this by defining a change of basis.

Given $X = \{x_1, x_2, \dots, x_n\}$ and $Y = \{y_1, y_2, \dots, y_m\}$

$$C = \begin{matrix} \sum_X & \sum_{X,Y} \\ \sum_Y & \sum_{Y,X} \end{matrix} \quad \max_{a,b} \text{Corr}(U,V) = \frac{a' \sum_{12} b}{\sqrt{a' \sum_{11} a} \sqrt{b' \sum_{22} b}}$$

[Gianoulis et al., PNAS (in press, 2009)]

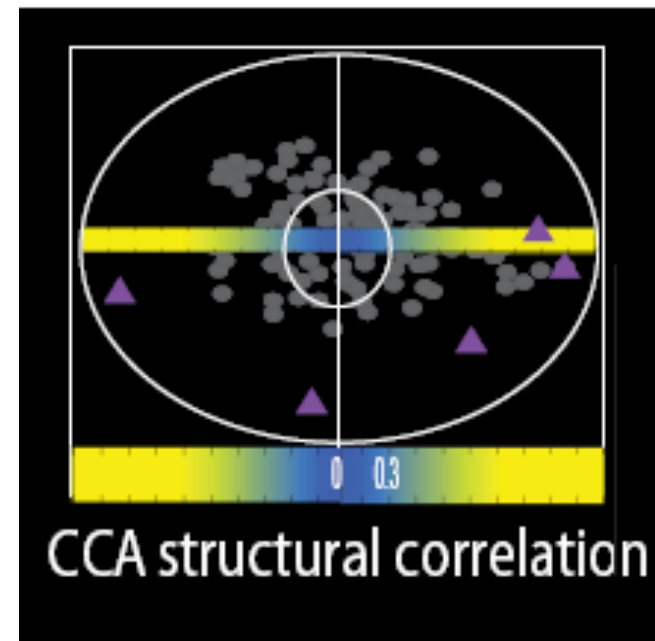


Strength of Pathway co-variation with environment



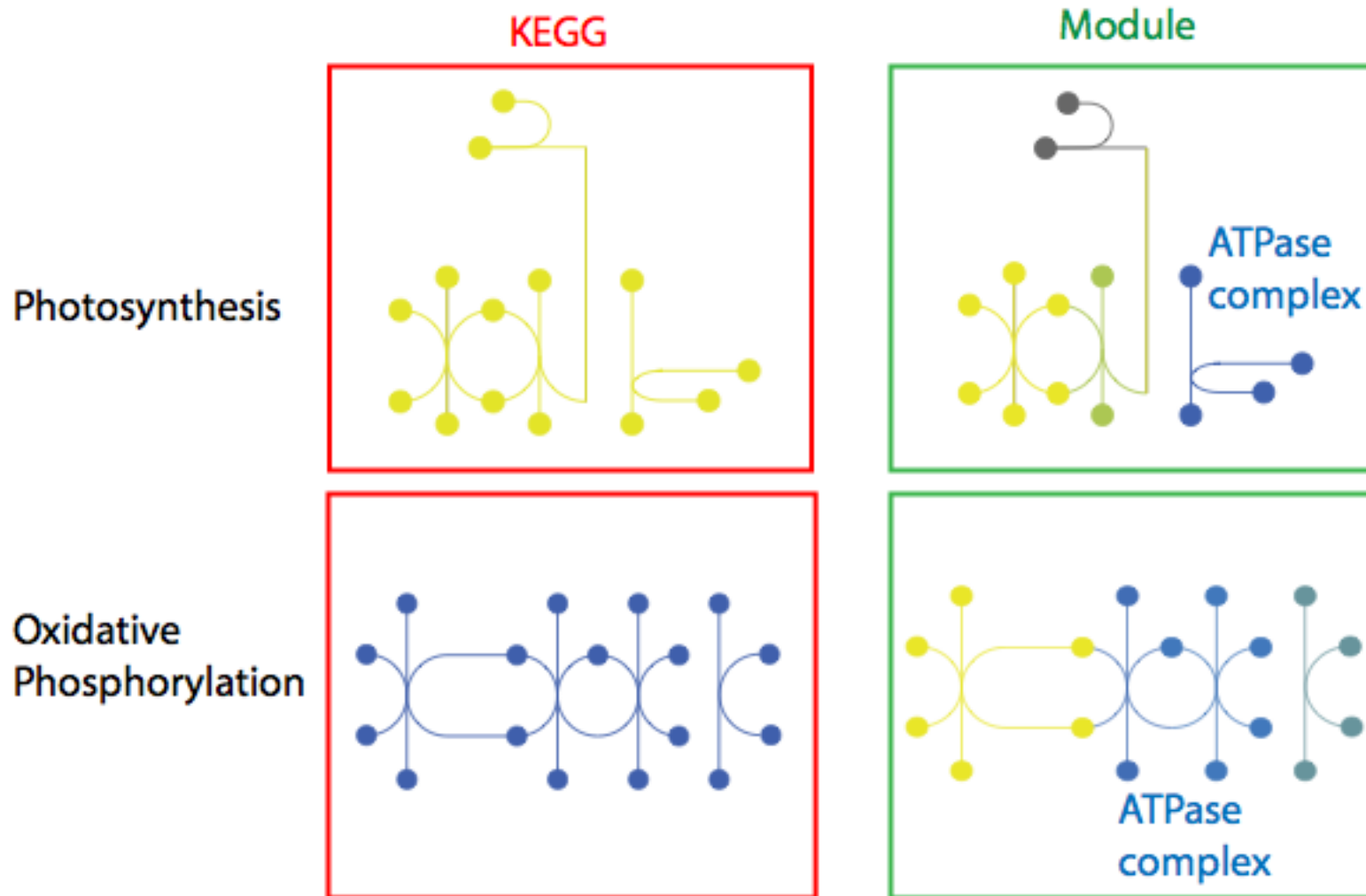
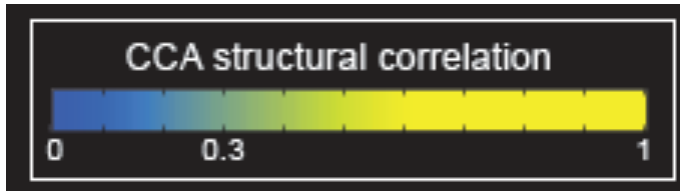
Environmentally
invariant

Environmentally
variant

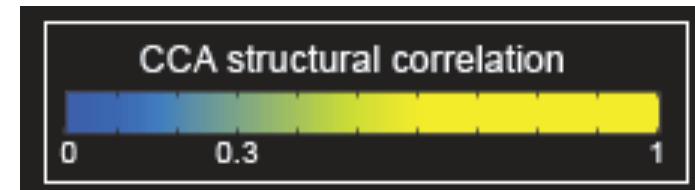
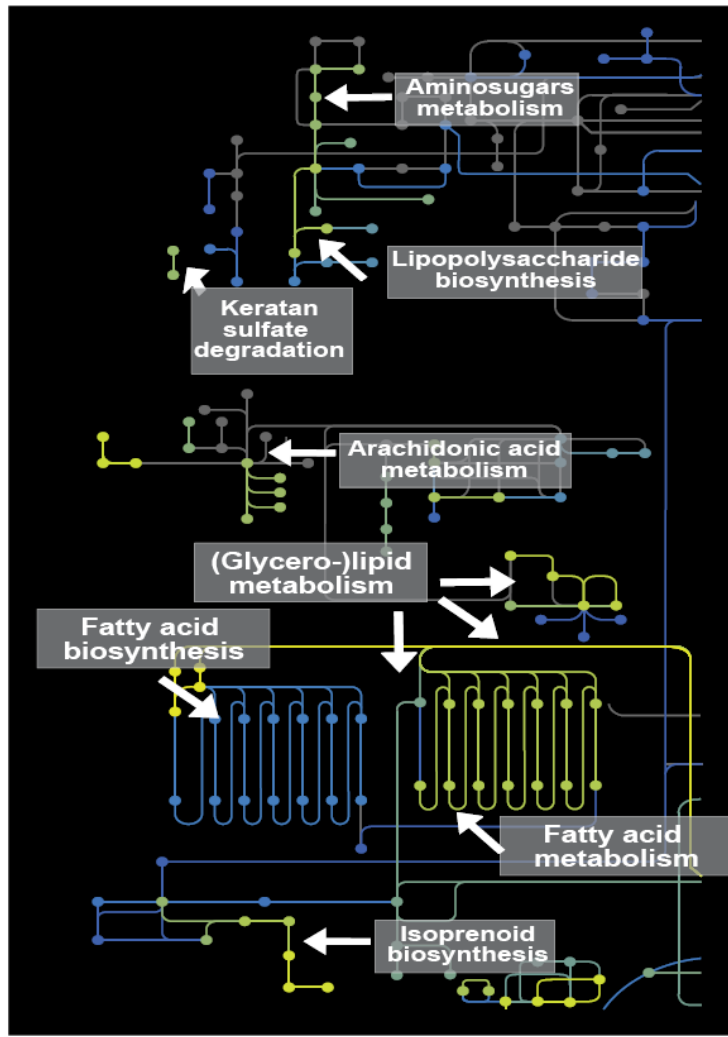


[Gianoulis et al., PNAS (in press, 2009)]

Conclusion #1: energy conversion strategy, temp and depth

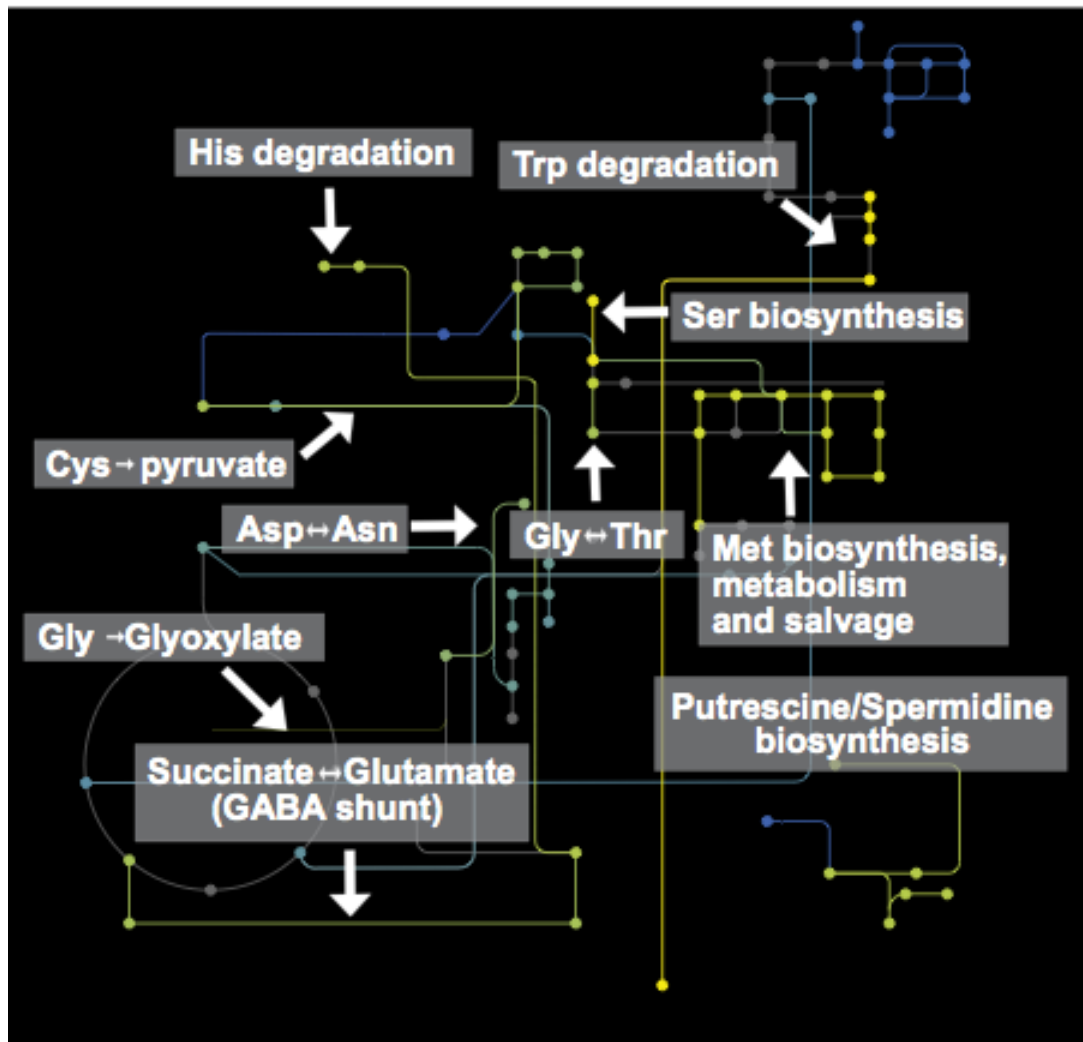


Conclusion #2: Outer Membrane components vary the environment



[Gianoulis et al., PNAS (in press, 2009)]

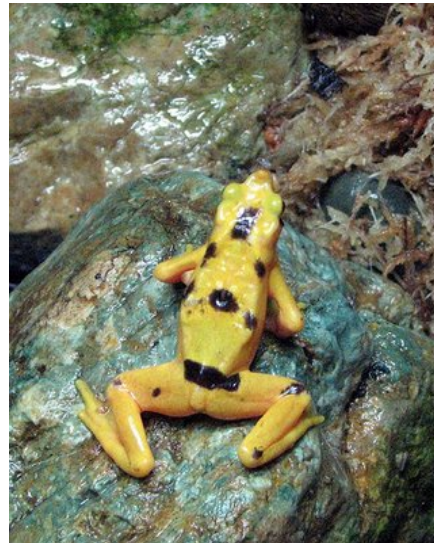
Conclusion #3: Covariation of AA biosynthesis and Import



Why is their fluctuation in amino acid metabolism? Is there a feature(s) that underlies those that are environmentally-variant as opposed to those which are not?

[Gianoulis et al., PNAS (in press, 2009)]

Biosensors: Beyond Canaries in a Coal Mine



[Gianoulis et al., PNAS (in press, 2009)]

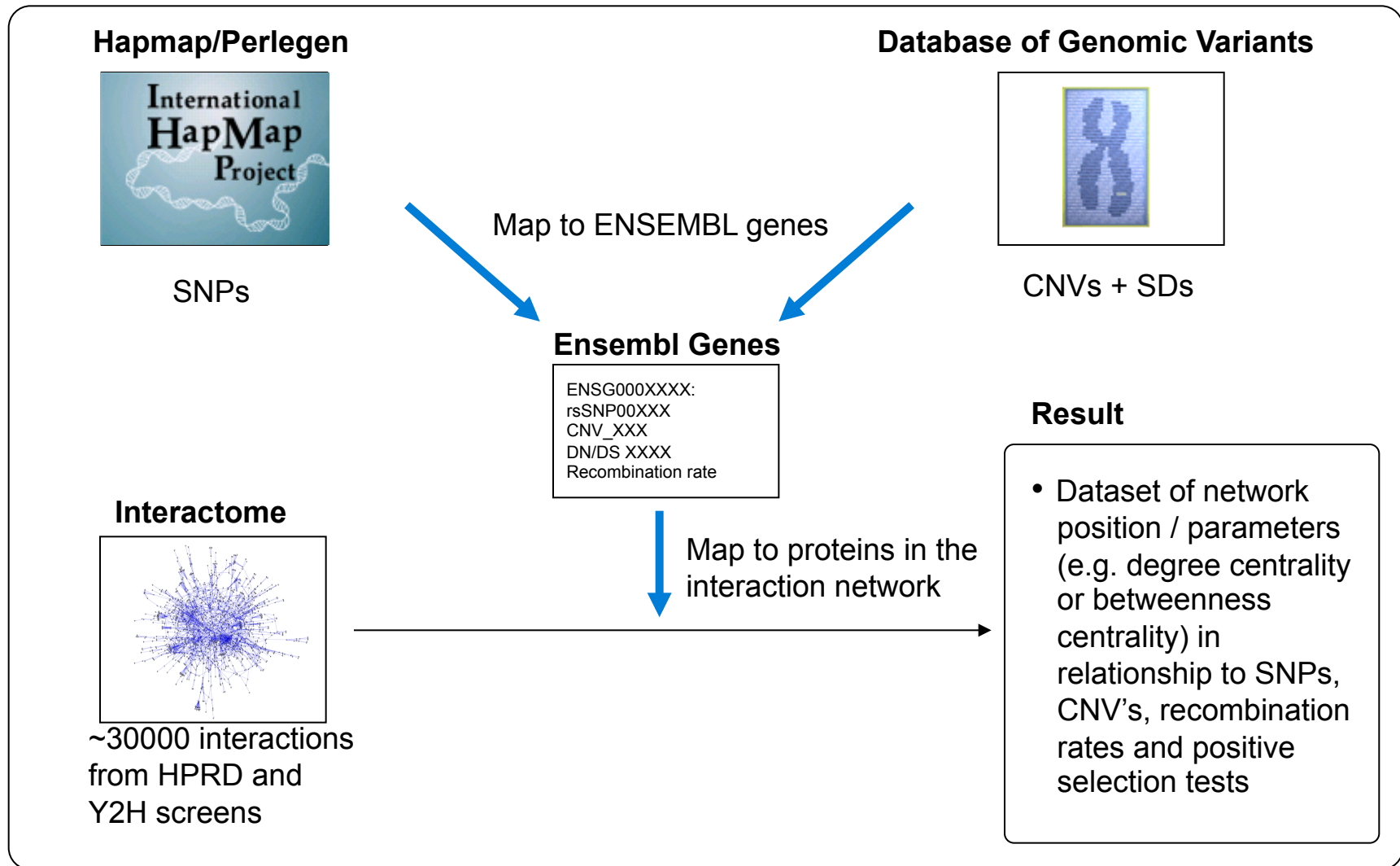
Networks & Variation

Which parts of the network vary most in sequence?
Which are under selection, either positive or negative?



METHODOLOGY: MAP SNP AND CNV DATA ONTO ENSEMBL GENES, AND THEN MAP ENSEMBL GENES TO THE KNOWN INTERACTOME

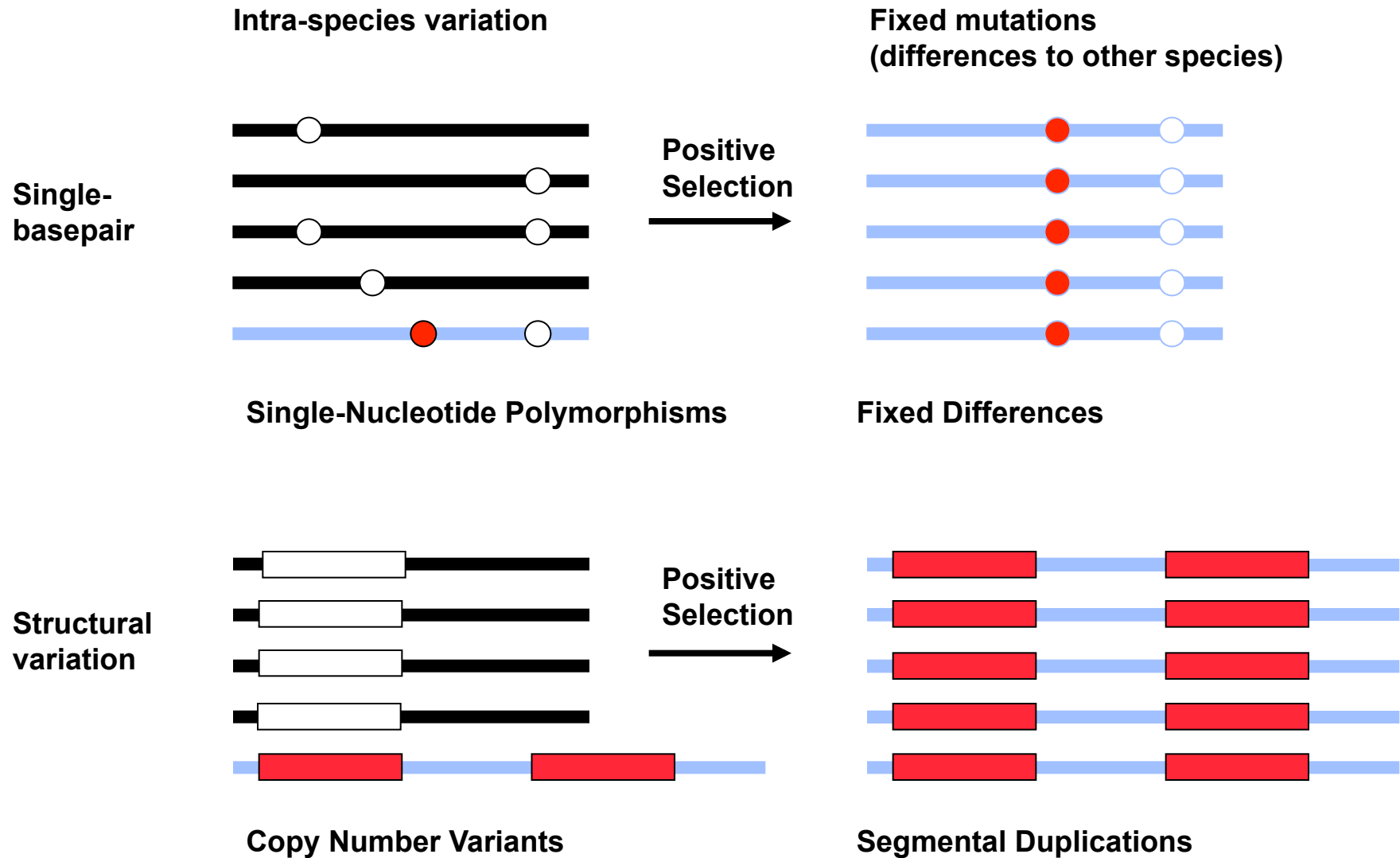
ILLUSTRATIVE



* From Nielsen et al. *PLoS Biol.* (2005) and Bustamante et al. *Nature* (2005)

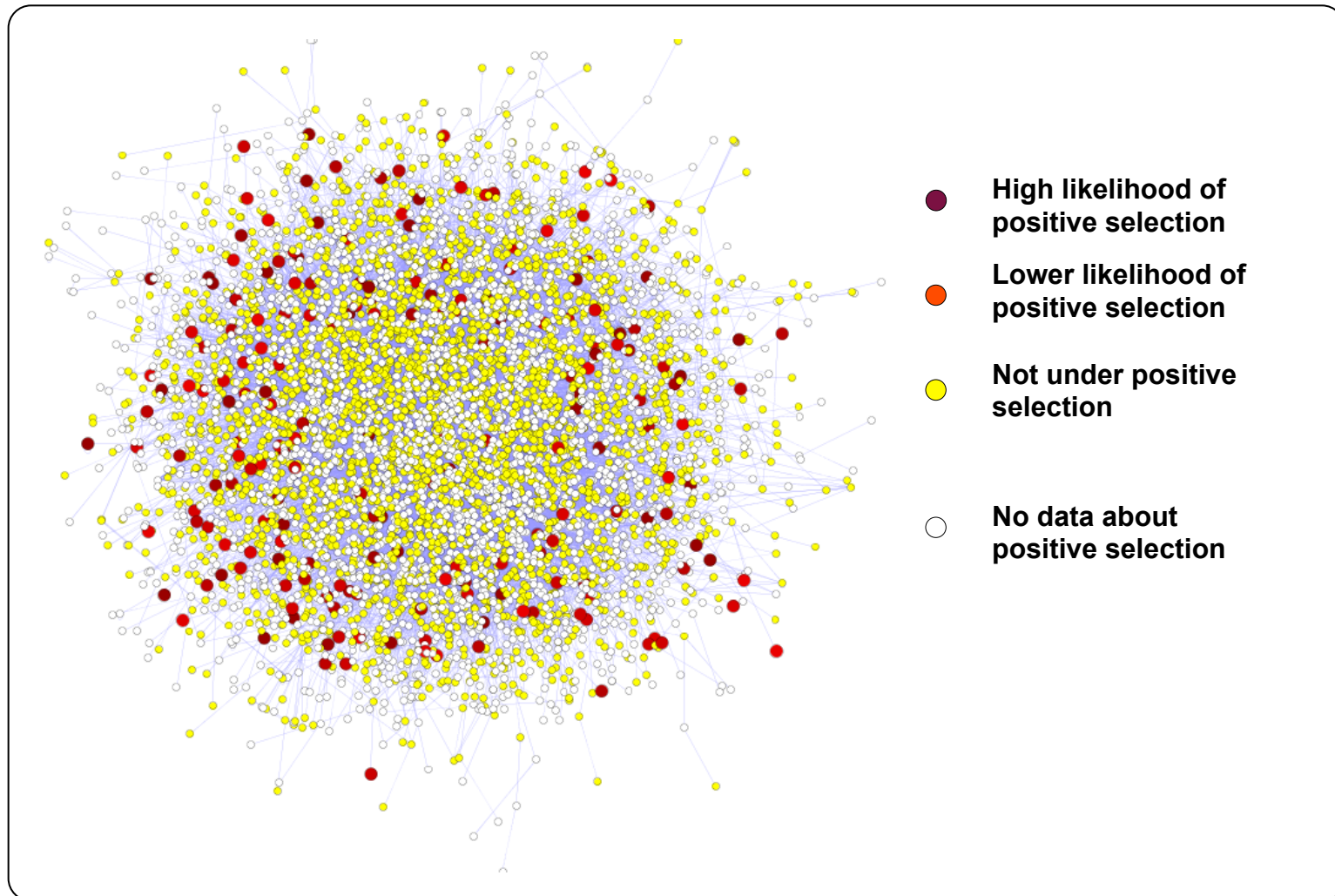
Source: PMK

ADAPTIVE EVOLUTION CAN BE SEEN ON TWO DIFFERENT LEVELS



POSITIVE SELECTION LARGELY TAKES PLACE AT THE NETWORK PERIPHERY

Positive selection in the human interactome

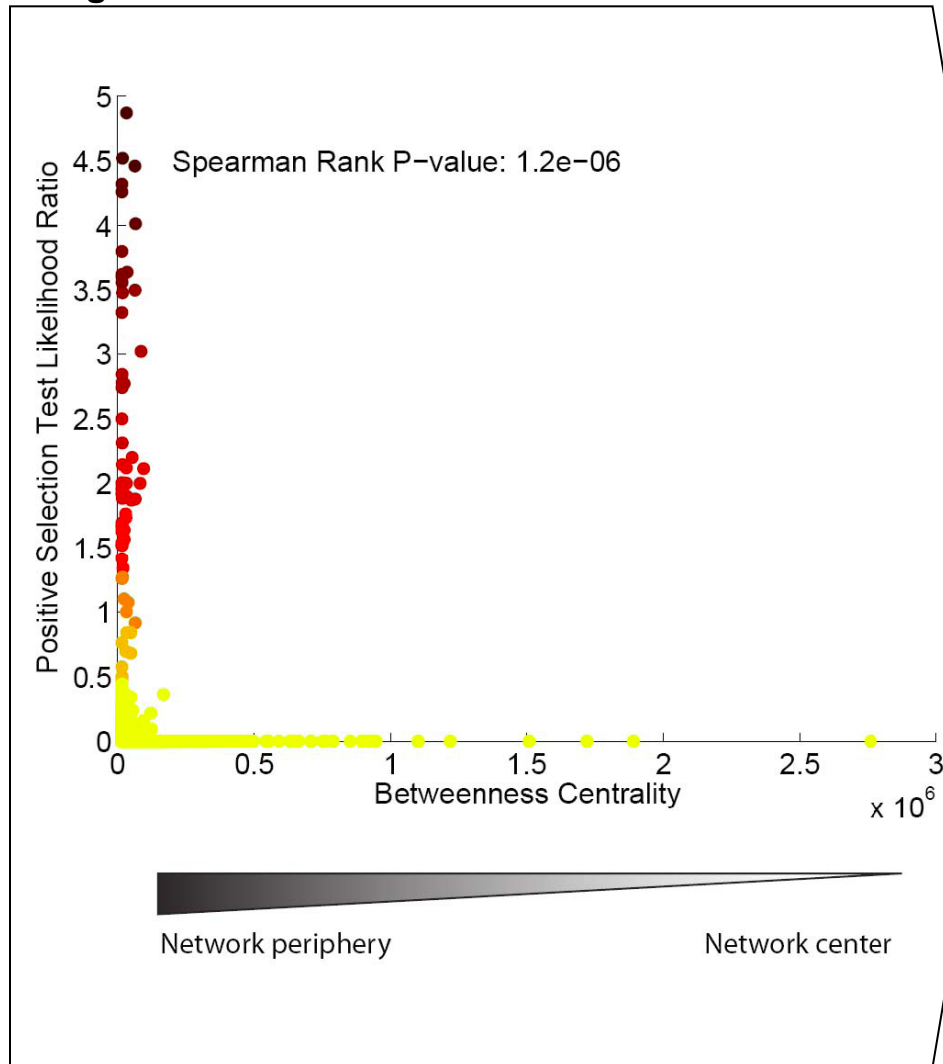


Source: Nielsen et al. *PLoS Biol.* (2005), HPRD, and Kim et al. *PNAS* (2007)

CENTRAL PROTEINS ARE LESS LIKELY TO BE UNDER POSITIVE SELECTION

▢ Hubs

Degree vs. Positive Selection



Reasoning

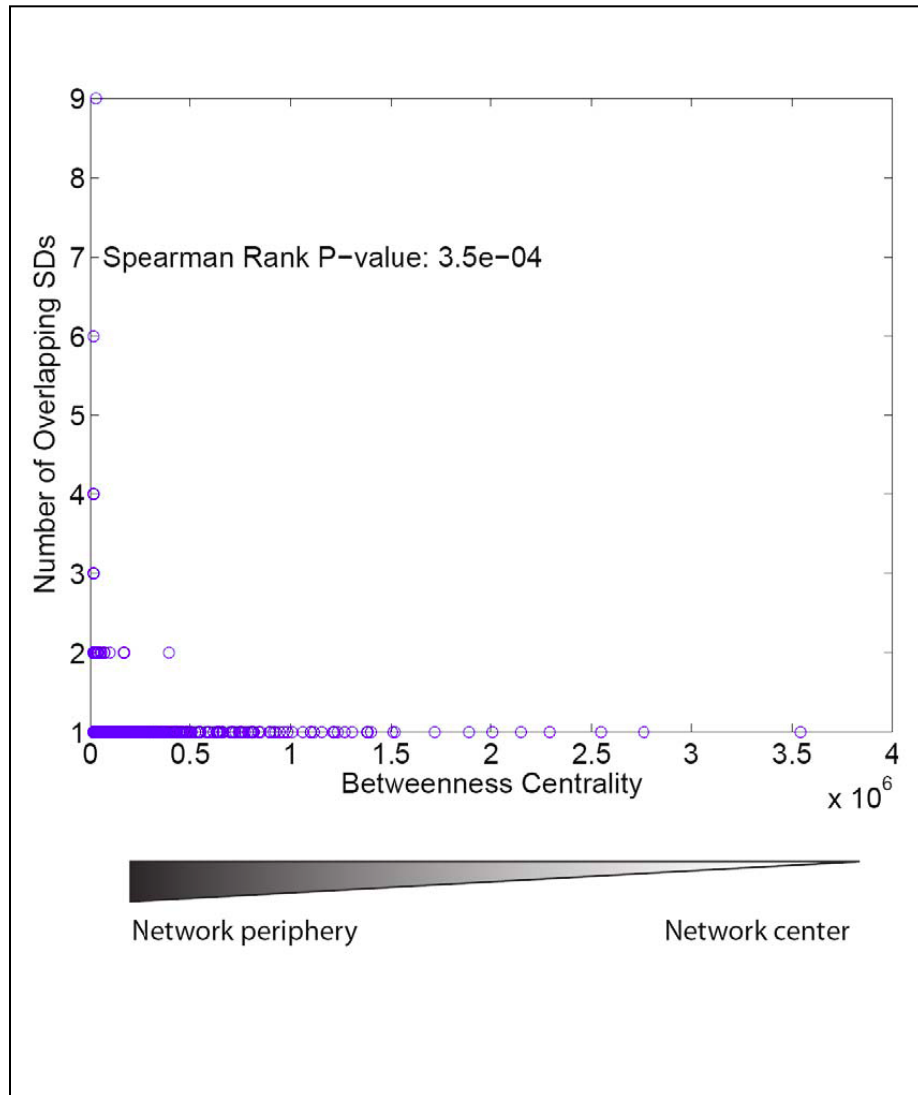
- Peripheral genes are likely to under positive selection, whereas hubs aren't
- This is likely due to the following reasons:
 - Hubs have stronger structural constraints, the network periphery doesn't
 - Most recently evolved functions (e.g. “environmental interaction genes” such as sensory perception genes etc.) would probably lie in the network periphery
- Effect is independent of any bias due to gene expression differences

* With a probability of over 80% to be positively selected as determined by Ka/Ks. Other tests of positive selection (McDonald Kreitmann and LDD) corroborate this result.

Source: Nielsen et al. *PLoS Biol.* (2005), Bustamante et al. *Nature* (2005), HPRD, Rual et al. *Nature* (2005), and Kim et al. *PNAS* (2007)

CENTRAL NODES ARE LESS LIKELY TO LIE INSIDE OF SDs

Centrality vs. SD occurrence



Reasoning

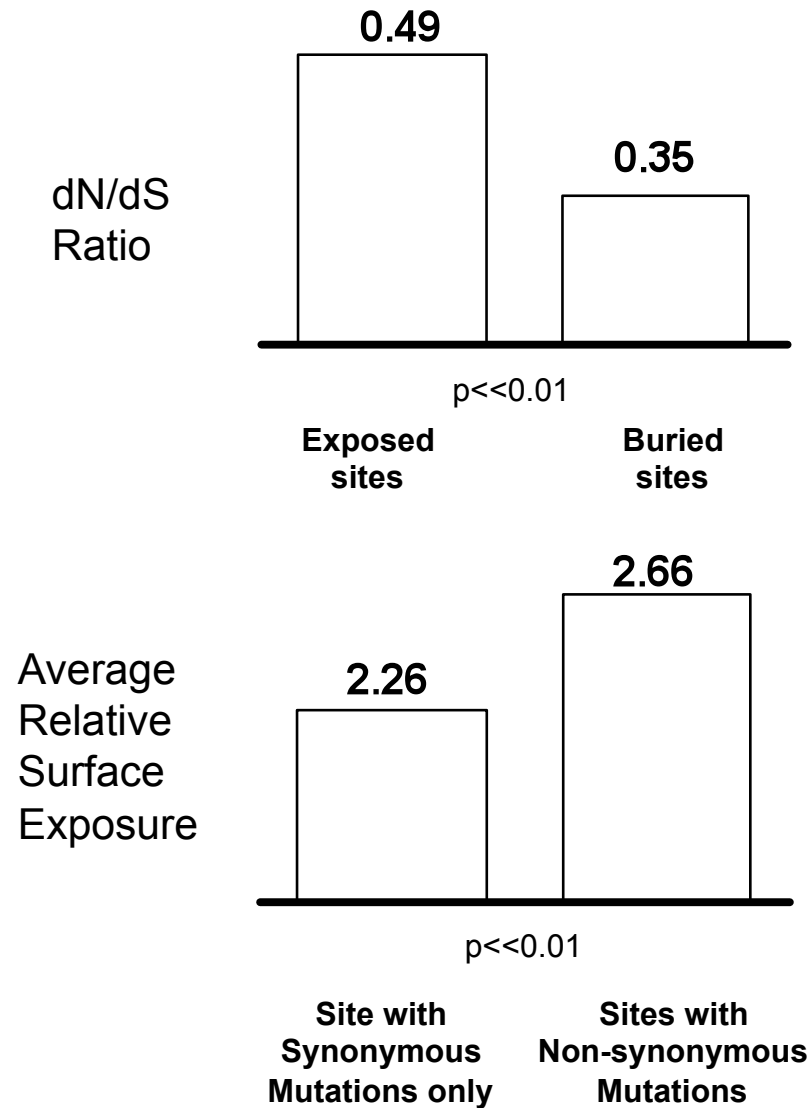
- This result also confirms our initial hypothesis – peripheral nodes tend to lie in regions rich in SDs.
- Since segmental duplications are a different mechanism of ongoing evolution, the less constrained peripheral proteins are enriched in them.
- Note that despite the small size of our dataset for known SD's we get significant correlations. It is to be expected that the correlations will get clearer as more data emerges*

* Specifically, a number of the SDs are likely not fixed, but rather common CNVs in the reference genome

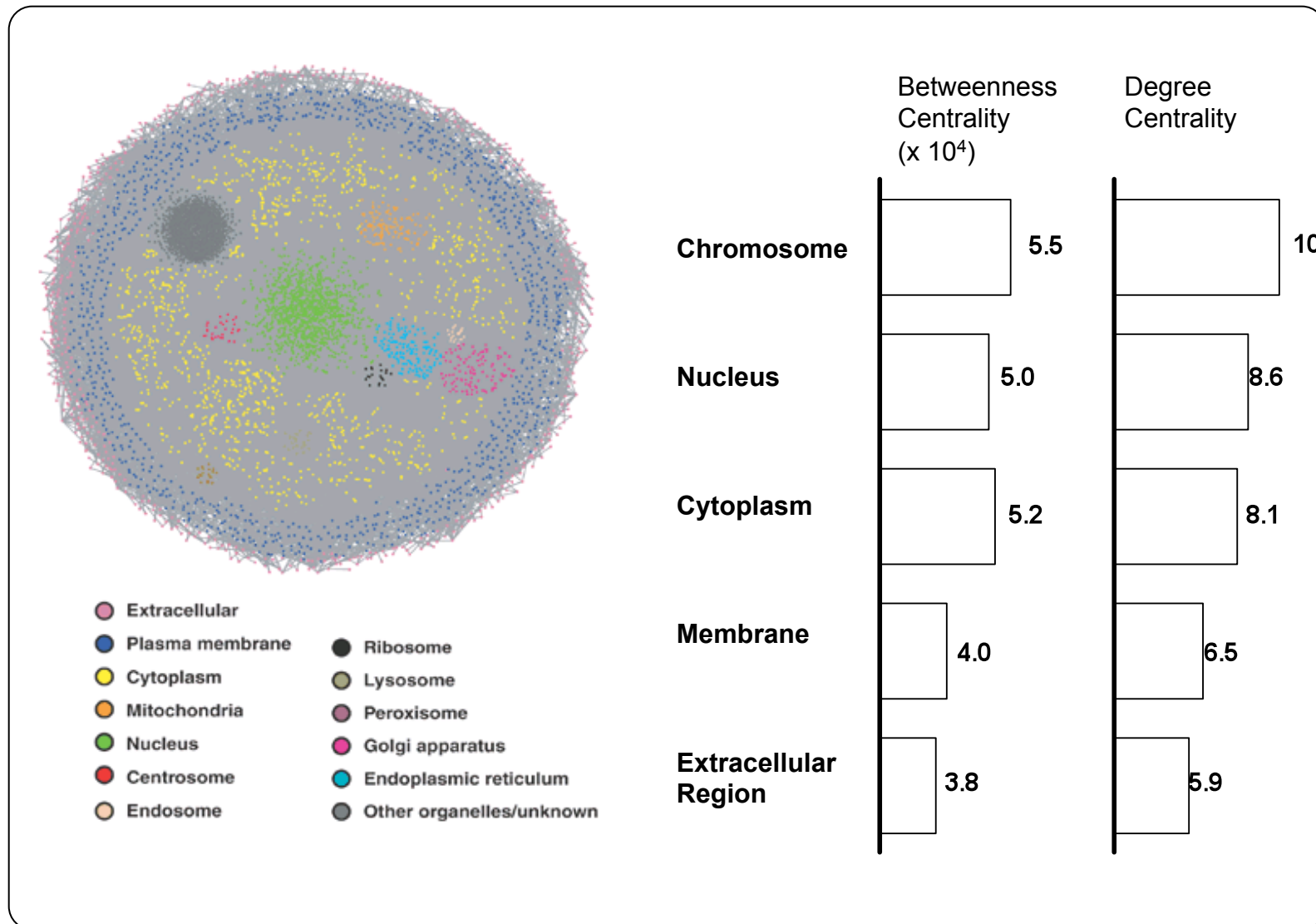
Source: Database of genetic variation, HPRD, Rual et al. *Nature* (2005), and Kim et al. *PNAS* (2007)

Why do we observe this? Perhaps central hub proteins are involved in more interactions & have more surface buried.

**BURIED SITES ARE
CONSERVED AND
MUCH LESS LIKELY
TO HARBOR NON-
SYNONYMOUS
MUTATIONS**



Another explanation: THE NETWORK PERIPHERY CORRESPONDS TO THE CELLULAR PERIPHERY



Source: Gandhi et al. (*Nature Genetics* 2006), Kim et al. PNAS (2007)

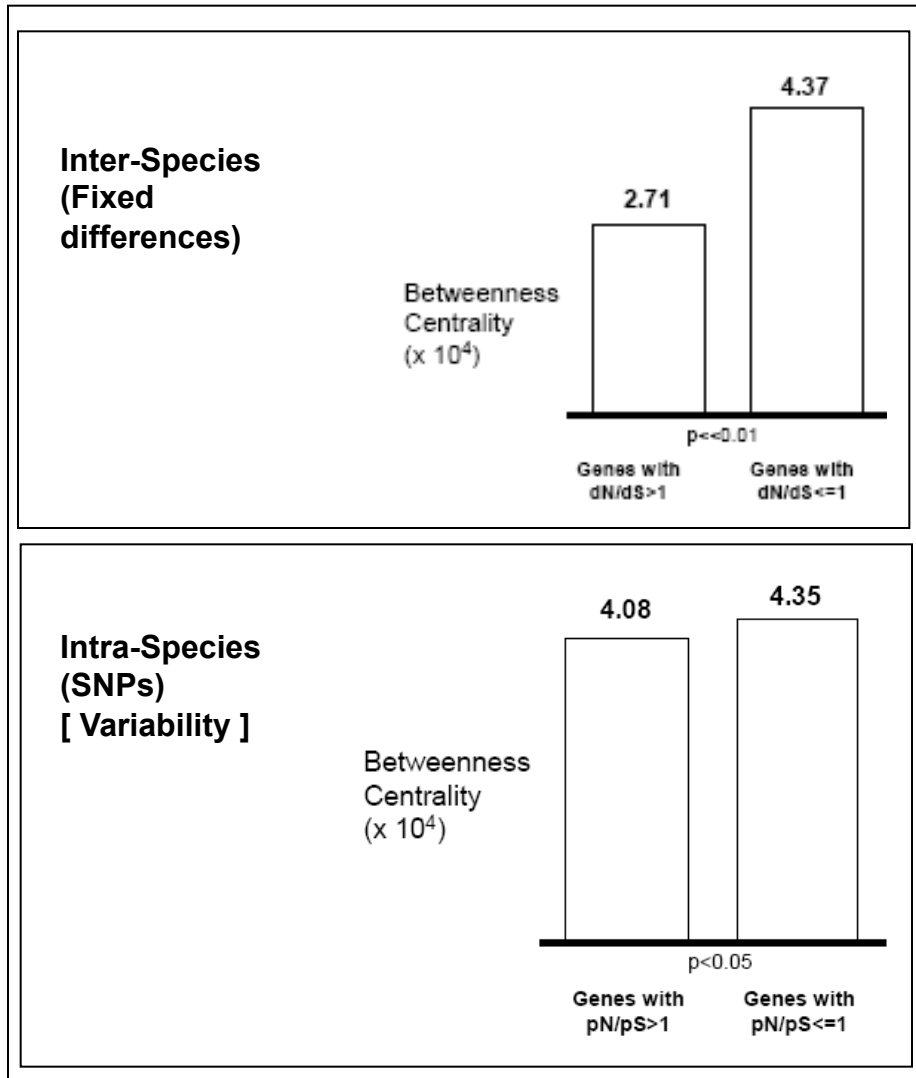
IS RELAXED CONSTRAINT OR ADAPTIVE EVOLUTION THE REASON FOR THE PREVALENCE OF BOTH SELECTED GENES AND SDs AT THE NETWORK PERIPHERY?

ILLUSTRATIVE

	Relaxed Constraint	Adaptive Evolution
Inter-Species Variation (Fixed differences)	<ul style="list-style-type: none">• Increases inter-species variation – more variable loci are under less negative selection• Can be seen in higher Ka/Ks ratio or SD occurrence	<ul style="list-style-type: none">• Increases inter-species variation – more variable loci are under less negative selection• Can be seen in higher Ka/Ks ratio or SD occurrence
Intra-Species Variation (Polymorphisms)	<ul style="list-style-type: none">• Increases intra-species variation – for the very same reason• Can be seen in both SNPs or CNVs	<ul style="list-style-type: none">• Should not have effects on intra-species variation

SOME, BUT NOT ALL OF THE SINGLE-BASEPAIR SELECTION AT THE PERIPHERY IS DUE TO RELAXED CONSTRAINT

Inter vs. Intra-Species Variation in Networks



Reasoning

- There is a difference in **variability** (in terms of SNPs) between the network periphery and the center
- However, this difference is much smaller than the difference in **selection**
- This most likely means, that part of the effect we're seeing is due to relaxed constraint (and higher variability)
- But, not the entire effect*

* But it's hard to quantify

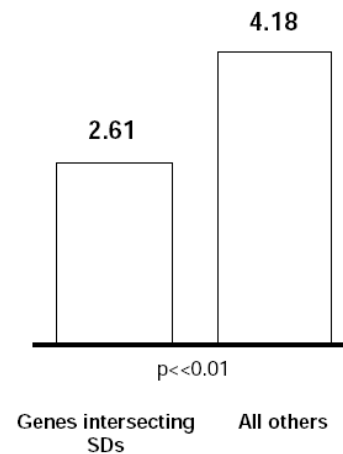
Source: Kim et al. (2007) PNAS

Similar Results for Large-scale Genomic Changes (CNVs and SDs)

Inter vs. Intra-Species Variation in Networks

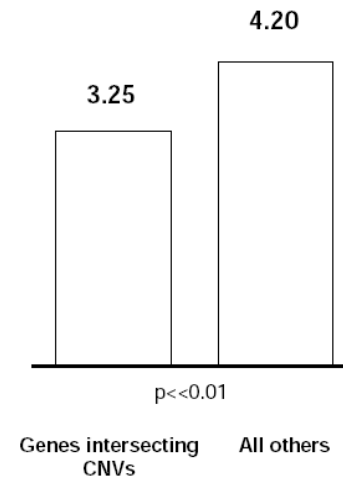
Inter-Species (SDs)

Betweenness Centrality ($\times 10^4$)



Intra-Species (CNVs) [Variability]

Betweenness Centrality ($\times 10^4$)

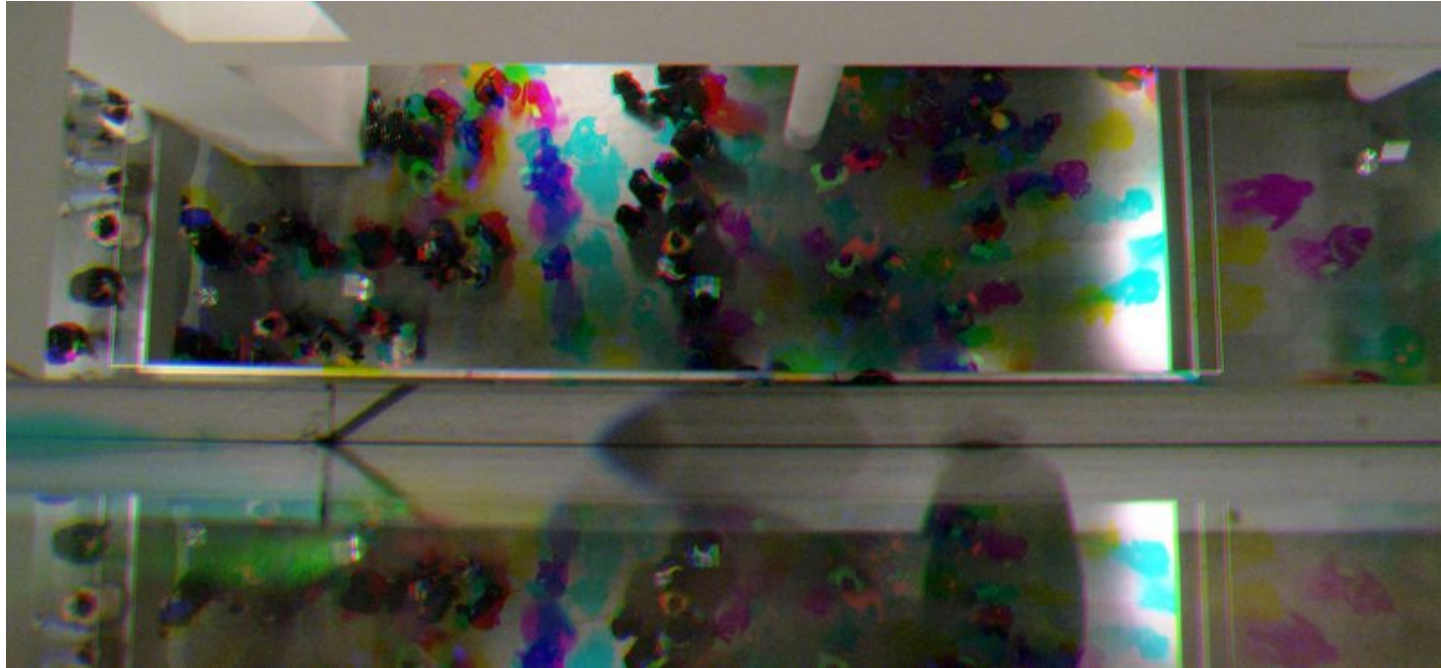


Reasoning

- There a small difference in **variability** (in terms of CNVs) between the network periphery and the center
- But, there is a (as shown before) marked difference in fixed (and hence, presumably, **selected**) SDs at the network periphery and center

Networks & Variation 2

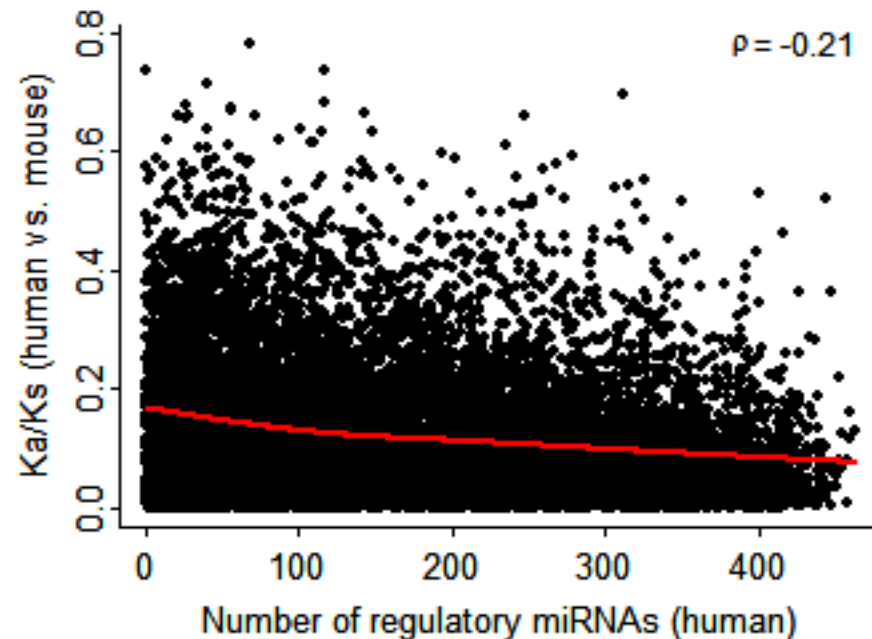
Variation in the miRNA network



Analyze Regulation in microRNA-target Network

- Relationship between target in degree (number of micro-RNAs that regulate gene) & evolutionary rate of gene?
 - ◊ In deg. related 3' UTR size
- Expectation: more regulation, more constraint

Relationship between microRNA regulation and protein evolution



Important genes are regulated more intensively regulated by the microRNAs

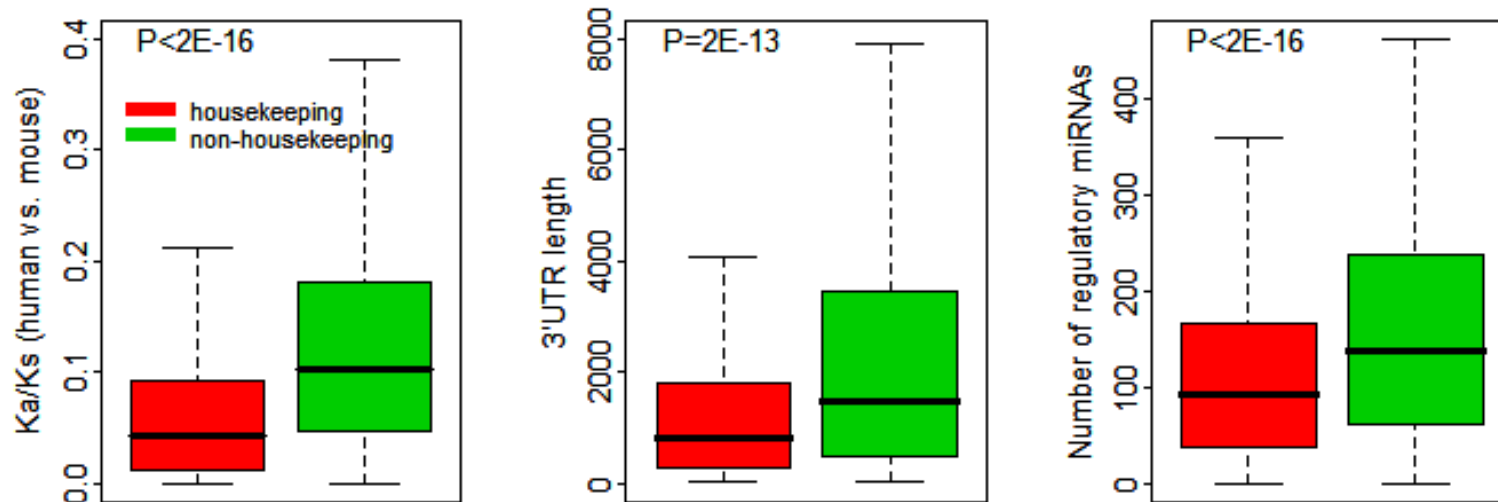
Human vs.	Number of genes	Correlation	P-value
chimpanzee	11326	-0.11	2.E-32
mouse	13280	-0.21	7.E-128
rat	12270	-0.20	4.E-107
cow	11683	-0.21	8.E-115
chicken	8061	-0.18	1.E-57

[Cheng et al., BMC Genomics, 2009 (in press)]

MicroRNA regulation: a two-way strategy

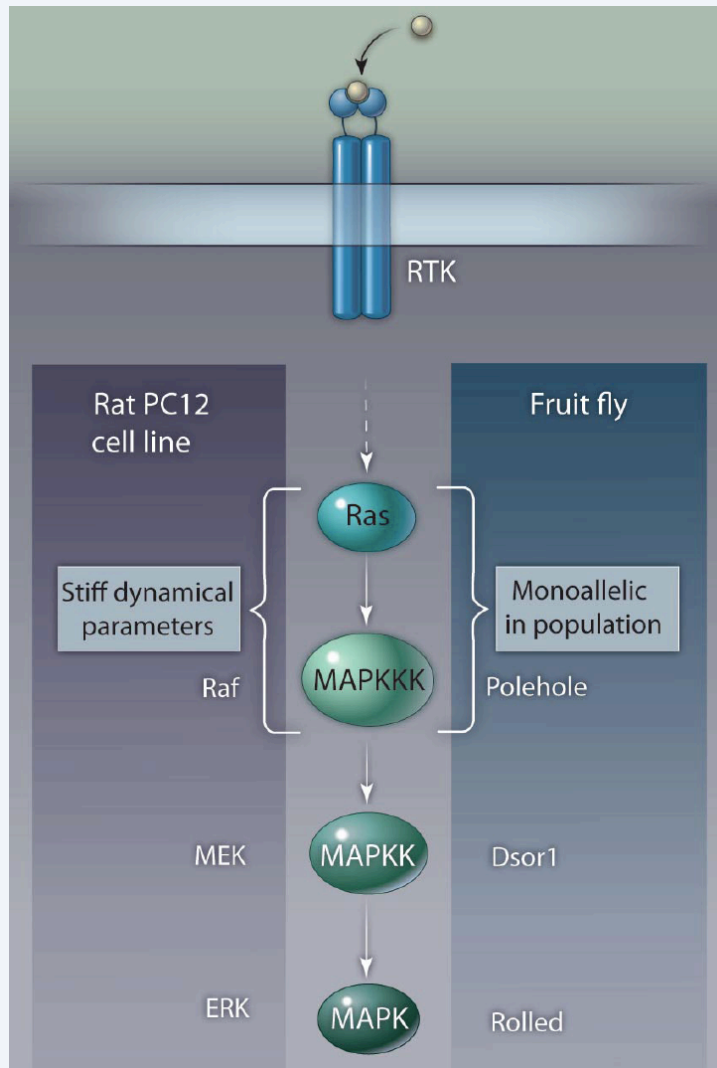
For non-housekeeping genes, functionally critical genes are intensively regulated by miRNAs and prefer long 3'UTR.

housekeeping genes, however conserved, are selected to have shorter 3'UTRs to avoid miRNA regulation.



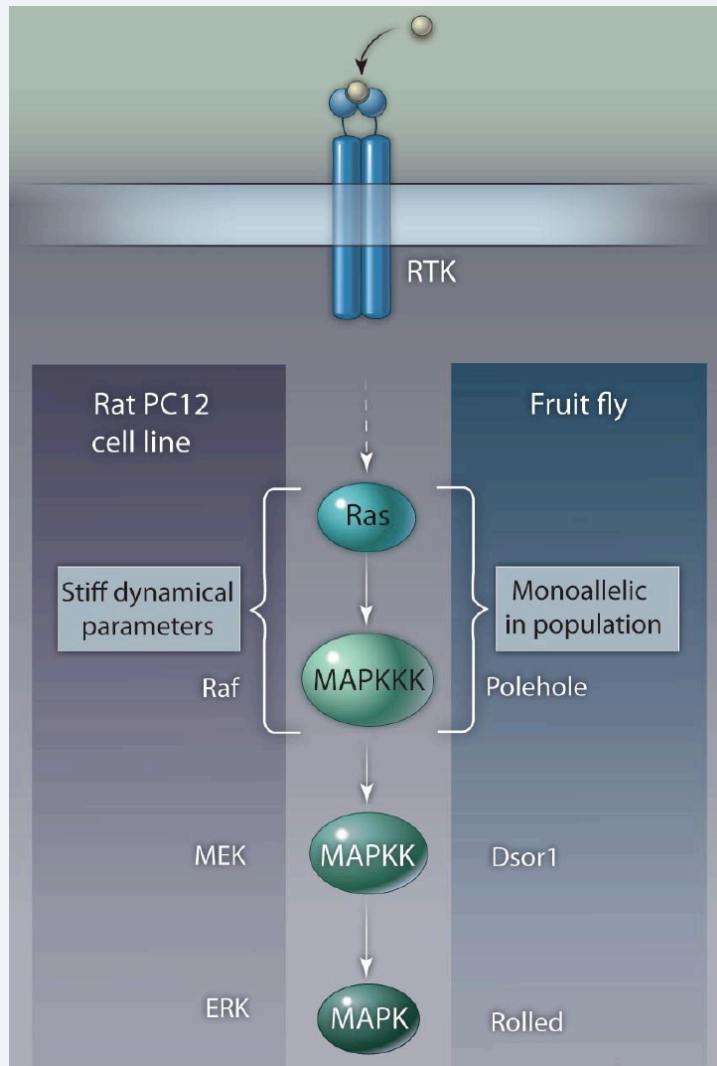
[Cheng et al., BMC Genomics, 2009 (in press)]

Network dynamics constrain evolution



Hypothesis: Nodes in a molecular network with the strongest impact on dynamic behavior should be under strong purifying selection and thus exhibit the least genetic variation.

Network dynamics constrain evolution

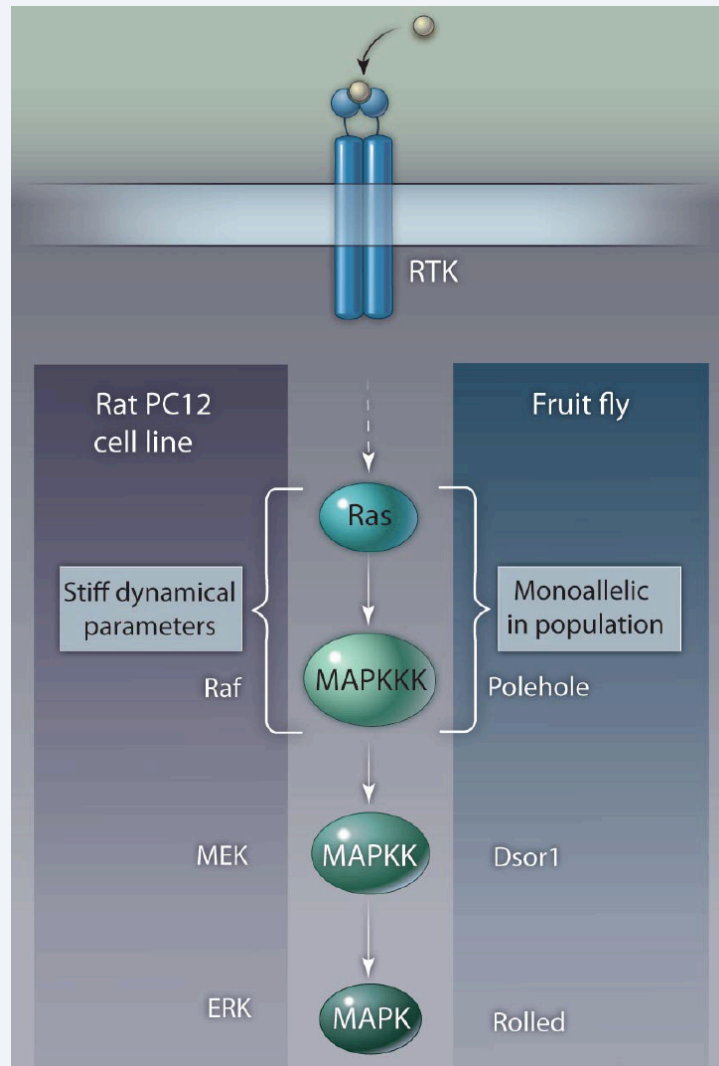


Hypothesis: Nodes in a molecular network with the strongest impact on dynamic behavior should be under strong purifying selection and thus exhibit the least genetic variation.

Algorithm:

- 1) Reconstruct families of molecular networks from genomic data.
- 2) Map some kind of genetic variation onto the networks.
- 3) Analyze sensitivity of dynamical model of the generic network.

Speculation: Why more tightly regulated gene might have less variation



Example: MAP Kinase signaling pathway

Dynamic model:

- ODE model with Michaelis-Menten kinetics
- parameters fit to time series data of protein activities in response to EGF and NGF from rat PC12 cell line

In sensitivity analysis, stiff parameters cluster around Ras and Raf.

Population study in fruit flies:

- allele variation based on PCR of pathway genes

Ras and Raf have less allele variation than other proteins in the network.

Analogies show it reasonable for more variable part of network to be periphery

- **Computer Networks**

- Servers in center have much depending on them; thus, can't be frequently updated & patched
- Servers on periphery often attacked and so need frequent patches

- **Social Networks**

- Individuals at center under more constraint (to conform), whereas those at periphery have more freedom to experiment

Outline: Molecular Networks

- Why Networks?
- Predicting Networks (yeast ppi)
 - ◇ Propagating known information
- Central Points in Networks
 - ◇ Hubs & Bottlenecks (yeast ppi & reg. net)
 - ◇ Tops of Hierarchies (yeast reg. net)
 - ◇ Identified by score (human miRNA-targ. net)
- Dynamics of Networks
 - ◇ Across environments (in prokaryote metab. pathways)
- Protein Networks & Variation (human ppi & miRNA-targ. net)



Conclusions on Networks: Predictions



- Predicting Networks
 - ◇ Extrapolating from the Training Set
 - ◇ Principled ways of using known information in the fullest possible fashion
 - Prediction Propagation
 - Multi-level learning

Conclusions:

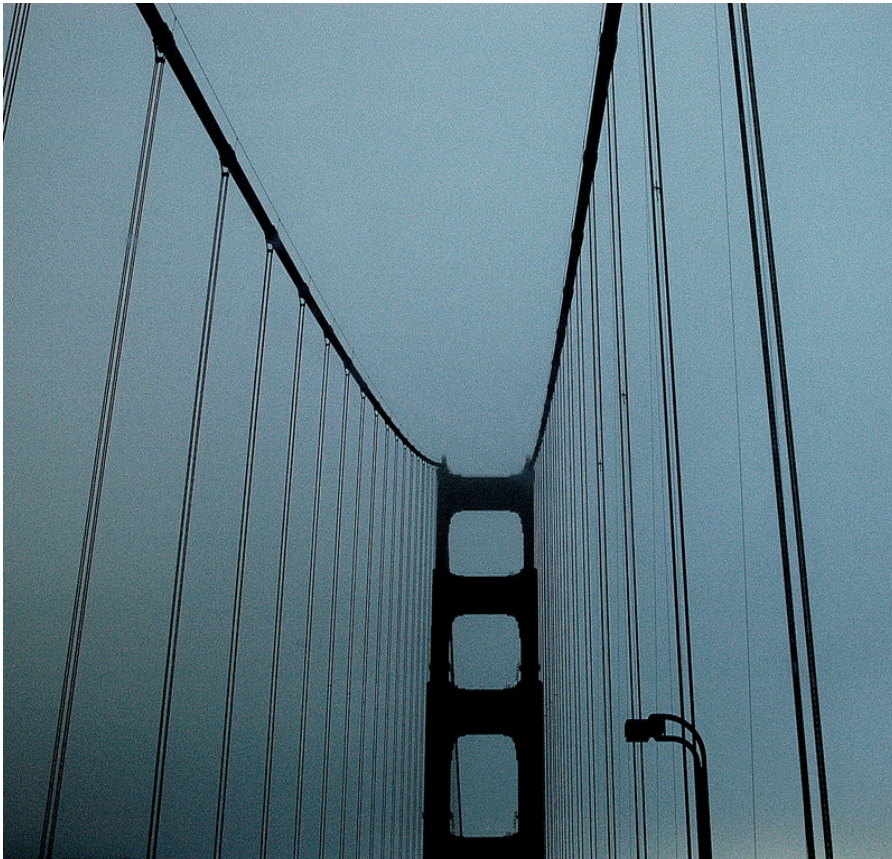
Analysis of Network Structure



- Centrality Measures in Protein Network
 - ◇ Hubs & Bottlenecks
 - ◇ Importance of later in regulatory networks
- Regulatory Network Hierarchies
 - ◇ Middle managers dominate, sitting at info. flow bottlenecks
 - ◇ Paradox of influence and essentiality
 - ◇ Topmost proteins sit at center of interaction network

Conclusions:

Points of Network Centrality



- RE-score measures degree of (down) regulation of targets vs. non-targets
- Application to miRNA network
- Different RE-score of miRNAs can be used in cancer classification

Conclusions: Networks Dynamics across Environments



- Developed and adapted techniques to connect quantitative features of environment to metabolism.
- Applied to available aquatic datasets, we identified footprints that were predictive of their environment (potentially could be used as biosensor).
- Strong correlation exists between a community's energy conversion strategies and its environmental parameters (e.g. temperature and chlorophyll).
- Suggest that limiting amounts of cofactor can (partially) explain increased import of amino acids in nutrient-limited conditions.

Conclusions: Connecting Networks & Variation



- We find ongoing evolution (positive selection) at the network periphery.
 - ◇ This trend is present on two levels:
 - On a sequence level, it can be seen as positive selection of peripheral nodes
 - On a structural level, it can be seen as the pattern of SDs that display significantly higher allele frequencies in non-central genes
 - ◇ 2 possible mechanisms for this : adaptive evolution at cellular periphery & relaxation of structural constraints at the network periphery
 - We show that the latter can only explain part of the increased variability

Conclusions: Connecting Networks & Variation 2



- More highly regulated genes are under more constraint in miRNA-target networks
- Exception for housekeeping genes
- Speculation as to why variation at periphery is quite reasonable



- an automated web tool

tYNA

(vers. 2 :

"TopNet-like

Yale Network Analyzer")

tYNA

Getting started API WSDL Download tYNA Installation guide Plugins for Cytoscape Contact Known problems

You are logged in as kevin. [Logout](#) View: Simple Advanced

List Owned Biological networks with (Attribute name) = (Attribute value) List

Workspace manager

Load an existing network

Load: 14. Uetz 2000 yeast two ...

Into: workspace 0

Categorized by: Nil

Load

Current working networks in your workspaces:

Workspace 0: statFilter(degrees, geq, 1, value, neighbors=false, intersection("Uetz 2000 yeast two hybrid", "Ito 2001 yeast two hybrid"))

Workspace 1: (empty)

Workspace 2: (empty)

Workspace 3: (empty)

Multiple network analysis

Networks in database (upload download)

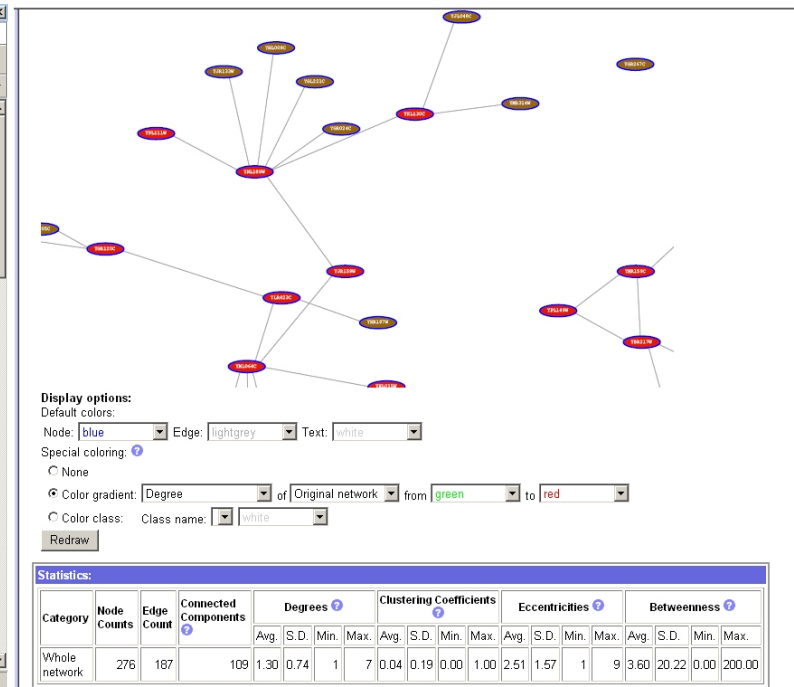
ID	Name	Creator	Creation date	
14	Uetz 2000 yeast two hybrid	kevin	21-Feb-06	Delete
15	Ito 2001 yeast two hybrid	kevin	21-Feb-06	Delete
16	Ho 2002 pull down	kevin	21-Feb-06	Delete
17	Gavin 2002 pull down	kevin	21-Feb-06	Delete
18	Jansen 2003 PIT	kevin	21-Feb-06	Delete
19	MIPS yeast PPI	kevin	21-Feb-06	Delete
21	BIND yeast data	kevin	21-Feb-06	Delete
22	DIP yeast data	kevin	21-Feb-06	Delete
23	Kim 2006 structural interaction	kevin	21-Feb-06	Delete
24	Han 2004 FYI data	kevin	21-Feb-06	Delete
25	Luscombe 2004 regulatory	kevin	21-Feb-06	Delete

Categories in database (upload download)

ID	Name	Creator	Creation date
----	------	---------	---------------

Statistics:

Category	Node Counts	Edge Count	Connected Components	Degrees				Clustering Coefficients				Eccentricities				Betweenness			
				Avg.	S.D.	Min.	Max.	Avg.	S.D.	Min.	Max.	Avg.	S.D.	Min.	Max.	Avg.	S.D.	Min.	Max.
Whole network	276	187	109	1.30	0.74	1	7	0.04	0.19	0.00	1.00	2.51	1.57	1	9	3.60	20.22	0.00	200.00



Normal website + Downloaded code (JAVA)
+ Web service (SOAP) with Cytoscape plugin

[Yu et al., NAR (2004); Yip et al. Bioinfo. (2006);
Similar tools include Cytoscape.org, Idekar, Sander et al]

H Yu
P Kim
K Yip
T Gianoulis
C Cheng

A Paccanaro
P Alves
T Emonet
P Cayting
M Seringhaus
Y Xia
J Korbel
A Sboner
P Patel
P Bork
J Raes
E Franzosa
M Snyder
N Bhardwaj
R Alexander

Acknowledgements



Networks.GersteinLab.org

Job opportunities currently for postdocs & students

More Information on this Talk

TITLE: Understanding Protein Function on a Genome-scale through the Analysis of Molecular Networks

SUBJECT: Networks

DESCRIPTION:

Network Biology: Understanding metabolic and protein interactions, Mathematical Biosciences Institute, Columbus, OH; 2009.09.14, 13:30-14:30; [I:**MBINETS**] (Long networks talk, adding in for the first time: **rescore***, **mirnatargevolrate*** & **netdynamicsrev***. Fits easily into 55' w. 5' questions. PPT works on mac & PC and has many photos.)

(Paper references in the talk were mostly from Papers.GersteinLab.org. The above topic list can be easily cross-referenced against this website. Each topic abbrev. which is starred is actually a papers "ID" on the site. For instance, the topic **pubnet*** can be looked up at <http://papers.gersteinlab.org/papers/pubnet>)

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